PTC&B

Plant Regeneration through Somatic Embryogenesis from Leaf Sheath Derived Callus of Sugarcane (*Saccharum officinarum* L.) var. Isd -16

Mohashweta Roy*, M. Hossain¹, A. Biswas, M. K. Biswas and R. Islam

Institute of Biological Sciences, Rajshahi University, Rajshahi-6205, Bangladesh

Key words: Plant regeneration, Leaf sheath, Somatic embryogenesis, Sugarcane

Abstract

Leaf sheath explants of an indigenous variety Isd-16 of sugarcane (*Saccharum officinarum* L.) produced light yellow friable callus after culturing on to MS with 2,4-D (2 - 4 mg/l) and NAA (3 - 5 mg/l) singly. Callus formation was the maximum on MS + 3 mg/l 2,4-D. Callus underwent embryogenesis producing huge number of somatic embryos when subcultured on MS with 15 - 30 mg/l L-proline, 3 mg/l 2,4-D + 5 - 10% coconut water (v/v) and 3 mg/l 2,4-D + 10% CW (v/v) + 300 - 500 mg/l CH. L-proline significantly enhanced somatic embryogenesis and 25 mg/l L-proline in MS was the best culture medium formulation. Most of the somatic embryos germinated and developed plantlets after 1 - 2 weeks of incubation in proline-supplimented medium. On the other hand, maturation and germination of embryos were achieved on half-strength MS with or without 0.25 - 1.0 mg/l L-proline, and 5% coconut water (v/v). Somatic embryos derived plantlets were then successfully transferred to natural condition through successive phages of acclimation.

Introduction

Sugarcane is an important industrial crop and it is the only source of sugar production in Bangladesh. Due to lack of breeding facilities, biotechnological approach using gene transfer technology is successfully used for genetic improvement of sugarcane during past few years. Successful genetic transformation attempts have mostly employed embryogenic callus or cell cultures as the target tissue in several crop plants (Nayak et al. 1997, Newell et al. 1995) including sugarcane (Aftab and Iqbal 1999, Khalil 2001). Somatic embryogenesis is considered an efficient and high volume propagation system

^{*}Author for correspondence. <mroyshilpi@yahoo.com>. 1Department of Botany, Rajshahi University, Rajshahi-6205, Bangladesh.

for the production of large number of plantlets in fewer steps, with a concomitant reduction in labour, time and cost (Ammirato 1987). It offers a great potentiality in crop improvement by cell selection, genetic transformation, somatic hybrid and polyploid plant production. In recent years somatic embryos are used for developing synthetic seeds, shortening the breeding cycle and transformation. Induction of somatic embryogenesis from young leaves of sugarcane was reported by Ahloowalia and Maretzki (1983), Chen et al. (1988), Marcano et al. (2000).

In Bangladesh research in this area of tissue culture has been conducted on efficient *in vitro* plant regeneration (Islam et al. 1982, Begum et al. 1995, Samad and Begum 2000, Karim et al. 2002). So far, recent years no work has been done on the improvement of protocol for induction of embryogenic culture, which helps in transgenesis. The present study was undertaken to develop efficient and stable plant regeneration through somatic embryogenesis for a popular sugarcane var. Isd -16 of Bangladesh.

Materials and Methods

Immature leaf sheath from 6 months old field-grown plants were used as explant. Apical portion of healthy shoots were stripped to the terminal bud and attached immature leaf rolls were surface sterilized with 0.1% HgCl₂ for 9 min. Approximately 3 slices (5 × 3 mm) were taken from each cylindrical leaf roll and cultured on MS supplemented with 2 - 4 mg/l 2,4-D and 3 - 5 mg/l NAA alone and incubated in dark for callus induction. Sub-cultures were done at every 3 weeks after incubation. In order to induce somatic embryogenesis, the proliferated friable callus was transferred to MS containing 15 - 30 mg/l L-proline, 15-30 mg/l L-glutamine, 3 mg/l 2,4-D + 5 - 10% (v/v) coconut water (CW) and 3 mg/l 2,4-D + 10% (v/v) CW + 300 - 500 mg/l casein hydrolysate (CH) with 3% (w/v) sucrose (BDH, England). The pH of the medium was adjusted to 5.6 and gelled with 0.7% (w/v) agar. The cultures were also incubated in a growth chamber at 25 ± 1°C in dark. Number of somatic embryogenic callus.

