



Hematological and histo-architectural damages in the kidney and liver of Nile tilapia on exposure to kinalux

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Abstract

Study was conducted to assess the histo-architectural damages of kidney and liver and hematological parameters in Nile tilapia (*Oreochromis niloticus*) after sub-lethal exposure to kinalux. Fish was exposed to two sub-lethal concentrations (10% and 50%, 0.052 and 0.259ppm of median lethal concentration, respectively) of kinalux for 90 days and a parallel control was run simultaneously. Kidney and liver of exposed individuals exhibited some remarkable changes in their histology in comparison to control. Significant changes also occurred in the number of red blood cell (RBC) and white blood cell (WBC). Duration of exposure appears to have a profound effect on kidney and liver as with increasing duration of exposure histo-architectural damages become more severe.

Key words: Hematology, histopathology, kinalux 25EC, *Oreochromis niloticus*, organophosphate pesticide

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Introduction

Pesticides have been widely used all over the world to control insects, pests and disease vectors and they are one of the most potentially harmful chemicals introduced into the environment, though they have contributed considerably to human welfare, their adverse effects on non-target organisms are significant (Velisek *et al.*, 2009). These pesticides ultimately find their way into aquatic habitats such as rivers, lakes and ponds, and have been found to be highly toxic not only to fish but also to the organisms, which constitute the food chain (Arjmandi *et al.*, 2010). Sometimes this pollution may cause sudden death of fish and other aquatic organisms.

Kinalux is an organophosphate pesticide and it is an extensively used pesticide in our area. Extensive use of organophosphate compounds has resulted in a wide spread distribution of these chemicals in the

environment. They are much less persistent than the organochlorines and do not accumulate in fatty tissues. Due to their rapid biodegradability and lesser persistency in the environment, the organophosphate compounds replaced the more persistent organochlorine compounds. Organophosphorous pesticides to a large extent replaced the persistent chlorinated pesticides in the 1970s and at the beginning of 1980s. The main advantage of the organophosphate is their low cumulative ability and short-term persistence in the environment (Özcan *et al.*, 2006). These pesticides leave residues in the soil and water for several days after their application, and pose a constant threat to the non-target organisms especially fishes (Magare and Patil, 2000). Therefore, the usage of organophosphate pesticides has an impact on environment leading to the development of several adaptations such as morphological, physiological, biochemical and

behavioral ones at various levels of organization in the organisms to suit their environment.

Histopathological changes have been widely used as biomarkers in the evaluation of the health of fish exposed to contaminants, both in the laboratory (Thophon *et al.*, 2003) and field studies (Schwaiger *et al.*, 1997). One of the great advantages of using histopathological biomarkers in environmental monitoring is that this category of biomarkers allows examining specific target organs, including gills and liver that are responsible for vital functions, such as respiration, excretion and accumulation and biotransformation of xenobiotics in the fish, and serve as warning signs of damage to animal health (Gernhofer *et al.*, 2001). The present study was performed to evaluate the sub-lethal effects of kinalux on histopathological alterations in the vital organs like kidney and liver and hematological changes of Nile tilapia, *Oreochromis niloticus* as a laboratory animal model. The Nile tilapia was selected for the bioassay experiments since it is one of the most economically important freshwater fish that is extensively cultured in Bangladesh, India, Philippines, Vietnam, and many other countries.

Materials and Methods

Kinalux 25EC toxicity was assessed using tilapia as an aquaculture model in this experiment. Two hundred live and healthy tilapia (*Oreochromis niloticus*) purchased from local fish farm of Mymensingh. Standard length and average body weight of the experimental fish were 6.21 ± 0.67 cm and 4.17 ± 1.46 g, respectively and allowed to acclimatize to the laboratory conditions for two weeks before experiment.

