

Chemotaxis of Biofilm Producing *Pseudomonas* spp. towards Refined Petroleum Oil

S. Dutta* , P. Singh

Department of Microbiology, Kanya Gurukul Campus, Gurukul Kangri University, Haridwar-249407, India

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Abstract

The bioavailability of organic contaminants to the degrading bacteria is a major limitation to efficient bioremediation of sites contaminated with hydrophobic pollutants. Studies were conducted to analyze the presence of biodegrading bacteria on the sites of refinery and garage oil spills with abundance of hydrocarbons. Contaminated soils were collected from these rich hydrocarbon sites and bacteria were isolated from samples using regular bacterial media enriched with petroleum oil. These isolates were characterized for their efficiency to utilize refining petroleum oil as their energy source and then the most common isolate was characterized. Further investigations examine the formation of biofilm by naturally existing oil-degrading bacteria on refined petroleum oil degradation. Here also studied the abilities of these *Pseudomonas* strains to respond chemotactically to refined petroleum oil. Microbial chemotaxis plays an important role for formation of biofilm. 16S rDNA analysis of the best degrader was found to belong to the *Pseudomonas* species. Interestingly, one of the best isolates was found to be close to *Pseudomonas aeruginosa* family.

Keywords: Bioremediation; Biofilm; Chemotaxis; *Pseudomonas aeruginosa*.

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1. Introduction

Petroleum is one of the most important energy resources and a raw material of the chemical industry. The world depends on oil and the use of oil as fuel has contributed to intensive economic development. Although petrochemical plants and oil refineries are beneficial to society, they produce a large amount of hazardous waste. Moreover, oil spills during exploration, transportation, and refining, have caused serious environmental problems [1]. Eight million tons of petroleum is spilled into the

* Corresponding author: shreyasridutta@gmail.com

environment every year worldwide. Oil contamination is a severe threat for our environment and therefore invites general concern. Consequently, the remediation of oil-polluted sites has become an important issue worldwide. Petroleum was a mixture of a very large number of different hydrocarbons; the most commonly found molecules were alkanes (paraffins), cycloalkanes (naphthenes), aromatic hydrocarbons, or more complicated chemicals like asphaltenes. Petroleum diesel, also called petro diesel or fossil diesel was the most common type of diesel fuel. It was produced from the fractional distillation of crude oil between 200°C (392°F) and 350°C (662°F) at atmospheric pressure, resulting in a mixture of carbon chains that typically contain between 8 and 21 carbon atoms per molecule. As well as kerosene was a mixture of hydrocarbons. The chemical composition depends on its source, but it usually consists of about 10 different hydrocarbons, each containing 10 to 16 carbon atoms per molecule. The main constituents were saturated straight-chain and branched-chain paraffins, as well as ring-shaped cycloparaffins (also known as naphthenes). Kerosene was less volatile than gasoline. Therefore, refined oils (i.e petrol, diesel, kerosene) have been considered as priority pollutants which exert bio-hazardous effects on both human and other living organisms in the environment. Fortunately, this mixture represents an excellent substrate in the study of hydrocarbon bioremediation due to its composition. The emphasis of research now is to exploit the hydrocarbon degradation abilities of the microbial population to raise the rate of biodegradation found naturally to significantly higher rates. The enhancement of refine petroleum oil bioremediation only depends on potential strains to run the process effectively. In biological treatment it is always necessary to perform laboratory feasibility test to determine the microbial potential to degrade the pollutant [2]. One of the major factors that impede the process bioremediation is bioavailability of hydrophobic contaminants to the hydrocarbon utilizing microorganisms. Although numerous studies focused on biofilm reactor in the field of bioremediation [3], pollution by oil and other hydrocarbons solely during the oil spillage needs further attention in the context of bioavailability of microorganisms. It has been investigated earlier that this major limitation can be improved by exploiting chemotactic bacteria [4-6]. Microbial chemotaxis plays important role in surface colonization and biofilm formation [7]. Biofilm community is diverse and relatively stable for longer period of time [8]. The phenomenon of chemotaxis by the organisms towards the pollutants and the simultaneous attachment-detachment process maintains a constant load of biomass to the affected site in the water bodies. Potential strains have the ability of chemotaxis towards petroleum hydrocarbon. It was shown that *Pseudomonas* has positive chemotaxis to naphthalene and growth on naphthalene or salicylate induced the chemotactic response [9]. Several authors have also studied the bacterial chemotaxis toward environmental pollutants [10]. Ortege *et al.* [11] studied bacterial chemotaxis towards environmental pollutants.

