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Effect of gamma irradiation on shelf life and quality of indigenous chicken meat

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ABSTRACT

The experiment was conducted on fresh indigenous chicken meat treated with 0 (non-irradiated), 1, 2 and 3 kGy ⁶⁰Co gamma irradiation and stored for 0, 30 and 60 days at -20°C to investigate the effects on proximate components, sensory attributes, and physicochemical, biochemical and microbial changes in meat quality. Data were analyzed under 4x3 factor CRD design of experiment in GLM procedure of SAS statistical package. The results showed that irradiation groups had significantly (p<0.05) higher color and tenderness of meat compared to that of non-irradiated group. The 2 kGy group showed significantly (p<0.05) higher Dry matter (DM) and Ether extract (EE) whereas the cooking loss, Free fatty acid (FFA), Peroxide value (PV), and Thiobarbituric acid reactive substances (TBARS) levels were higher in 3 kGy irradiated group. With the advancement of storage periods pH significantly (p<0.05) decreased. The 2 kGy irradiation group showed significantly (p<0.05) lower numbers of Total viable count (TVC), Total coliform count (TCC), Total yeast and mold count (TYMC) compared to non-irradiated group. From this study, it may be concluded that the 2 kGy irradiated group had positive effects on sensory evaluation, biochemical and microbial qualities of indigenous chicken meat to increase the shelf life and the quality of indigenous chicken meat.

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Introduction

Now-a-days many preservation techniques have been developed which include cooking, freezing, fermenting, salting, drying and pickling (Choi et al. 2009; Kim et al. 2009). These methods have been used to reduce the number of microorganisms and increase the shelf-life and safety of meat (Farkas, 2004). Irradiation is one of the safest methods to maintain quality and safety of meat and meat products. Therefore, meat scientists have made an effort to develop new technologies that can be used not only to secure the safety issues but also to improve the quality of meat (Artes et al., 2007). Irradiation is recognized as an effective, widely applicable food processing technique. Currently, several countries have permitted food irradiation and more than half a million tons of food are irradiated annually (Eustice and Bruhn, 2013). Gamma irradiation is a physical means of sterilization or decontamination where products are exposed to gamma rays.

It is well established that meat has several key nutritional factors, like lipids, proteins with high biological value, trace elements, and vitamins (Wyness *et al.*, 2013). Meat quality has intrinsic characteristics such as color, flavor, tenderness, texture, juiciness, and overall acceptability. The nutritional properties depend

on animal genetics, feeding, and livestock practices and on the post-mortem processes that take place during the conversion of muscle into meat (Hocquette *et al.*, 2012).

The chemical and biochemical reactions with the free radicals produced by irradiation results in modification of the oxidation-reduction environment within meat products, and accelerates lipid oxidation, protein oxidation, off odor (Xiao et al., 2011), and alters meat color (Nam and Ahn, 2002). With the approval of irradiation to improve the safety of poultry meat, concerns have been raised about the negative effects of irradiation on meat quality, which include lipid oxidation, protein oxidation, color, and odor. The negative effects of irradiation on indigenous chicken meat quality not yet been studied in our country. Lately Bangladesh is producing 72.60 Lakh metric ton of meat vis-à-vis demand of 72.14 Lakh MT (DLS, 2018) where chicken is contributing more share (around 50%). It indicates we are in surplus 0.46 Lakh MT meats production per year. As a result now we have opportunity to seek foreign markets to export our excess meats. To overcome the international trade barrier irradiation can be an effective way to increase the shelf life and safety of meats. To best of our knowledge our research team conducted first experiment in The Animal Science Laboratory of BAU on gamma irradiation of

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different meats (chicken, beef, chevon and mutton) to evaluate the quality and shelf life in Bangladesh. Therefore, the study was carried out to determine the effect of gamma irradiation on sensory, proximate, biochemical and microbial qualities of chicken meat to increase the shelf life and the quality of indigenous chicken meat.

Materials and Methods

Sample collection and processing

The study was conducted in 2017 in the Department of Animal Science, Bangladesh Agricultural University, Mymensingh. About 3.5 kg of fresh indigenous chicken meat samples from four birds at the age of 12-18 month (on the basis of seller interview) were collected from local market of Mymensingh. The treatment time of sample was 24 hours after slaughtered.