Kinalux 25EC was purchased from the authorized dealer of Mymensingh. Ten fishes were randomly selected from the stock and exposed to different concentrations of kinalux (0.3, 0.4, 0.7, 0.8, 1.0 ppm) for 96 hours to determine the median lethal concentration (LC_{50}). Water was replaced daily with fresh kinalux mixed water to maintain constant level of kinalux during exposure period. The LC_{50} value for kinalux was 0.518 ppm. Based on the result of the 96h LC_{50} of kinalux, 100 fishes of Nile tilapia were exposed for 90 days to the sub-lethal concentrations

of 10% and 50% value of the LC_{50} of the kinalux. A control group was maintained simultaneously. All these experiments were performed in triplicates.

For hematological and histological studies, three fishes were removed from each treatment including control at fifteen days interval up to the end of 90 days exposure period. They were randomly selected from each group and immediately sacrificed by pinning through the brain and samples like blood, liver and kidney were collected from the dissected fishes. Blood samples were used to measure red blood cell count and white blood cell count (RBC and WBC) which was done immediately. RBC and WBC counts were carried out in a Neubauer chamber after saline (0.9% NaCl solution) dilution of the blood. For the histological study specimens were preserved in 10% formalin and histological examination was done using standard histological technique. All data obtained in test was analyzed using probit method and one way analysis of variance (ANOVA) followed by post-hoc testing using Duncan mean, performed with SPSS version 16. The data are presented as mean \pm standard deviation of the means.

Results

No fish died during the acclimation period before kinalux exposure, and no control fish died during toxicity tests. The probit analysis showed that the lethal concentration for 50% mortality of the fishes at 96h was 0.52ppm. In the present study, exposure of fish to sub-lethal concentrations of kinalux (0.052 and 0.259ppm) for 90 days caused significant alterations in hematological parameters (Table 1). The alterations observed in hematological parameters such as red blood cell, RBC values were decreased significantly up to 45 days of exposure and white blood cell, WBC values increased significantly after 90 days of exposure periods, respectively, in comparison with control. Furthermore, dose dependent variations were depicted in the hematological parameters of the kinalux treated fishes.

Kidney tubules and hematopoietic cells were normal and systematically arranged in the control treatment. Kinalux exposed kidney sections showed

disintegration of convoluted tubules with large intracytoplasmic vacuoles in the epithelial cells and lumen. Shrinkage and degeneration of the glomeruli, dilation within the Bowman's space were also recorded (Figure 1 A-G).

The hepatocytes and other cells of the liver in control treatment were normal and systematically

arranged. The liver of the kinalux exposed fishes showed changes including degeneration of cytoplasm in hepatocytes, rupture in blood vessels and disappearance of cell boundaries result in releasing of blood causes hemorrhage, leading to the centrilobular degeneration. Dead red blood cells were also seen in necrotic area (Figure 2 A-G).

Table 1. Mean Red Blood Cell (RBC) & White Blood cell (WBC) of tilapia exposed at 0, 0.052 and 0.259 ppm concentrations of kinalux at different days.

Parameters	Treatments	Exposure time (days)					
		15	30	45	60	75	90
RBCs ($\times 10^6/\text{mm}^3$)	0 ppm	1.46 \pm 0.055 ^a	1.91 \pm 0.051 ^a	1.89 \pm 0.035 ^a	2.31 \pm 0.032 ^a	2.7 \pm 0.035 ^a	3.01 \pm 0.094 ^a
	0.052 ppm	1.35 \pm 0.031 ^b	1.46 \pm 0.025 ^b	1.29 \pm 0.025 ^b	1.95 \pm 0.041 ^b	1.59 \pm 0.050 ^b	2.80 \pm 0.058 ^b
	0.259 ppm	1.25 \pm 0.050 ^b	1.18 \pm 0.031 ^c	1.18 \pm 0.026 ^c	1.86 \pm 0.040 ^b	1.63 \pm 0.045 ^c	2.73 \pm 0.030 ^b
WBCs ($\times 10^4/\text{mm}^3$)	0 ppm	2.13 \pm 0.252 ^a	1.59 \pm 0.074 ^a	1.92 \pm 0.038 ^a	1.85 \pm 0.040 ^a	2.11 \pm 0.096 ^a	2.63 \pm 0.076 ^a
	0.052 ppm	2.88 \pm 0.091 ^b	2.84 \pm 0.061 ^b	2.75 \pm 0.107 ^b	2.70 \pm 0.106 ^b	2.67 \pm 0.051 ^b	2.71 \pm 0.87 ^a
	0.259 ppm	3.17 \pm 0.246 ^c	3.10 \pm 0.215 ^b	3.18 \pm 0.124 ^c	2.77 \pm 0.122 ^b	2.75 \pm 0.072 ^b	2.79 \pm 0.099 ^c