The objective of this study was to isolate and identify potential microorganisms from contaminated site and examined their ability to form biofilm and chemotaxis towards refine petroleum oil.

2. Experimental

2.1. Soil sample

Soil samples were collected from Indian Oil refinery, Haldia (W.B, India). Subsurface soil samples contaminated with refined products were collected from local area, Haridwar, India. The soil samples were collected in pre-sterilized plastic bags. The samples duly labelled were stored at 4°C for further study.

2.2. Culture enrichment isolation and characterization of strain

Soil samples were sieved moist using a 2 mm mesh screen and thoroughly mixed. 10 g of soil was added to 95 mL deionized water and then vortexed. The suspension was allowed to settle down and supernatant (5 mL) was used as an inoculum in 100 mL MSM (Minimal Salt Medium) containing 2.5 mL of synthetic mixture of refined petroleum oil (petrol, diesel and kerosene; 1:1:1) in an Erlenmeyer flask for 48 hr at 35-37°C on a rotary shaker at 100 rpm. An enrichment of culture was carried out in three consecutive batches, each having a span of 15 days and enriched by using previous growth as inoculums for the next. MSM [12] containing following composition [in (g/L)]: KNO₃ (1), MgSO₄·7H₂O (1), CaCl₂·6H₂O (0.1), FeSO₄ (0.05), trace element sol, 250 mL; phosphate buffer (1M; pH 6.8), 20 mL; and distilled water, 980 mL. Trace element solution comprised (g): SnCl₂ (0.05); KI (0.05), LiCl (0.05), MnSO₄·4H₂O (0.08), HBO₃ (0.05), ZnSO₄·7H₂O (0.10), CoCl₂·6H₂O (0.10), NiSO₄·6H₂O (0.10), BaCl₂ (0.05), ammonium molybdate (0.05) and distilled water 1000 mL (all salts were dissolved in defined sequence only). In final stage, 1 mL of this suspension was evenly spread on to a refined petroleum oil agarose plate and then incubated at 37 °C for 3 days. Selection of microorganisms was based on better ability to grow in presence of refined petroleum oils as sole source of carbon in growth media. The isolated microorganisms were tested on the basis of colonies's size, shape, margin, consistency, opacity, elevation, pigmentation, Gram reaction and cell morphology as described in Bergey's Manual of Determinative Bacteriology [13]. Biochemical properties tested include, production of catalase, indole, urease, oxidative fermentation of sugars, methyl red test, Voges Proskauer test and citrate utilization test, gelatin utilization/liquefaction test, starch utilization test etc. [13]. Motility tests were done by stabbing cells in semisolid nutrient agar (0.7% agar) [14].

2.3. Growth characteristics in different oil

This experiment was performed according to the method of Salam *et al.* [15] with slight modification. For that purpose, all selected bacterial species were grown in MSM with excess amount (5 mL) of refined petroleum oil separately such as petrol, diesel and kerosene. The individual bacterial isolate from overnight culture at log phase of growth were adjusted with sterile distilled water to give a bacterial cell count of 1.0X10² CFU/g

and transferred to 100 mL conical flasks containing 50 mL of sterile-define MSM with 5 mL (10% v/v) of petrol, diesel and kerosene separately for each isolate. The flasks were then incubated in a shaker at 150 rpm at normal temperature for 7 days. At the end of the experiment, sets of flasks were used for the enumeration of the microbial population by pour plate technique and serial dilution method on plate count agar. Optical densities of inoculated sets of flasks were also measured by spectrophotometry (Systronics-model no: 2205) at 600 nm against control flasks.

2.4. Oil biofilm development

The isolated strains were first tested for biofilm formation on 18 mm glass cover slips being immersed in 15 mL MSM with 1% synthetic mixture of refine petroleum oils in 50 mL sterile falcon tubes. The organism was inoculated and incubated at 30°C for 4 days. The cover slips were recovered from the culture tubes, washed thoroughly in 1% saline solution aseptically, air-dried and Gram-stained. Formation of biofilm was viewed under 100X oil immersion objective using compound light microscope [14].