The samples were divided into four treatment groups. Each group was exposed to the irradiation dose of 0 (T0), 1 (T1), 2 (T2) and 3 kGy (T3)) at the Bangladesh Institute of Nuclear Agriculture. Meat sample was irradiated at Cobalt 60 GC-5000 (BRIT, India) machine; whose central dose rate was 4.29 kGy /hr. Time had taken for each group of sample was 14 min, 28 min and 35 min 55sec which was treated with 1.00, 2.00, and 3.00 kGy, respectively.

Proximate components

Dry Matter (DM), Ash, Crude protein (CP), Ether extract (EE) was determined as per the standard procedures of Association of Official Analytical Chemists (AOAC, 1995). All determination was done in triplicate and the mean value was reported. The proximate determination was conducted to know the nutrient composition of chicken meat changed with irradiation.

Sensory evaluation

Sensory evaluation was executed by a trained 6-member panel (color, flavor, tenderness, juiciness and overall acceptability). Prior to sample evaluation, all panelists participated in orientation sessions to familiarize with the scale attributes (color, smell, juiciness, tenderness, and overall acceptability) of indigenous chicken meat using an intensity scale. Each sample was evaluated by using a 9-point hedonic scale (9 = like extremely, 8 = like very much, 7 = like moderately, 6 = like slightly, 5 = neither like nor dislike, 4 = dislike slightly, 3 = dislike moderately, 2 = dislike very much and 1 = dislike extremely) (Pena *et al.*, 2016). Sensory evaluation was accomplished at 0 days and repeated at 30 and 60 days, respectively.

Physicochemical and bio-chemical assessment

pH value of raw meat and cooking loss was measured using pH meter (Hanna HI99163) from raw meat homogenate. The homogenate was prepared by blending 5 g of meat with 10 ml distilled water. FFA value, POV value and TBARS value were determined by (Sharma *et al*, 2012). All determination was done in triplicate and mean value was reported.

Microbial assessment

Ten grams of sample were aseptically homogenized after adding 90 ml of sterile solution in a sterile Stomacher bag for 2 min (BagMixer® 400, Interscience, France). Consequently the diluents were planted onto aerobic plated count agar (Difco Laboratories), incubated at 37°c for 45 h. The total number of colonies observed on plate of each sample after incubation was counted and expressed as log of colony forming units per gram (Log CFU/g).

Statistical model and analysis

The proposed model for the planned experiment was a factorial experiment with two factors-A (Treatments) and B (Days of Intervals) is:

$$yijk = \mu + Ai + Bj + (AB)ij + åijk i = 1,...,A; j = 1,...,B;$$

 $k = 1,...,n$

Where: yijk = observation k in level i of factor A and level j of factor B

 μ = the overall mean

Ai =the effect of level i of factor A

Bj = the effect of level j of factor B

Data were statistically analyzed using SAS Statistical Discovery Software (2002-2003), NC, USA. DMRT test was used to determine the significance of differences among treatment means.

Results and Discussion

Sensory evaluation

Color, flavor, tenderness, juiciness and overall acceptability score at different treatment was 5.33 to 6.33, 6.33 to 5.44, 5.55 to 6.17, 5.88 to 6.22 and 4.55 to 5.16, respectively (Table 1). Color was significantly (p<0.05) increased with the increasing level of treatments but significantly decreased color of meat with longer storage time. The range values for three observations of different days of intervals for the color, flavor, tenderness, juiciness and overall acceptability score were 6.58 to 5.33, 6.33 to 5.16, 6.75 to 5.08, 6.66 to 5.16 and 5.50 to 4.33, respectively. Kim et al. (2002) also found that the development of red color in irradiated meat was due to the production of gas, especially CO. Contradictory reports were found by Souza et al. (2007) who investigated the influence of radiation on the levels of iron and color of pigments of thighs and chicken

