Original Article



Development of a Cost Effective Medium for Enhanced Production of *Bacillus* thuringiensis δ-endotoxin

Anamika Bhowmik, Muntahi Mourin, Md. Asaduzzaman Shishir, Shakila Nargis Khan and Md. Mozammel Hoq*

Department of Microbiology, University of Dhaka, Dhaka-1000, Bangladesh.

The present study was carried out to develop a sustainable production medium using locally available cheap raw materials for biopesticide production by *Bacillus thuringiensis* subsp. kurstaki (Btk) HD-73. In submerged fermentation (SmF) condition, the conventional Luria-Bertani (LB) medium which was enriched with nitrogen source (10% defatted soybean meal) supported 28.57% sporulation and 125% endotoxin increase over LB (alone). The effect of cystine on sporulation and endotoxin synthesis was highly pronounced in LB-soybean medium (LBS) with a range of 19.54% and 131.35% higher endotoxin yield respectively in SmF condition. While basal salts supplemented in soybean-cystine (SMc) medium, it resulted in 7.65% endotoxin yield compared to LB-soybean-cystine (LBSc) medium. Addition of molasses balanced the C: N ratio in the SMc medium thus helping 84.85% higher endotoxin synthesis after 24 hours fermentation. Substitution of basal salts with cost effective sea water yielded about 19.27% less endotoxin. The optimum medium thus obtained consisting of soybean extract-molasses-cystine with sea water was used in 3.0 L bioreactor cultivation for endotoxin synthesis by Btk HD-73 under 30% saturation of dO_2 through cascade of agitation and aeration. The production rate obtained was 1.67 fold higher in bioreactor than in shake flask culture.

Keywords: Bacillus thuringiensis subsp. kurstaki HD-73, molasses, delta-endotoxin, Bt endotoxin production

Introduction

Bioinsecticide preparations based on the spores and insecticidal crystal proteins (ICPs) of Bacillus thuringiensis (Bt), were proved to be eco-friendly agents for controlling many pests in agriculture and disease causing vectors. Bt is a naturally occurring entomopathogenic bacterium, commonly found in soil throughout the world but the use of Bt based biopesticides is limited in developing countries for their unavailability and cost. The high cost of Bt biopesticides is due to the production being located mostly in developed countries incurring higher production costs and the shipment cost to the operational sites as well. The expense for raw materials is one of the principal costs involved in overall Bt production. In conventional Bt production process, the expense of raw materials varied between 30% and 40% of the total costs depending on the plant production capacity¹. Therefore, the production of Bt biopesticide based on locally available cheap raw materials including agro-industrial by products in developing countries like Bangladesh will reduce the cost significantly^{2,3}.

Bt biopesticides are usually composed of spores and crystals protein mixtures, harvested from the production media, readily produced by aerated liquid fermentation. They are easily harvested and have a long shelf life when formulated properly⁴. Optimizing different culture conditions and regulating some critical factors, it is possible to obtain higher yield in terms of cell mass, Cry protein concentration and toxicity to develop efficient

Bt formulations^{3,5}. Critical factors e.g. sugars have significant impact on cell growth but when used at high concentrations, they can cause adverse effects on sporulation due to the acids produced by Bt ä-endotoxin from carbohydrates⁵ and the balance of the ratio between carbon and nitrogen itself is important for the crystal protein production⁶.

Other important components for the production of crystal proteins are the trace minerals⁷. Again, amino acids are important in the formation of spores and crystal proteins⁸. Moreover, it was reported that decreasing the proteolytic activity in the fermentation medium increased the \ddot{a} - endotoxin titer which is susceptible to the proteolytic degradation^{3,9}. So, the optimization of nutrients and other factors could help in attaining Bt formulations with high spore count and high toxin titer.