Values are mean \pm standard deviation, different alphabetic superscripts a,b,c indicates significant differences at $p < 0.05$ level.

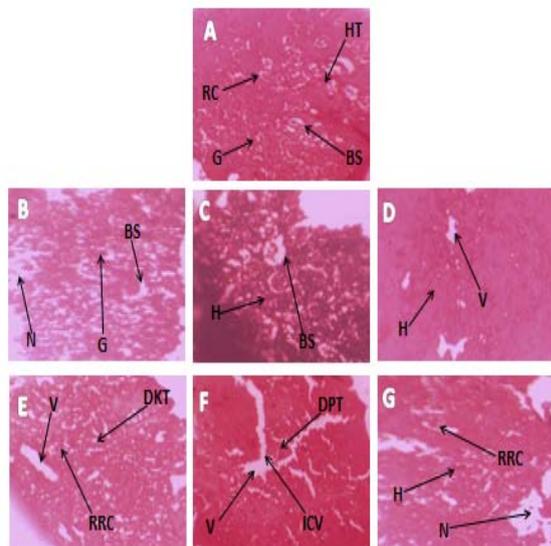


Figure 1. Histoarchitectural changes in kidney (H & E stained, X100) exposed to Kinalux (A) 0 ppm, (B), (C) and (D) at 0.052 ppm; (E) and (F) and (G) at 0.259 ppm at 30, 60 and 90 days. Arrows are indicating RC-Renal corpuscle, G-Glomeruli, HT-Haepatopoietic tissue, BS-Bowman's space, N-Necrosis, DKT-Degenerated kidney tissue, V-Vacuolation, RRC-Ruptured renal corpuscle, DHT-Degenerated haepatopoietic tissue, ICV-Intra-cytoplasmic vacuoles, H-Hemorrhage

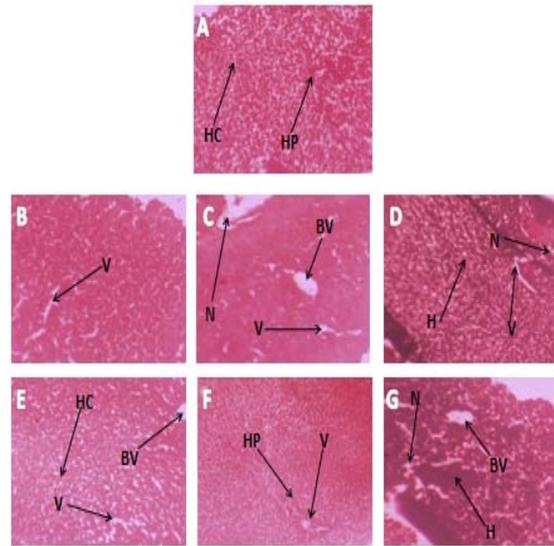


Figure 2. Histoarchitectural changes in liver (H & E stained, X100) exposed to Kinalux (A) Control, (B), (C) and (D) at 0.052 ppm; (E) and (F) and (G) at 0.259 ppm at 30, 60 and 90 days respectively. Arrows are indicating HC - Haepatic cell, HP - Haepato-pancreas, V - Vacuoles, H - Hemorrhage, BV - Blood vessel, N - Necrosis

Discussion

The present study revealed that an organophosphate pesticide, kinalux exposure induced histo-architectural alterations in kidney and liver and hematological parameters of Nile tilapia, severity of the alterations were dose and duration dependent. The exposure of aquatic organisms to sub-lethal concentration of pesticides in their environment may result in various biochemical, physiological and histological alterations in vital tissues (Geraldine *et al.*, 1999).