For germination, the embryogenic calli with developing somatic embryos were transferred to MS with or without L-proline (0.25 - 1.00 mg/l) and 5% CW (v/v) and incubated in 16/8h light/dark condition. Regenerated plantlets with well-developed root systems were transferred to pots containing soil : manure : sand (1 : 2 : 1) mixture through successive phases of acclimation. Data collected on different quantitative characters were analyzed for mean and standard error. Each experiment was performed thrice using 15 replicates. Analysis of variance was carried out and the test of significance was done by DMRT. The data in

percentage were transformed using arc-sine/square root transformation before analysis and were transformed back to percentage for presentation. All statistical analyses were conducted using MSTAT-C software.

Results and Discussion

Young sugarcane leaves are known to be good explant sources for callus production (Ho and Vasil 1983, Brisibe et al. 1994). Therefore, young leaves of sugarcane var. Isd-16 were used as starting materials on MS containing high concentrations of 2,4-D and NAA alone for callus induction. The explants were induced to develop callus in all culture media formulations but the degree of callus proliferation varied from 13 - 91% (Fig. 1). The highest frequency (91%) of explants induced callus formation in medium having 3 mg/l 2,4-D. This result corroborates with that of Heinz and Mee (1969) and Chengalrayan and Gallo-Meagher (2001). After two subcultures at 2 - 3 weeks intervals on the same medium, these calli became light yellow, friable and embryogenic in nature.



Fig. 1. Effect of different concentrations of 2,4-D and NAA on induction of callus from leaf sheath explants of sugarcane Isd-16. Bar represents Mean ± SE. Bars marked by the same letter indicate that values are not significantly different from one other at 5% level according to DMRT.

In order to induce somatic embryogenesis, the proliferated friable calli were transferred to MS with various concentrations and combinations of L-proline, L-glutamine, 2,4-D + CW, 2,4-D + CW + CH (Table 1). Amino acids such as L proline, L glutamine, glutamic acid, asparagine play a key role for induction of

somatic embryogenesis in sugarcane. The development of somatic embryo in presence of amino acids was reported in maize (Carvalho et al. 1997), in wheat (Yadava and Chawla 2002), and also in sugarcane (Sinha et al. 2000 and Desai et al. 2004). In the present investigation, L-glutamine did not show any positive

Supplements (mg/l)	Days to embryo initiation (Mean ± SE)	% of callus induced somatic embryogenesis	No. of somatic embryos/150 mg embryogenic callus (Mean ± SE)	Remarks	
L-proline					
15	$12.84 \pm 0.22a$	51de	31.95 ± 3.72d	cal + emb	
20	8.07 ± 0.30cd	80b	$59.13 \pm 0.82 \mathrm{b}$	cal + emb + minisht	
25	$6.60 \pm 0.14 d$	91a	$75.58 \pm 2.90a$	cal + emb + ms/pt	
30	8.11 ± 0.31cd	71bc	$50.16 \pm 2.20c$	cal + emb + minisht	
L- glutamine					
15 - 30	-	-	-	cal	
2,4-D + CW% (v/v)					
3.0 + 5	$12.85 \pm 0.15a$	44e	36.69 ± 5.06d	cal + emb	
3.0 + 10	$10.12 \pm 0.18 bc$	71bc	54.36 ± 3.32bc	do	
2,4-D + CW% (v/v) + CH					
3.0 + 10 + 300	$12.98 \pm 0.65a$	60de	35.89 ± 1.23d	cal + emb	
3.0 + 10 + 400	9.58 ± 1.43 bc	80b	$58.91 \pm 1.11 \mathrm{b}$	do	
3.0 + 10 + 500	11.69 ± 1.70ab	69bcd	$49.64 \pm 0.37c$	do	

Table 1. Effect of various concentrations and combinations of L-proline, L-glutamine, 2,4-D, CW and CH on somatic embryogenesis through callus culture of sugarcane var. Isd-16.