In Bangladesh, agro-industrial by-products like defatted soybean meal, defatted mustard seed meal, molasses, rice husk, rice bran, citrus peels etc are generated in huge amount every year ¹⁰. These can be utilized in formulating cost effective media to promote biopesticide production and its application here³. The goal of this work was, therefore, to develop a low-cost bio-insecticides production medium based on the defatted soybean meal, one of the most common and cheap by products of edible oil industry, as raw material and to determine other influential factors such as cystine, mineral salts in sporulation and ä-endotoxin synthesis by *Bacillus thuringiensis*.

Materials and Methods

Bacterial strain and inoculum preparation

The reference *Bt kurstaki* strain HD-73 was obtained from Okayama University, Japan and preserved in 15% glycerol stock at -70°C. Inoculum was prepared by inoculating a single *Bt* colony into a 250 ml Erlenmeyer flask containing 50 ml of LB (Luria-Bertani) broth (per litre: yeast extract, 5.0 g; tryptone, 10.0 g; NaCl, 10.0 g)¹¹ and incubated overnight at 30°C and 180 rpm which was used in all media assessment studies if not otherwise stated. Each time, inoculum was aseptically added into the medium by micropipette so that the process starts with an OD_{600nm} = 0.1.

Media used and the culture conditions

Medium 1 (LB): Luria bertani (LB) broth was used for inoculum prepration and as standard medium for Bt biopesticide production. The pH of the medium was adjusted to 7.0 ± 0.2 .

Medium 2 (LBS): The LB broth was supplemented with 10% of finely ground defatted soybean meal and the inoculum was added in it to assess the efficiency of soybean. The pH was adjusted to 7.0 ± 0.2 before autoclaving.

Medium 3 (LBSc): The LBS broth was supplemented with 300 il of 10% cystine. Stock solution of 10% cystine was prepared by suspending and autoclaving 6.0 g cystine in 54.0 ml of phosphate buffer.

Medium 4 (SMc): 10 g of defatted Soybean meal was added into 90 ml of basal salt solution [(g/L): MgSO $_4$.7H $_2$ O, 0.5; MnSO $_4$.H $_2$ O, 0.1; FeSO $_4$.7H $_2$ O, 0.02; ZnSO $_4$.7H $_2$ O, 0.02; CaCl $_2$, 0.01 & KH $_2$ PO $_4$ 1.0] 12 in a 500 ml conical flask. It was then supplemented with 300 il of 10% cystine.

Medium 5 (SMmc): 0.5 g of molasses was added into the 100 ml of previously described SMc medium. The pH was adjusted to 7.0 ± 0.2 before autoclaving.

Medium 6 (SSwMmc): Sea water was used in SMmc medium as a substituent of basal salts. In this case everything was same as SMmc medium except 20 ml of sea water instead of 90 ml of basal salts and the volume was adjusted with distilled water up to 100 ml.

Medium 7 (SeSwMmc): In this medium soybean extract was use instead of soybean mass. 100 ml of 10% soybean suspension was boiled for 10 min and the aqueous part was separated from the solid mass. This suspension was used in SwMmc medium instead of 10.0 g of soybean.

Estimation of spore count

The spore counting was performed each time in triplicate from the 1.0 ml sample collected during the culture. The sample was heat treated at 80°C for 15 min, serially diluted and plated on the LB agar medium. The plates were then incubated at 37°C for 24 hrs. The colonies were then counted and multiplied by the dilution factor to estimate their number.

Partial purification and determination of ä-endotoxin concentration

The purification of crystal protein was done by the modified method ^{13, 14} where 1 ml culture was washed twice with sterile distilled water by centrifugation at 10,000 rpm for 10 min. The pellet was treated with 1.0 M NaCl and 5.0 mM EDTA and later with 5.0 mM EDTA alone. Finally the pellet was resuspended in 1.0 ml 0.1 N NaOH solution and incubated for 1 hr at room temperature to solubilize the crystal proteins. Thus the partially purified crystal protein concentration in the supernatant was estimated by Bradford method¹⁵.

