

Molecular Characterization of Tropical Strawberry Genotypes

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Abstract

Genetic variability of tropical strawberry genotypes was determined through Random Amplified Polymorphic DNA (RAPD) primers. Out of 14 RAPD primers six showed reproducible and polymorphic bands. The results revealed that the maximum polymorphic bands were produced by the primer OPB 12. Ten genotypes were differentiated into two clusters on the basis of Unweighted Pair Group Method With Arithmetic Averages (UPGMA). Genotypic variation based on molecular characterization indicated that genotypes belonging to two different clusters depend on their genetic component. So, selection of parents from different clusters will provide the maximum heterosis in yield.

Introduction

Strawberries are important climacteric soft fruit crop widely grown in the world, adapted in geographically diverse area (Biswas et al. 2008). In Bangladesh there is no statistics about the area and production of this crop, since it has recently been introduced into the country. Growers' level extension of strawberry farming can bring a new horizon to the agriculture sector in Bangladesh. There has been a bright prospect of farming strawberry, a high-value crop, everywhere in the country except the coastal districts. Strawberry farming has started gain in popularity in northern region for the last couple of years. Many fresh initiatives have been taken to develop the trade in Panchagarh, Dinajpur, Tangail, Rangpur, Kurigram, Mymensingh, Noakhali, Laxmipur, Jessore, Magura, Faridpur, Madaripur and Dhaka district of Bangladesh (Anon. 2009).

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There are more than 20 *Fragaria* species and numerous cultivars commercially cultivated in several countries (Gaafar and Sarker 2006). A germplasm collection with good variability for the desirable characters is the basic requirement of any crop improvement program (Singhania et al. 2006). In addition, crop improvement is primarily based on extensive evaluation of germplasm. Molecular and morphological characters of a plant are most important for nature as well as yield of a crop. Hence, studies about these important traits are necessary for successful cultivation of this crop in a new area like Bangladesh. In Bangladesh, some varietal trials of strawberry (Hossan et al. 2013) have been conducted but the number of sustainable strawberry cultivars is absent for the country's climate. Molecular markers have been developed as a powerful tool to analyze genetic relationships and genetic diversity. As an extension to the variety of existing techniques using polymorphic DNA markers, the Random Amplified Polymorphic DNA (RAPD) technique may be used in to determine the genetic relationships of different crops (Hasan and Raihan 2015). So, the present study has been planned to find out the variations at molecular level of the newly introduced strawberry genotypes in Bangladesh.

Materials and Methods

The following ten genotypes were used for polymorphism study: V₁- FA 006, V₂- BARI strawberry 1, V₃- FA 015, V₄-FA 024, V₅- FA 016, V₆- FA 005, V₇- FA 007, V₈- FA 008, V₉- FA 023 and V₁₀- FA 017.

DNA extraction and RAPD amplification: DNA was isolated using the modified CTAB method and DNA samples were evaluated both quantitatively and qualitatively using spectrophotometer (SPECORD 50, Analytikjena, Germany) at 260 nm and agarose gel electrophoresis, respectively. Then, RAPD amplification reactions were performed in a total volume of 25 µl mixing with 2.5 µl *Taq* Buffer A10X (Tris with 15 mM MgCl₂), 0.5 µl dNTPs 2.5 mM, 0.2 µl *Taq* DNA polymerase 5U/µl, 1.0 µl primer, 19.8 µl sterile deionized distilled water and 1.0 µl DNA template. The results obtained with this primer were later confirmed by six other standard primers, OPA-02, OPA-03, OPB-05, OPB-06, OPB-11 and OPB 12 (Operon Technologies Inc., Alameda, CA, USA), which yielded satisfying results in the same conditions.

PCR amplification for RAPD: PCR mplifications were carried out in a Perkin-Elmer thermal cycler 480 (Perkin-Elmer, Milan, Italy). In each thermal cycling a negative control (water instead of template) was included to rule out amplification products due to external contamination. All amplifications were repeated twice for each sample on 1% agarose gel. The optimum amplification cycle was as follows:

45 times	}	Initial denaturation	94°C	For	5 min
		Denaturation at	94°C	For	1 min
		Annealing at	36°C	For	30 sec
		Extension at	72°C	For	3 min
		Final extension at	72°C	For	5 min

After completion of cycling programme, the reactions were held at 4°C.

Electrophoresis of the amplified products and documentation: The amplified products were separated electrophoretically on 1% agarose gel. The gel was prepared using 1.0 g agarose powder containing ethidium bromide and 100 ml 1 × TAE buffer. Agarose gel electrophoresis was conducted in 1 × TAE buffer at 50 Volts and 100 mA for 1.5 hrs. One molecular weight marker 1 kb DNA ladder was electrophoresed alongside the RAPD reactions. DNA bands were observed on UV-transilluminator and photographed by a gel documentation system.