breast meat irradiated at doses 0, 1 and 2.0 kGy and found that color was not influenced by those doses. The present findings also were not in agreement with Al-Bachir *et al.* (2010) who found that color of chicken kabab product were not influenced by the irradiation treatment. It may be due to species difference. Flavor, tenderness, juiciness and overall acceptability were significantly decreased with increasing days of interval. Kanatt *et al.* (2015) found that in chicken, lamb and buffalo meat tenderization is increased with dose-

dependent manner. The lowest test score of juiciness was reduced to 5.16 in all treatments after 60 days of storage. Juiciness influenced by the cut of meat and how long the meat is cooked. Among all treatment groups except flavor higher dose 3 kGy showed higher value due to higher lipid oxidation of chicken meat. There was no interaction between treatment and days of interval among all treatments for all variables.

Table 1. Sensory-attributes (mean ± SE) in irradiated indigenous chicken meat samples compared at different storage period

Parameters	DI		Mean ± SE	Level of significance					
		T_0	T_1	T_2	T ₃	-	Т	DI	T*DI
	0	6.33 ± 0.58	6.33 ± 0.33	6.67 ± 0.33	7.00 ± 0.00	$6.58^{a} \pm 0.31$			
	30	5.33 ± 0.33	6.33 ± 0.33	6.67 ± 0.33	6.33 ± 0.33	$6.17^a \pm 0.33$	0.0020	<.0001	0.4481
Colour -	60	4.33 ± 0.33	5.66 ± 0.33	5.67 ± 0.33	5.66 ± 0.33	$5.33^{b} \pm 0.33$	0.0020		
Coloui	Mean	$5.33^{b} \pm 0.41$	$6.11^a \pm 0.33$	$6.33^a \pm 0.33$	$6.33^a \pm 0.22$				
	0	6.66 ± 0.33	6.33 ± 0.33	6.33 ± 0.33	6.00 ± 0.57	$6.33^{a} \pm 0.39$			
	30	6.33 ± 0.33	6.00 ± 0.57	5.33 ± 0.33	5.66 ± 0.67	$5.83^{ab} \pm 0.48$	0.1724	0.0125	0.9595
Flavour -	60	6.00 ± 0.58	5.00 ± 0.57	5.00 ± 0.57	4.67 ± 0.67	$5.16^{b} \pm 0.60$	0.1724	0.0125	0.9393
Tavoui	Mean	$6.33^a \pm 0.41$	$5.77^{a} \pm 0.49$	$5.55^a \pm 0.41$	$5.44^{a} \pm 0.64$				
	0	6.67 ± 0.33	6.67 ± 0.33	6.66 ± 0.33	7.00 ± 0.00	$6.75^{a} \pm 0.25$		<.0001	0.9282
Tenderness	30	5.33 ± 0.33	6.00 ± 0.00	6.00 ± 0.00	6.17 ± 0.44	$5.87^{b} \pm 0.19$			
	60	4.66 ± 0.33	5.33 ± 0.33	5.00 ± 0.58	5.33 ± 0.33	$5.08^{\circ} \pm 0.39$	0.1074		0.9282
	Mean	$5.55^{b} \pm 0.33$	$6.00^{ab} \pm 0.22$	$5.89^{ab} \pm 0.30$	$6.17^{a} \pm 0.26$				
	0	6.33 ± 0.33	6.67 ± 0.33	6.66 ± 0.33	7.00 ± 0.00	$6.66^{a} \pm 0.25$			
Juiciness	30	5.66 ± 0.67	5.33 ± 0.33	6.33 ± 0.33	6.33 ± 0.33	$5.92^{b} \pm 0.42$	0.0656	< 0001	0.2564
	60	5.66 ± 0.33	4.33 ± 0.33	5.33 ± 0.33	5.33 ± 0.33	$5.16^{\circ} \pm 0.33$	0.0656	<.0001	0.3564
	Mean	$5.88^{ab} \pm 0.44$	$5.44^{b} \pm 0.33$	$6.11^{a} \pm 0.33$	$6.22^{a} \pm 0.22$				
Overall	0	5.33 ± 0.33	5.33 ± 0.67	5.66 ± 0.33	5.67 ± 0.33	$5.50^{a} \pm 0.42$			
acceptability	30	4.33 ± 0.33	4.67 ± 0.33	5.33 ± 0.33	5.17 ± 0.44	$4.87^{b} \pm 0.36$	0.2357	0.0018	0.0662
	60	4.00 ± 0.57	4.33 ± 0.33	4.33 ± 0.33	4.67 ± 0.33	$4.33^{b} \pm 0.39$			0.9663
	Mean	$4.55^a \pm 0.41$	$4.77^{a}\pm0.44$	5.11a ±0.33	$5.16^{a}\pm0.37$		=		