Bioinsecticide production in 3.0 L bioreactor

Production experiments were carried out at 30 °C in a 3.0 L fully controlled bioreactor (New Brunswick Scientific, USA) containing 1.5 L of the finally optimized medium. 30% dissolved oxygen (DO) level in the medium was automatically controlled by the aeration, set at 1.0 SLPM (Standard liter per minute) and agitation was set at 250 rpm. Dissolved oxygen was continuously monitored by an oxygen sensor but the pH was not controlled.

Results

Effect of defatted soybean meal with LB on sporulation and äendotoxin formation

The defatted soybean mass is a readily available rich nitrogen source for bacterial growth hence could be cost effective for large scale production of the *Bt* biopesticides as a substituent of yeast extract and tryptone in the LB medium. The performance of the defatted soybean meal was, therefore, examined supplementing in the LB medium on sporulation and ä-endotoxin synthesis by *Btk* HD-73 against LB medium as control. Maximum sporulation and ä-endotoxin concentration were obtained as 7.447 log CFU/ml and 0.371 mg/ml respectively after 48 hours under comparable cultivation conditions in LBS medium whereas LB medium (as control) produced its maximum after 72 hours with relatively lower yield (7.217 log CFU/ml and 0.149 mg/ml respectively). Based on the productivity index, the defatted soybean mass caused 2.5 fold higher yield in ä- endotoxin production than that of the LB alone (Figure 1).

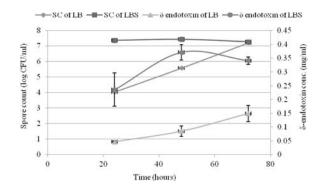


Figure 1. Evaluation of soybean meal & LB medium on sporulation & endotoxin synthesis by Btk HD 73.

Role of cystine in defatted soybean meal with LB

Since, cystine promotes the growth and sporulation of \hat{A} . thuringiensis 16 , cultivation of the Btk HD-73 was carried out in the medium, LBS under comparable conditions with or without cystine (300µg/ml) for 72 hours. Maximum sporulation and ä-endotoxin concentration were obtained after 48 hours in both cases. A significant change was observed on the ä- endotoxin concentration when cystine was added. The sporulation and ä-endotoxin were increased as 19.54% (7.447 log CFU/ml to 9.06 log CFU/ml) and 131.35% (0.371 mg/ml to 1.791 mg/ml) respectively over the LBS medium without cystine (Figure 2).

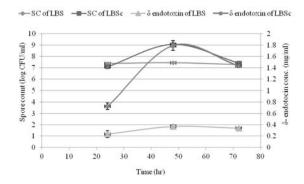


Figure 2. Role of cystine in defatted soybean meal.

Replacement of LB with basal salts in soybean-cystine medium

As, soybean was found to serve as a good C and N source, basal salt solution was added in the medium removing tryptone and yeast extract in this experiment. Maximum sporulation and ä-endotoxin concentration were obtained at 48 hours which were 9.352 log CFU/ml and 1.659 mg/ml respectively with this formulation on defatted soybean meal with cystine and basal salts (SMc) (Figure 3). Though the endotoxin was 7.65% less than that of LBSc, it was still comparable and acceptable because of its cost effectiveness.

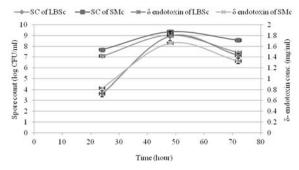


Figure 3. Effect of basal salt solution in soybean-cystine medium.

Improvement of yield with molasses

The yield in sporulation and ä-endotoxin synthesis was improved by adding molasses in SMc medium. This promoted higher endotoxin formation (2.376 mg/ml versus 1.659 CFU/ml) after 48

hours under comparable conditions over the same medium composition without molasses (Figure 4).

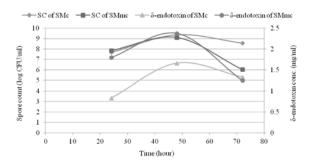


Figure 4. Effect of molasses supplementation in Soybean-cystine medium.