RAPD data analysis: Following electrophoresis, the size of amplification products were estimated by comparing the migration of each amplified fragment with that of a known size fragments of 1 kb molecular weight marker. All distinct bands or fragments (RAPD marker) were there by given identification numbers according to their position on the gel and scored visually on the basis of their presence (1) or absence (0), separately for each individual varieties and each primer. The scores obtained using all primers in the RAPD analysis were then combined to create a single data matrix. Jaccards similarity and dissimilarity coefficients were then calculated between pairs of tracks and strains were grouped by using the Unweighted Pair Group Method With Arithmetic Averages (UPGMA), with a dendrogram construction utility software.

Results and Discussion

Despite of an important horticultural crop in the world, little information is available on genetic structure of strawberries. The results obtained from the experiment have been presented and discussed under the following headings.

Primer selection and RAPD pattern: Fourteen primers were initially screened on 10 strawberry genotypes for their ability to produce polymorphic patterns and six primers (OPA-02, OPA-03, OPB-05, OPB-06, OPB-11 and OPB-12) gave reproducible and distinct polymorphic amplified products.

A total of 38 RAPD bands were scored of which 38 (100%) polymorphic amplification products were obtained by using these arbitrary primers. The size of the amplification products ranged from 250 - 5000 bp (Table 1). The dissimilar

numbers of bands were generated by primer OPA-02, OPA-03, OPB-05, OPB-06, OPB-11 and OPB-12 (Table 2).

Table 1. RAPD primers with corresponding bands score and their size range together with polymorphic bands observed.

Primer code	Sequence (5'-3')	Total No. of band scored	Size range (bp)	No. of polymorphic band	Proportion of polymorphic loci (%)
OPA-02	TGC CGA GCT G	8	600-1500	8	100
OPA-03	AGT CAG CCA C	8	400-2000	8	100
OPB -05	TGC GCC CTT C	2	300-500	2	100
OPB -06	TGC TCT GCC C	4	250-500	4	100
OPB -11	GTA GAC CCG T	3	500-1800	3	100
OPB -12	CCT TGA CGC A	13	300-5000	13	100
Total		38		38	100
Average		6.33		6.33	100

Table 2. Number of polymorphic bands observed in ten variants of strawberry after PCR amplification with RAPD primers OPA-02, OPA-03, OPB-05, OPB-06, OPB-11 and OPB-12.

Strawberry genotype	OPA-02	OPA-03	OPB-05	OPB-06	OPB-11	OPB-12	Total bands
V ₁	2	0	0	2	0	12	14
V ₂	8	3	0	0	1	8	16
V ₃	7	5	0	1	1	3	12
V ₄	0	4	0	1	0	0	5
V ₅	2	5	0	0	0	7	12
V ₆	7	4	0	0	0	0	8
V ₇	7	1	2	0	0	0	8
V ₈	6	1	1	3	0	2	11
V ₉	0	3	0	2	3	3	11
V ₁₀	0	6	0	1	3	2	8
Total Bands	39	32	3	10	8	37	129

Maximum number of band (13) was shown by the primer OPB-12 and the minimum number of polymorphic band (2) by the primer OPB-05 followed by the primer OPB-11(3). A total of 129 polymorphic bands were amplified from six RAPD primers. The primers OPA-02, OPA-03, OPB-05, OPB-06, OPB-11 and OPB-12 produced 39, 32, 3, 10, 8 and 37 polymorphic bands, respectively in 10

variants of strawberry. The present experiment produced 6.33 scorable bands per primer and 6.33 polymorphic bands per primer.

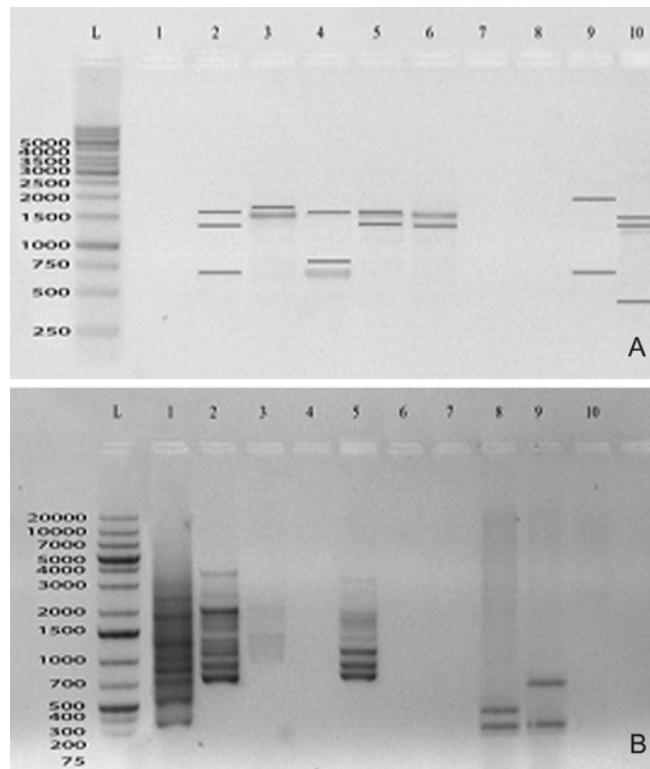


Fig. 1. A RAPD profile of 10 genotypes using primer OPA-03. B. Same as A but using primer OPB-2 OPB-12. L: Molecular weight marker (DNA ladder) Lanes 1-10: 10 genotypes of strawberry..