2.5. Scanning electron microscopy analysis

The cell-substrate physical interaction on cover slip was examined by scanning electron microscopy. In the case of the scanning electron microscopy, bacterial aggregates on glass cover slip were fixed with 5% glutaraldehyde for 30 min at room temperature and stored in refrigerator for overnight. Finally, samples were transferred in Advanced Instrumentation Research Facility, Jawaharlal Nehru University, New Delhi-110067 for SEM (Zeiss EVO40) analysis [16].

2.6. Chemo taxis movement

Chemo taxis was tested with drop assay [17]. For this assay, 40 mL of cells were harvested in the logarithmic phase of growth and resuspended in 12 mL of chemotaxis buffer (100 mM potassium phosphate [pH 7.0], 20 mM EDTA). A small amount of a test attractant (synthetic mixture of refine petroleum oils) was added to the center of a dish (1 mL). A chemotaxis response of cells to the added compound resulted in the formation of a ring of turbidity near the center of the petri dish after 24 h.

2.7. Molecular characterization of selected bacterial species

Bacterial strain of PS-I was identified by 16 S rDNA sequence structure from Royal Life Sciences Pvt. Ltd. (affiliated to MIDI Sherlock, USA). The 500 bases sequence of 16S rDNA was obtained by sequencing. The electrophoregram of partial sequence was aligned with blast search of NCBI database.

3. Results and Discussion

Three bacterial strains capable of using hydrocarbon as sole source of carbon, were isolated of which two (Ps-II, Ps-III) were recorded from the soil sample of Haldia and one from oil contaminated soil (Ps-I) of a nearby garage of Haridwar. The pure refine oils utilizing bacterial strains were isolated on petroleum agarose plate. After cultural (Table 1) morphological (Table 2) and biochemical identification (Table 3), these tested isolates were identified as *Pseudomonas* spp.(Ps-I, Ps-II, Ps-III).

Table 1. Cultural characteristics of selected bacterial isolates.

Isolated Bacteria	Size	Shape	Gram's reaction	Motility
Ps-I	Small	Rods	Gram negative	Motile
Ps-II	Small	Rods	Gram negative	Motile
Ps-III	Small	Rods	Gram negative	Motile

Pseudomonas is the most frequently reported and so far most studied of hydrocarbon degradation genus [18-20]. The isolated strains were examined further for substrate utilization. Over the year, substrate specificity in newly isolated organisms has been a routine diagnostic test for speciation and identification. After substrate utilization study, it was concluded that out of three *Pseudomonas* spp., Ps-I showed maximum growth in presence of kerosene (56.27×10^7 CFUg⁻¹) and diesel (145.90×10^7 CFUg⁻¹) whereas Ps-II showed maximum growth on petrol (4.60×10^7 CFUg⁻¹). But in case of turbidity, Ps-II showed maximum growth in presence of petrol (0.19 ± 0.07) and diesel (0.30 ± 0.029) whereas Ps-I showed maximum growth in presence of kerosene (0.95 ± 0.013). Although Ps-III was a *Pseudomonas* spp. but it showed very poor growth in presence of petrol, diesel and kerosene (Table 4). These results clearly indicate that different oils were degraded and utilized by all the strains in various proportions, depending on the complexity and aliphatic and aromatic nature of the sample oil dependent on bacterial species as well. The biofilm study demonstrated that the isolated bacterial strains possess the ability to form biofilm on glass surface when grown in MSM in presence of synthetic mixture of refined oils as a sole source of carbon (Fig. 1).

Table 2. Morphological characteristics of selected bacterial isolates.

Isolated Bacteria	Size	Shape	Elevation	Margin	Opacity	Texture	Pigment
Ps-I	Big	Irregular	Slightly raised	Irregular	Opaque	Smooth	Green
Ps-II	Big	Irregular	Slightly raised	Irregular	Opaque	Smooth	Brownish green
Ps-III	Big	Irregular	Slightly raised	Irregular	Opaque	Smooth	Bluish green

Table 3. Biochemical characterisation of bacteria.