Comparison between basal salts and sea water as mineral source

In order to reduce the further cost of bioinsecticide production, sea water was used as the possible supply of the minerals instead of basal salt solutions in SSwMmc medium. In this experiment, maximum sporulation and ä- endotoxin production were 8.176 log CFU/ ml and 1.918 mg/ml after 48 hours as compared to 9.114 log CFU/ ml and 2.376 mg/ ml in the media, SMmc with basal salts (Figure 5). Though the medium containing basal salt solutions gave higher ä-endotoxin yield in comparison to medium containing sea water, this slight difference can be compensated due to its cost effective production.

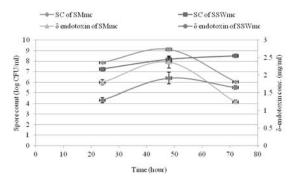


Figure 5. Replacement of basal salts with sea water in soybean-molasses-cystine medium

Efficacy of soybean extract as substituent of soybean mass

For large scale production there are few drawbacks for the medium with soybean mass. For this purpose a defatted soybean extract-molasses- cystine medium was designed for trial. The sporulation and ä- endotoxin concentration was 8.596 log CFU/ml and 1.386 mg/ml at 24 hours. But ä- endotoxin concentration decreased from 1.359 mg/ml to 0.471 after 72 hours (Figure 6).

Production kinetics of *Btk* HD-73 under controlled conditions in a 3.0 L bioreactor

From the results, it was observed that the maximum ä-endotoxin yield was 2.3 mg/ml after 24 hours as compared to that after 48 hrs

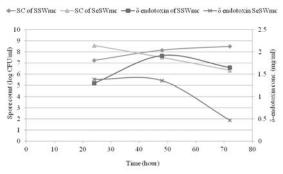


Figure 6. Effect of soybean extract in place of soybean meal in the soybean-molasses-cystine medium

in shake flask culture (Figure 7). An increase in 1.67 fold of ä-endotoxin production was obtained in bioreactor than in shake culture. The spore counts or growth of *Bt* was highly correlated kinetically with endotoxin formation as a growth associated manner.

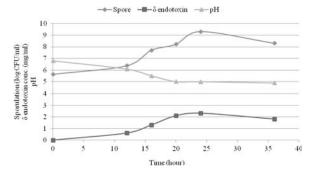


Figure 7. *Total spore count and ä-endotoxin concentration in 3L bioreactor.*

Discussion

For large scale production of Bt biopesticide, several different approaches were employed to design media supporting higher sporulation and \ddot{a} - endotoxin synthesis. As sporulation and endotoxin production occur simultaneously, it is very important to optimize the factors including nutrients and culture conditions to attain maximum biomass yield which might result in maximum sporulation and high \ddot{a} - endotoxin titer. Carbon and nitrogen sources and their critical concentrations not only affect the growth rate of microbes in general but also the sporulation and the rate of endotoxin production. This aspect is rather more important with Bt as the endotoxin (Cry protein) synthesis is critically related to sporulation rate.

Carbon and nitrogen sources and their critical concentrations not only affect the growth rate of microbes in general but also the sporulation and the rate of endotoxin production. This aspect is rather more important with Bt as the endotoxin (cry protein) synthesis is critically related with sporulation rate^{17, 18, 19}. So, proper optimization of nutritional and other factors are essential for large scale production of Bt biopesticide with a selective Bt strain. Luria-Bertani (LB) is one of the balanced medium with

respect to C, N, NaCl for bacterial culture and propagation. However, the components, partially of the defined medium, are usually suitable for lab scale experiments. However, this medium was not designed with the aim of enhanced sporulation and äendotoxin synthesis rather for routine culture, propagation and culture maintenance. Hence, this medium could be used to compare the efficiency of the experimentally designed media in spore and endotoxin production and thus selected as the model one to develop a cost effective medium with locally available raw materials such as soybean meal and molasses. In this connection, experiments were designed to replace yeast extract and tryptone (as N source) of LB composition with defatted soybean meal (or extract) and molasses respectively on the growth, sporulation and endotoxin formation by a reference strain Btk HD-73. The effect of cystine in the growth and sporulation enhancement as reported in many articles was evaluated in this study. And the feasibility of sea water instead of basal salts as the source of trace elements required for microbial growth was also assessed.