Genetic distance of the studied genotypes is presented in Table 3. The smaller number of pair-wise differences (high genetic similarity) among some genotypes was likely due to their genetic relatedness. Comparatively higher genetic distance 23.0 was found among V_2 vs. V_9 . The lowest genetic distance 6.0 was revealed among V_6 vs. V_7 as the genotype was morphologically very similar. Considering the genetic distance values, the results indicated that the variants were genetically different from each other which could be used in crop improvement program.

UPGMA dendrogram: Ten variants of Strawberry have been differentiated into two main clusters: V_1, V_3, V_2, V_5 and $V_4, V_9, V_{10}, V_6, V_7, V_8$ were grouped in cluster I and cluster II, respectively (Fig. 2).

Table 3. Genetic distance matrix based on Jaccard coefficient

	V ₁	V ₂	V ₃	V ₄	V ₅	V ₆	V ₇	V ₈	V ₉	V ₁₀
V ₁	0	0.667	0.815	0.950	0.696	0.920	0.917	0.792	0.826	0.923
V ₂		0	0.560	0.958	0.640	0.524	0.696	0.778	0.893	0.857
V ₃			0	0.833	0.750	0.579	0.632	0.682	0.826	0.727
V ₄				0	0.882	0.933	0.929	0.875	0.857	0.692
V ₅					0	0.684	0.909	0.920	0.913	0.762
V ₆						0	0.500	0.737	0.952	0.850
V ₇							0	0.562	1.000	0.952
V ₈								0	0.800	0.864
V ₉									0	0.467
V ₁₀										0

The cluster I was divided into two sub-clusters. In sub-cluster A of cluster I, there was V₁ and V₃ and in sub-cluster B of cluster I, V₂ and V₅ and their genetic relationship was present between sub-sub clusters.

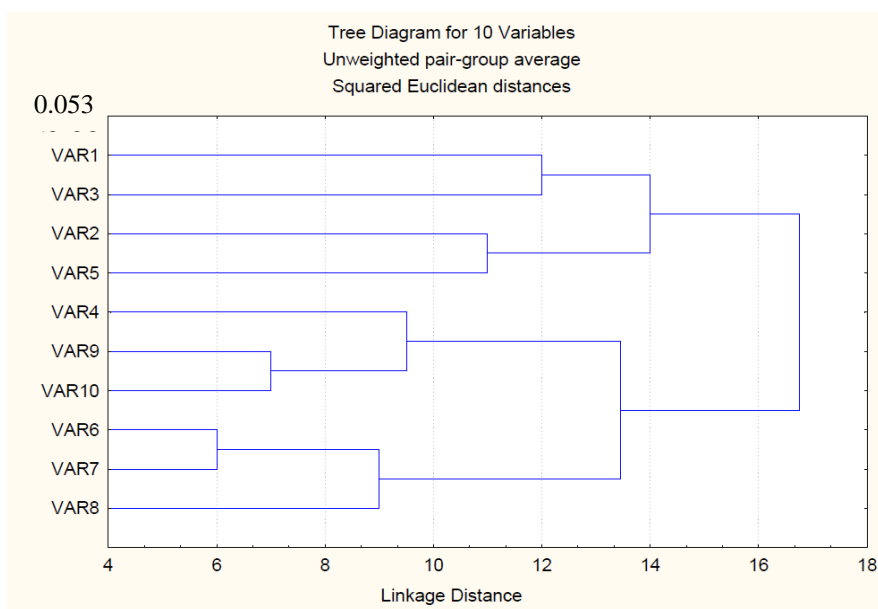


Fig. 2. Unweighted pair group method of arithmetic mean (UPGMA) dendrogram based on Jaccard coefficient, summarizing data on differentiation in 10 genotypes according to RAPD-PCR analysis.

The cluster II was divided into two sub-clusters. In sub-cluster A of cluster II, there was in V₄; V₉ and V₁₀ formed one sub-sub-cluster and in sub cluster B of cluster II, there was in V₆; V₇ and V₈ formed another sub-sub-clusters, genetic relationship were present between sub-sub-clusters.

Genotypic variations based on molecular characterization indicated that genotypes belonging to different clusters depend on their genetic components itself, but not at geographical origin at all. Therefore, it could be concluded that for further research program, especially for hybridization, genotypes selected from different clusters would provide maximum heterosis.

Ten variants of strawberry have been differentiated into two main clusters after molecular characterization. It could be concluded that for further research program, especially for hybridization, genotypes selected from different clusters would provide maximum heterosis regarding the yield.

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