Name of Biochemical test	Code number of isolates		
	Ps-I	Ps-II	Ps-III
Urease production	+	+	+
Nitrate reduction	+	+	+
Oxidase	+	+	+
Catalase production	+	+	+
Gelatin utilization	+	+	+
Starch hydrolysis	-	-	-
Indole production	-	-	-
M.R test	+	+	+
V.P test	-	-	-
Citrate utilization	+	+	+
Glucose utilization	+	+	+
Sucrose utilization	-	-	-
Mannitol utilization	+	+	+
Lactose utilization	+	+	+
Maltose utilization	+	+	+

Interestingly, a thick biomass was observed to be aggregated near the oil water interface on the glass slide. The bioavailability of organic contaminants to the degrading bacteria is a major limitation to efficient bioremediation of sites contaminated with hydrophobic pollutants; such limitation of bioavailability can be overcome by steady state biofilm-based reactor. The isolated *Pseudomonas* spp. in this study showed their ability to form biofilm on glass surface in MSM in presence of refine oil. In case of Ps-III showed lower attachment than other two *Pseudomonas* spp. (Ps I & Ps II). In addition to this, the chemotaxis elucidated by the bacterial adhering on biofilm could also support the fact of bacterial motility towards the refine oil. A positive chemotaxis response was visualized by the accumulation of a cloud of motile cells around the drop of oil after 24 h. Cells of Ps-I formed a turbid ring around near the drop of refine oil to the centre of petridish. Cells of Ps-II also responded to chemotaxis movement and formed a broader and diffuse ring around the drop of oil but Ps-III formed a ring far away from the drop of oil. Ps-I showed a much denser ring of turbidity around the site of drop. These results indicate that out of three species of Ps-I had a greater chemotaxis response towards refine oil within very short period of time, whereas Ps-II slowly response but Ps-III was not sufficiently sensitive towards refines oil (Fig. 2).

Table 4. Growth characteristics of different oil. (* Average of triplicates)

Code no of isolate	Petrol		Diesel		Kerosene	
	Turbidity*	C.F.U/mL*	Turbidity*	C.F.U/mL*	Turbidity*	C.F.U/mL*
Ps-III	0.09±0.002	1.52X10 ⁷	0.10±0.012	25.60X10 ⁷	0.13±0.005	5.6X10 ⁷
Ps-II	0.19±0.079	4.60X10⁷	0.30±0.029	117.25X10 ⁷	0.07±0.014	1.04X10 ⁷
Ps-I	0.18±0.058	2.57X10 ⁷	0.28±0.012	145.90X10⁷	0.95±0.013	56.27X10⁷

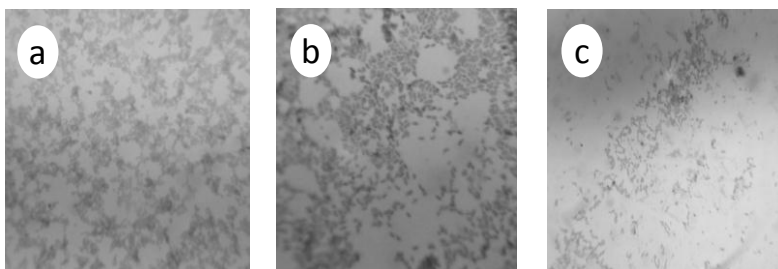


Fig. 1. Oil-biofilm formation of three bacterial isolates, [a] Ps-I, [b] Ps-II and [c] Ps-III.

Ability to form biofilm on various surfaces was always advantageous for the microorganisms in terms of survival, metabolism, adaptation, and propagation [21,22]. One of the major limitations faced in the process of bioremediation was the bioavailability of organic compounds on site [23]. Early studies [24] that indicate biofilm forming bacteria could be employed to overcome this limitation although the application of steady state biofilm in bioremediation was not well established. Studies indicate that biofilm-mediated bioremediation was a proficient approach and safer option since cells in biofilm had better chance of survival and adaptability especially during the stressed conditions [25,26]. Establishment of biofilm on glass slides was reported previously [26] where the artificially glued microorganisms showed excellent attenuation of crude oil in liquid waste in batch culture. Peacock *et al.*, [22] showed altered profile of expressed proteins, specifically type VI secretion system in biofilm forming *Marinobacter hydrocarbonoclasticus SP17* at alkane water interface. In this study Ps-I & Ps-II showed good result on biofilm formation but Ps-III very limited attachments. SEM analysis permitted a 2-dimensional observation of Ps-I attached on glass cover slip. Cells were seen densely gathered on the cover slip, (Fig. 3A), linked together as clusters (Fig. 3B).