Initial experiment, performed to evaluate the compatibility of the soybean meal as nitrogen source in comparison to LB broth on the formation of spores and ä- endotoxin by *Btk* HD-73 revealed that the yield increased at 58.35% and 133.85% respectively after 24 hours. The soybean meal in the medium also supported faster growth to attain the maximum spore count and ä- endotoxin synthesis unlike LB medium that took 72 hrs. The increase in sporulation and ä-endotoxin yield however may be due to the complex rich nitrogen source with a composition of Soybean meal: 91.52% dry matter, 43.30% crude protein²⁰.

Certain amino acids help in growth, sporulation and ä-endotoxin formation as the effect of cystine on the growth, sporulation and endotoxin production by Bt subsp kurstaki reported by Vora and Shethna ²¹. In the present study, the effect of 300 mg/ L cystine was observed in cystine supplemented LB-Soybean medium (LBSc) for 72 hours. It could be clearly observed from Figure 2 that maximum sporulation and endotoxin yield was obtained after 48 hours which was 19.54% and 131.35% higher than that of the LBS medium. This increase may be due to the fact that cystine might have interfered with some of the macromolecular changes during sporulation and parasporal crystal formation²². Lower cystine concentration facilitates sporulation related specific events such as dipicolinic acid synthesis by Bt whereas higher concentration inhibits this phenomena. They have showed that maximum sporulation and ä- endotoxin concentration were obtained at 300 mg/L of cystine on Cystine Basal Media (CBM) and it was 1523 ig/ml.

Basal salts have very specific role in the metabolism and growth of the bacteria. Hence the basal salts were used instead of the LB broth in the defatted soybean meal- cystine medium. In this medium, the endotoxin formation was 1.791 mg/ml for LBSc and 1.659 mg/ml for SMc. The endotoxin concentration was 7.65% less on SMc medium than that of LB-soybean-cystine medium (LBSc). However, the lower yield might not be due to the basal

salts but due to the lack of balanced carbon sources with the SMc medium.

Since nitrogen sources (yeast extract and tryptone) of LB medium has been substituted with defatted soybean meal, to balance carbon in the SMc medium, molasses was used to observe the effect on sporulation and ä- endotoxin synthesis by *Btk* HD-73 (Figure 4). It has been seen that after 48 hours the spore count and ä- endotoxin concentration both were increased in the SMc medium as compared to the same without molasses. 84.85% higher endotoxin yield may be due to balances of C: N ratio and growth factor present in the molasses. Molasses also aided the earlier growth of the organism in 48 hrs as compare to 72 hrs.

Sea water contains (g/L): Na⁺ 12; Cl⁻22; K⁺0.4; Ca⁺0.14; HCO³-0.40; Mg⁺ 1.3; Fe²⁺ SO₄²⁻ 2.640 ⁴. In order to reduce the cost of bioinsecticide production, sea water was used to replace the basal salts (Figure 5) required for ä- endotoxin production in the culture medium of *Bt*. The ä- endotoxin formation was 2.376 mg/ml and 1.918 mg/ml at 48 hrs in the medium with basal salts (SMmc) and in medium with sea water respectively. Though the medium containing basal salt solutions gave 21% higher ä-endotoxin yield in comparison to medium containing sea water, it is more cost effective and suitable for large scale production of biopesticide. These results seem to be encouraging to examine further studies with respect to the composition of sea water based on sources and seasonal variations.

The ä- endotoxin concentration decreased from 1.918 mg/ml to 1.649 mg/ml and from 2.376 mg/ ml to 1.246 mg/ ml in SSWmc and SMmc media respectively after 48 hrs may be due to the degradation by proteases synthesis by *Bacillus* species during growth and sporulation as has been reported^{3, 23}.