Mean values with different superscript in same row for the treatment and in same column for the days interval varies significantly at p < 0.05. T_0 =Control group, T_1 = 1 KGy irradiated group, T_2 = 2 KGy irradiated group T_3 = 3 KGy irradiated group, DI=Days of Intervals, T= Treatment, T*DI=Interaction of Treatment and Days of Intervals.

Proximate analysis

From Table 2 DM, CP, EE and Ash content at different treatments were found 28.66 to 31.31, 22.84 to 22.90, 1.28 to 2.65 and 1.32 to 1.09 %, respectively. The range of different days of interval DM, CP, EE and Ash content were found 28.69 to 31.68, 24.41 to 21.09, 2.66 to 1.73 and 1.12 to 1.30, respectively. The result showed that increasing irradiation dose increased the DM content significantly (p<0.05) as a result the shelf life of meat increased. DM content also increased with storage time. Similar results also found by Al-Bachir and Zeinou (2014), Konieczny et al. (2007) and Fallah et al. (2010). Nitrogen content in turkey meat is significantly (p<0.05) increased with irradiation doses. Storage period also significantly (p<0.05) decreased of CP content. Irradiated treated samples had significantly (p<0.05) higher amounts of EE in comparable with the control group. This trend was similar to the study revealed by Al-Bachir and Zeinou (2014). EE was significantly increased with increasing level of treatment but decreased with storage time. Ash was significantly decreased with increasing level of treatment but increased with storage period. Similar results also found

(Al-Bachir and Zeinou, 2014) that Ash content of meat decreased with increasing irradiation dose. There was positive and significant interaction between treatments and days of interval for DM and EE (Table 2).

Physicochemical and biochemical properties

Raw pH

From Table 3 shows the range of different treatments of pH and cooking loss score was 6.00 to 5.97 and, 20.07 to 24.98%. The rage of different days of interval pH and cooking loss score was 6.41 to 5.69 and 23.47 to 22.31%, respectively. The pH value slightly decreased with increasing irradiation doses. The effect of irradiation decreased raw pH values in the irradiated samples in comparable with controlled (Table 3). The data showed a slight decrease in the raw pH values and increased acidity values for all samples along with storage time during the 60 days of storage as a result of the increase of free fatty acids due to rancidity. A similar result also found by (Aftab et al., 2015) in irradiated broiler meat, pH was slightly decreased as the dose increased with the storage time. No statistically significant differences were found of pH in nonirradiated and irradiated group. Only control group apparently showed slightly higher of pH level. The present findings were more similar with Kim *et al.* (2012) where they found that pH value of samples were not significantly influenced by irradiation. Modi *et al.* (2008) also found that the lack of change in pH reflects that there were not enough protein breakdowns by

irradiation to elicit increased pH. There were statistically significant (p<0.05) differences of pH in storage periods. The results were in agreement with Morales-delanuez *et al.* (2009) findings who reported that the increase in fat values in irradiated samples and during storage decreased in pH values.

Table 2. Proximate composition (mean ± SE) in irradiated indigenous chicken meat samples compared at different storage period