Mean values within a column having the same letter are not significantly different according to DMRT at 5% level. cal + emb = callus with embryos, cal + emb + minisht = callus + embryos with minishoots, cal + emb + ms/pt = callus + embryos with minishoots/plentlets.

effect on somatic embryogenesis. This observation is in agreement with the results obtained by Ho and Vasil (1983). On the other hand, L-proline was found to be effective for enhancing the frequency of embryogenesis. Among the different concentrations, 25 mg/l L-proline was proved to be the most effective formulation for somatic embryogenesis from leaf sheath derived calli. A large number of somatic embryos with different developmental stages such as, globular, oblong, heart shaped, scutellar, and coleoptilar stages appeared over the surface of the embryogenic callus. Moreover, embryo induction, maturation and germination simultaneously occurred in mass of the same callus (Fig. 2a-c). The addition of L-proline to the medium facilitated the increase in reduced nitrogen in the medium and became available to the cells that might enhance the development of embryo and subsequent germination of somatic embryos to complete plantlets. It was observed that the media containing 3 mg/l 2,4-D + 10%

CW and 3 mg/l 2,4-D + 10% CW + 400 - 500 mg/l CH also showed positive effect of somatic embryogenesis. Development of embryogenic callus subjected to high contents of 2,4-D and CH in combination with CW was also reported in sugarcane (Khail 2001). Somatic embryo germination was achieved on half-strength MS with or without low concentrations of L-proline and 5% CW but the best germination (84.44%) was recorded in half-strength MS0 (Table 2, Fig. d) which corroborated with the result of Ho and Vasil (1983).



Fig. 2. Somatic embryogenesis in sugarcane: (a) White cluster of somatic embryos with various developmental stages on 25 mg/l L-proline containing MS. (b) Globular (g) and coleoptiler (cl) embryos with scutellum (sc). (c) Germinating somatic embryos with leafy scutellum (ls) and roots (r). (d). Cluster of somatic embryos germinated on half-strength MS0. (e) Acclimatized plantlets in plastic pot. (f) Plantlets growing in the field.

The age of the callus was an important factor for somatic embryo induction. Only 6 - 10 weeks old callus induced somatic embryogenesis. According to Smith and Street (1974) changes in ploidy of the cultured cells may lead to loss of embryogenic potential in long-term culture. Somatic embryogenesis occurs in short time culture and their ability decreases with increasing duration. The plantlets were successfully transferred to field through successive phases of acclimatization (Fig. 2e-f).

Supplements	Germination	
	(%)	
Half strength MS	84.44a	
Half strength MS + proline mg/l		
0.25	71.12ab	
0.5	66.66bc	
1.0	51.11c	
Half strength MS + 5% CW (v/v)	53.33c	

Table 2. Effect of half-strength MS with or without different concentrations ofL-proline and CW on germination of somatic embryos of sugarcane var. Isd-16.

Mean values within a column having the same letter are not significantly different according to DMRT at 5% level.

Present study elucidated an efficient protocol for plant regeneration *via* embryogenesis of sugarcane (*Saccharum officinarum* L.) var. Isd-16 and this will be helpful in conducting genetic transformation.