Foaming, a process hampering problem in bioreactor, was not any exception in this study. Soybean extract is advantageous as compared to soybean mass because the former eases the difficulties by allowing proper mixing and dO₂ availability as well as less foaming in the bioreactor. The SeSWmc medium showed 8.596 log CFU/ml sporulation and 1.386 mg/ml endotoxin concentration after 24 hours whereas SSWmc showed 7.249 log CFU/ml and 1.296 mg/ml endotoxin concentration after 24 hours. This concentration remained quite same after 48 hours in SeSWmc medium but was increased in SSWmc medium up to 34.11% (Figure 6). The soybean extract medium caused highest ä- endotoxin production within 24 hours of fermentation. Thus it could be suitable for large scale production based on the fact that it would reduce the fermentation time at the same time reduce the production cost.

The optimized medium thus was used for production of biopesticide by *Btk* HD-73 in a 3.0 L bioreactor under controlled conditions. Temperature was controlled at 30ÚC and pH was kept uncontrolled. Initially, the pH was 6.8 and it became acidic later and after 36 hours the pH dropped to 4.98 (Figure 7). This might be due to acid production during sugar fermentation. For

Btk HD-73, ä- endotoxin concentration increased after 16 hours as vegetative cells entered into sporulation phase and maximum ä- endotoxin yield was 2317 mg/L at 24 hours. There was an increase of 1.67 fold in ä- endotoxin concentration in the bioreactor then that of shake flask culture. Thus the results of present bioprocess conditions by Btk HD-73 on locally available cheap raw materials can successfully be applied for pilot plant or commercial level production by potential indigenous Bt strains²⁴, with novel toxicity against different pest species especially $Bactrocera\ cucurbitae\ ^{25}$.

Acknowledgement

This work was supported by a Grant-in-Aid from the USDA as a project entitled "Production of *Bacillus thuringiensis* biopesticides by biotechnological approach for the control of vegetable pests in Bangladesh." We thank Okayama University, Japan for providing reference strain.

References

- Ejiofor AO. 1991. Production of *Bacillus thuringiensis* serotype H-14 as bioinsecticide using a mixture of 'spent' brewer's yeast and waste cassava starch as the fermentation medium. *Discov. Innovat.* 3: 85-88.
- Hasan MH, Akter A, Ilias M, Khan SN and Hoq MM. 2011. Growth, sporulation and toxin production by *Bacillus thuringiensis* isolates in media based on mustard-seed meal. *Bangladesh J. Microbiol.* 27(2): 51-55.
- Mourin M, Shishir MA, Khan SN and Hoq MM. 2015. Regulation of major cultural components for designing a cost effective medium to increase endotoxin synthesis by *Bacillus thuringiensis*. *African J. Biotech.* 14(16): 1379-1386.
- Ghribi D, Zouari N, Trigui W and Jaoua S. 2007. Use of sea water as salts source in starch- and soya bean-based media, for the production of *Bacillus thuringiensis* bioinsecticides. *Process Biochem.* 42:374-378.
- Dulmage HT, Yousten AA and Singer SLLA. 1990. Guidelines for production of *Bacillus thuringiensis* H-14 and *Bacillus sphaericus*. UNDP WORLD BANK WHO Special Programme for Research and Training in Tropical Diseases. p.42.
- Farrera RR, Perez-Guevara F and De la Torre M. 1998. Carbon: nitrogen ratio interacts with initial concentration of total solids on insecticidal crystal protein and spore production in *Bacillus* thuringiensis HD-73. Appl. Microbiol. Biotechnol. 49: 758-765.
- Rose AH. 1979. Production and industrial importance of secondary products of metabolism. In: Economic Microbiology, Academic Press, London, vol.3, pp.2-34.
- Sachidanandham R, Jenny K, Fiechter A, Jayaraman K. 1997. Stabilization and increased production of insecticidal crystal proteins of *Bacillus thuringiensis* subsp. *galeriae* in steady- and transientstate continuous cultures. *Appl. Microbiol. Biotechnol.* 47: 12-17.
- Ennouri HBH, Saoussan BK and Zouari NH. 2013. Concomitant production of delta-endotoxins and proteases of *Bacillus thuringiensis* subsp. *kurstaki* in a low-cost medium: effect of medium components. *Acta Biol. Szegediensis.* 57(1):13-19.
- Sheela SH, Ahmed MF and Gomes DJ. 2008. Fuel Ethanol Production from Molasses by Some Indigenous Yeast Isolates. *Bangladesh J. Microbiol.* 25(2): 129-133.
- Sambrook J and Russell DW. 2001. Molecular Cloning, a Laboratory Manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.