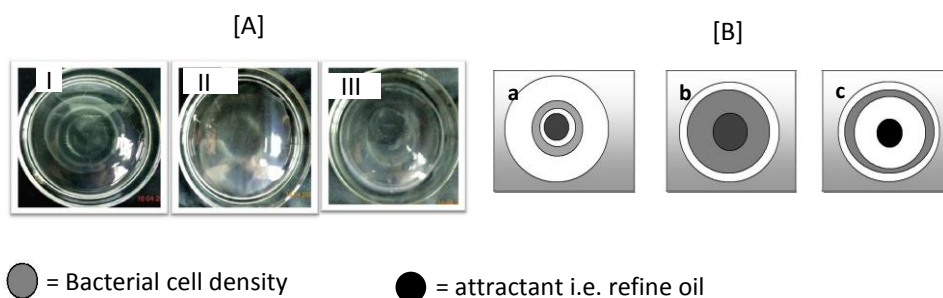


Fig. 2. [A] Photograph of chemotaxis responses of *Pseudomonas* species ([I]: Ps-I, [II]: Ps-II, [III]: Ps-III) to refine oil. [B] Diagram of the plate [a]=[I], [b]=[II], [c]=[III]. Drawings were used because often only a small amount of contrast was visible between the accumulated cells and the background. Note that Ps-I form a broader nearer to drop of refine oil. Cells of Ps-III form a ring far away from drop of SMRP oil than cells of Ps-II. Which form a diffuse ring around the drop of refine oil.

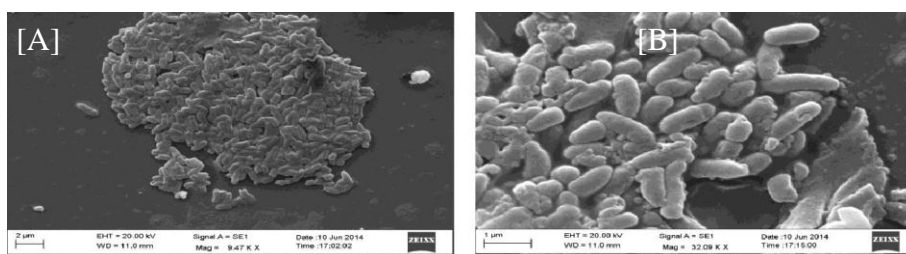


Fig. 3. (A) Cell cluster on Glass cover slip. (B) Clearly seen a rod shaped *Pseudomonas* sp. cells present on glass cover slip.

Recent results with the biodegradation of carbon tetrachloride and naphthalene had shown the potential of chemotaxis to enhance biodegradation in laboratory-scale microcosms. Indeed, chemotaxis had already been studied in bacteria able to degrade a wide variety of organic pollutants such as naphthalene, BTEX (Benzene, Toluene, Ethylbenzene, and Xylene), and pesticides. Parales *et al.* [27], showed toluene-degrading bacteria were chemotaxis towards the environmental pollutants, benzene, toluene, and trichloroethylene. In addition, Harwood and Hawkins [26], showed the chemotaxis of *Ralstonia* to herbicide. These results indicated that out of three species of *Pseudomonas* sp. I had a greater chemotaxis response towards refined oil within a very short period of time whereas *Pseudomonas* sp. II slowly response but *Pseudomonas* sp. III was not sufficiently sensitive. Bacterial chemotaxis toward environmental pollutants have an important role in bioremediation. Sequence aligned with NCBI database gave 98% similarity with *Pseudomonas aeruginosa*. Therefore, bacterial strain of Ps-I was identified as *Pseudomonas aeruginosa*.

4. Conclusion

Pseudomonas aeruginosa strain, isolated from petroleum oil contaminated sites, showed that their ability to form biofilm and chemotaxis response towards refined petroleum oil. Therefore, these isolated *Pseudomonas aeruginosa* strain could be considered for future use for bioremediation of contaminated spilled oil. However, further studies are needed to evaluate the potential of the isolated strains to degrade hydrocarbons in situ, in natural environmental conditions.

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