References

- Aftab F and Iqbal J (1999) Plant regeneration from protoplasts derived from cell suspension of adventive somatic embryos in sugarcane (*Saccharum* spp. *hybrid* cv. Col-54 and cv. CP-43/33). Plant Cell, Tiss. & Org. Cult. 53(3): 155-162.
- Ahloowalia BS and Maretzki A (1983) Plant regeneration *via* somatic embryogenesis in sugarcane. Plant Cell Rep. 2: 21-25.
- Ammirato PV (1987) In: Plant Tissue and Cell Culture, Plant Biology (eds. Green CE. et al.), Alan R. Liss, New York. Vol. 3. pp. 57-81.
- **Begum S, Hakim L** and **Azam MA** (1995) Efficient regeneration of plants from leaf base derived callus in sugarcane. Plant Tissue Cult. **5**(1): 1-5.
- Brisibe EA, Miyake H, Taniguchi T and Maeda E (1994) Regulation of somatic embryogenesis in long-term callus cultures of sugarcane (*Saccharum officinarum* L.). New Phytol. 126: 301-307.
- Carvalho CHS, Bohorova N, Bordall PN, Abreu LL, Valicente FH, Bressan W and Paiva E (1997) Type II callus production and plant regeneration in tropical maize genotypes. Plant Cell Rep. 17: 73-76.
- Chen WH, Davey MR, Power JB and Cocking EC (1988) Control and maintenance of plant-regeneration in sugarcane callus cultures. J. Exp. Bot. **39**: 251-261.
- **Chengalrayan K** and **Gallo-Meagher M** (2001) Effect of various growth regulators on shoot regeneration of sugarcane. In Vitro Cell. Dev. Biol. Plant. **37**: 434-439.
- **Desai NS, Suprasanna P** and **Bapat VA** (2004) Simple and reproducible protocol for direct somatic embryogenesis from cultured immature inflorescence segments of sugarcane. Current Sci. **87**(6): 764-768.

Plant Regeneration through Somatic Embryogenesis

- **Ho WJ** and **Vasil IK** (1983) Somatic embryogenesis in sugarcane (*Sacharrum officinarum* L.). I. The morphology and ontogeny of somatic embryos. Protoplasma. **118**: 169-180.
- Heinz DJ and Mee GWP (1969) Plant differentiation from callus tissue of *Saccharum* species. Crop Sci. 9: 346-348.
- **Islam AS, Begum HA** and **Haque MM** (1982) Studies on regeneration of *Saccharum officinarum* for disease resistant varieties. Plant Tiss. & Cell Cult. **5**: 709-710.
- Karim MZ, Amin MN, Hossain MA, Islam S, Hossain F and Alam R (2002) Micropropagation of two sugarcane (*Saccharum officinarum*) varieties from callus culture. Online J. Biol. Sci. 2(10): 682-685.
- Khalil SM (2001) Regeneration *via* somatic embryogenesis and microprojectile mediated co-transformation of sugarcane. Plant and Animal Genome IX conference, Town and country hotel, San Diego, Ca, January 13-17.
- Marcano AK, Guevara PM, Oropeza M and Garcia E (2000) Improvement of somatic embryogenesis in sugarcane Venezuelan cultivars. ACV. 53(4) ISSN 0001-5504.
- Nayak P, Basu D, Das S, Basu A, Ghosh D, Ramakrishnan NA, Ghosh M and Sen SK (1997) Transgenic elite indica rice plants expressing CrylAc o-endotoxin of *Bacillus thuringiensis* are resistant against yellow stem borer (*Scirpophaga incertulas*). Proceedings of the National Academy of Sciences of the USA. 94: 2111-2116.
- Newell CA, Lowe JM, Merryweather A, Rooke LM and Hamilton WDO (1995) Transformation of sweet potato [*Ipomoea batatas* (L.) Lam.] with *Agrobacterium tumefaciens* and regeneration of plants expression of cowpea trypsin inhibitor and drop lectin. Plant Sci. **107**: 215-227.
- **Samad MA** and **Begum S** (2000) Somaclonal variation of nonirradiated and irradiated calli of sugarcane (*Saccharum officinarum* L.). Plant Tissue Cult. **10**(1): 25-29.
- Sinha H, Gill M and Gosal S (2000) Regulation of somatic embryogenesis and plant regeneration in sugarcane (*Saccharum officinarum* L.). Indian J. Agri. Sci. 70(3): 181-183.
- Smith SM and Street HE (1974) The decline of embryogenic potential as callus and suspension cultures of carrot are serially subcultred. Annals of Bot. 38: 223-241.
- Yadava R and Chawla HS (2002) Role of genotypes, growth regulators and amino acids on callus induction and plant regeneration from different developmental stages of inflorescence in wheat. Indian J. Genet. 62(1): 55-60.