- Gangurde RP and Shethna YI. 1995. Growth, sporulation and toxin production by *Bacillus thuringiensis* subsp. *israelensis* and *B. sphaericus* in media based on mustard-seed meal. *World J. Microbiol. Biotechnol.* 11(2): 202-205.
- Liu WM, Bajpai MR and Bihari V. 1994. High-density cultivation of sporeformers. Ann. N.Y. Acad. Sci. 721: 310-325.
- Öztürk F, Açýk L, Ayvaz A, Bozdoðan B and Suludere Z. 2009. Isolation and Characterization of Native *Bacillus thuringiensis* trains from Soil and Testing the Bioactivity of Isolates against *Ephestia Kuehniella* Zeller (Lepidoptera: Pyralidae) Larvae. *Turk. J. Biochem.* 33: 202-208.
- Bradford MM. 1976. A rapid and sensitive method for the quantization of microgram quantities of protein utilizing the principle of proteindye binding. *Analyt. Biochem.* 72: 248–254.
- Rajalakshmi S and Shethna YI. 1977. The effect of amino acids on growth, sporulation and crystal formation in *Bacillus thuringiensis* var. thuringiensis. J. Ind. Ins. Sci. 59(12): 169-176.
- Nickerson KW and Bulla LA (Jr.). 1975. Lipid metabolism during bacterial growth, sporulation and germination: an obligate nutritional requirement in *Bacillus thuringiensis* for compounds that stimulate fatty acid synthesis. *J. of Bacteriol.*, 123(2): 598-603.
- Rogoff MH and Yousten AA. 1969. Bacillus thuringiensis: Microbiological considerations. Ann. Rev. Microbiol. 23: 357-386.

- Singer S, Goodman NS and Rogoff MH. 1966. Defined media for the study of Bacilli pathogenic to insects. Ann. NY. Acad. Sci. 139(1): 16-23.
- Jahan DA, Hussain L, Islam MA and Khan M. 2013. Comparative Study of Mustard Oil Cake and Soybean Meal Based Artificial Diet for Rohu, Labeo rohita (Ham.) Fingerlings. The Agriculturists, 11(1): 61-66.
- Vora D and Shethna YI. 1999. Enhanced growth, sporulation and toxin production by *Bacillus thuringiensis* subsp. *kurstaki* in oil seed meal extract media containing cystine. *World J. Microbiol. Biotechnol.* 15: 747-749.
- Rajalakshmi S and Shethna YI. 1980. Effect of L-cystine on macromolecular changes during spore and parasporal crystal formation in *Bacillus thuringiensis* var. thuringiensis. J. Biosci., 2: 311–319.
- Doi RH. 1972. Role of proteases in sporulation. Curr. Top. Cell. Regul. 6: 1-20
- Shishir A, Roy A, Islam N, Rahman A, Khan SN and Hoq MM. 2014.
 Abundance and diversity of *Bacillus thuringiensis* in Bangladesh and their cry genes profile. Front. Environ. Sci. 2: 20.
- Shishir MA, Akter A, Bodiuzzaman M, Hossain MA, Alam MM, Khan SA, Khan SN and Hoq MM. 2015. Novel toxicity of *Bacillus thuringiensis* strains against melon fruit fly, *Bactrocera cucurbitae* (Diptera: Tephritidae). *Biocontrol Sci.* 20(2): 115-123.