Insecticides may lead to alterations in the blood parameters and hematological profile of fish which can be investigated as biomarker in pollution monitoring (Banaee *et al.*, 2008). The present study shows significant decrease in RBC count following exposure to kinalux that might be due to hemolysis and shrinkage of hematopoietic blood cells by the toxic effect of pesticide. The increase rate of RBC breakdown or reduction rate of RBC formation might also be responsible for reduction in RBC count. While correlating with the histopathological changes, the destruction of the hematopoietic tissue in the head kidney was evident might be responsible for decrease in RBC (Das, 1998). The reduction number of erythrocytes suggested hypoxic effect prevailing over the body tissues of the fingerlings due to damaging effect on the gill tissue. Reduction in number of erythrocytes reported in this investigation indicated that *O. niloticus* exposed to sub-lethal concentrations of kinalux became anemic, which is similar to the findings of Wedemeyer *et al.* (1984). A reduction in the erythrocytes number of Atlantic salmon (*Salmo salar*) and channel catfish (*Ictalurus punctatus*) exposed to malachite green were also reported by Glagoleva and Malikova (1968) and there also was a reduction in erythrocytes number in Nile tilapia exposed to gammalin 20 and actellic 25EC by Omoregie *et al.* (1990).

In this study a significant increase in the WBC count was observed in fish exposed to sub-lethal dosages in 90 days. In the presence of foreign substances or under pathological conditions leucocytosis in fish may be the consequence of direct stimulation of immunological defense (John, 2007). As leukocytes

fight against any toxicant introduced into the blood stream, this increase suggests that fish has developed a certain degree of tolerance by stages during the stress conditions. The increase in WBC count can be correlated with an increase in antibody production which helps in survival and recovery of the fish exposed to lindane and malathion (Joshi *et al.*, 2002). This also helps in the removal of cellular debris of necrosed tissue at a faster rate (John, 2007). Consistent supports to the above several results showed a significant increase in the WBC (Adedeji *et al.*, 2009; Kumar *et al.*, 2011; Saravanan *et al.*, 2011).

In sub-lethal dosages of kinalux several alterations such as degeneration of kidney tubules and hematopoietic tissue, vacuolization, mild to severe necrosis, pyknosis and hemorrhage were recorded. Shrinkage and degeneration of the glomeruli, dilation within the Bowman's space were also seen. Similar to the present findings necrosis, pyknosis, hemorrhages, vacuolation, degeneration of kidney tubules and hematopoietic cells were reported on *Anabas testudineus*, *Channa punctatus* and *Barbodes gonionotus* at 3.75 ppm, 2.26 ppm and 2.26 ppm, of diazinon 60EC, respectively for 7 days by Rahman *et al.* (2002).

Fish liver histopathology is an indicator of chemical toxicity and it is a useful way to study the effects of exposure of aquatic animals to toxins present in the aquatic environment (Fernandes *et al.*, 2008). In the present study, histo-architectural changes of liver were observed after exposure to the kinalux such as hypertrophy of hepatocytes, mild to severe necrosis, blood spilling, ruptured central vein, lipid droplet and vacuolation. The recorded results in the present study were similar to those observed by Kunjamma *et al.* (2008) recorded pyknotic nucleus, protein precipitation, pancreatic acini appeared with the loss normal structure and necrosis of the hepatic and pancreatic tissue in freshwater fish (*Catla catla*) and (*Oreochromis mossambicus*) treated with chlorpyrifos.

Conclusion

The results of the present investigation reveal that under experimental condition, hematological

parameters and some organs of tilapia were sensitive to kinalux exposure. These findings permit us to conclude that kinalux is highly toxic to fish. Hence, the presence of kinalux in waterways could have adverse impact on the survival of the fish. Therefore it is necessary to monitor, the level of kinalux in aquatic environments.

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