Parameters	DI		Mean ± SE	Level of significance					
		T ₀	T_1	T ₂	T ₃	<u>-</u>	Т	DI	T*DI
DM (%)	0	27.39 ± 0.06	28.08 ± 0.06	29.16 ± 0.03	30.14 ± 0.05	$28.69^{b} \pm 0.05$			
	30	28.28 ± 0.65	27.54 ± 0.19	28.60 ± 0.28	29.88 ± 0.11	$28.57^{b} \pm 0.30$	- 0001	<.0001	0.0027
	60	30.32 ± 0.10	30.77 ± 0.12	31.75 ± 0.10	33.90 ± 0.12	$31.68^a \pm 0.11$	<.0001		
	Mean	$28.66^{\circ} \pm 0.27$	$28.79^{\circ} \pm 0.12$	$29.84^{b} \pm 0.14$	$31.31^a \pm 0.09$		_		
CP (%)	0	24.38 ± 0.11	24.41 ± 0.12	24.40 ± 0.13	24.46 ± 0.18	$24.41^a \pm 0.14$		<.0001	0.9935
	30	23.12 ± 0.02	23.10 ± 0.01	23.12 ± 0.02	23.12 ± 0.03	$23.12^{b} \pm 0.04$	0.8540		
	60	21.04 ± 0.01	21.14 ± 0.01	21.09 ± 0.05	21.13 ± 0.06	$21.09^{c} \pm 0.03$			
	Mean	$22.84^{a} \pm 0.05$	$22.88^a \pm 0.04$	$22.87^a \pm 0.07$	$22.90^a \pm 0.09$				
	0	1.09 ± 0.04	1.73 ± 0.10	1.97 ± 0.01	2.14 ± 0.05	$2.66^a \pm 0.05$			
EE (0/)	30	1.27 ± 0.03	2.19 ± 0.09	2.45 ± 0.05	2.66 ± 0.08	$2.14^{b} \pm 0.06$	< 0001	<.0001	<.0001
EE (%)	60	1.49 ± 0.02	2.94 ± 0.04	3.07 ± 0.05	3.15 ± 0.02	$1.73^{\circ} \pm 0.03$	<.0001		
	Mean	$1.28^{d} \pm 0.03$	$2.29^{\circ} \pm 0.08$	$2.49^{b} \pm 0.04$	$2.65^{a} \pm 0.05$		_		
Ash (%)	0	1.23 ± 0.02	1.16 ± 0.01	1.03 ± 0.02	1.05 ± 0.02	$1.12^{c} \pm 0.02$			
	30	1.31 ± 0.02	1.23 ± 0.02	1.12 ± 0.02	1.08 ± 0.03	$1.18^{b} \pm 0.02$	<.0001	<.0001	0.1726
	60	1.42 ± 0.03	1.37 ± 0.01	1.26 ± 0.01	1.15 ± 0.02	$1.30^{a} \pm 0.01$			0.1720
	Mean	$1.32^{a} \pm 0.02$	$1.25^{b} \pm 0.01$	$1.14^{c} \pm 0.02$	$1.09^{d} \pm 0.02$				

Mean values with different superscript in same row for the treatment and in same column for the days interval varies significantly at p < 0.05. T_0 =Control group, T_1 = 1 KGy irradiated group, T_2 = 2 KGy irradiated group T_3 = 3 KGy irradiated group, DI=Days of Intervals, T= Treatment, T*DI=Interaction of Treatment and Days of Intervals.

Table 3. Physicochemical and bio-chemical properties (mean \pm SE) in irradiated indigenous chicken meat samples compared at different storage period

Parameters	DI	Treatments (T) Mean \pm S				Mean ± SE	Level of significance		
		T ₀	T ₁	T_2	T ₃	•	T	DI	T*DI
	0	6.26 ± 0.16	6.42 ± 0.01	6.46 ± 0.01	6.52 ± 0.02	$6.41^a \pm 0.05$			
pН	30	5.97 ± 0.09	5.80 ± 0.01	5.79 ± 0.01	5.77 ± 0.01	$5.83^{b} \pm 0.03$	0.0504	<.0001	0.0061
	60	5.78 ± 0.03	5.67 ± 0.01	5.67 ± 0.01	5.62 ± 0.01	$5.69^{\circ} \pm 0.01$	0.8594		
	Mean	$6.00^{a} \pm 0.18$	$5.97^{a} \pm 0.01$	$5.97^{a} \pm 0.01$	$5.97^{a} \pm 0.01$		=		
	0	21.16 ± 0.53	23.55 ± 0.02	24.32 ± 0.05	24.87 ± 0.04	$23.47^a \pm 0.16$			
Cooking Loss	30	20.16 ± 0.47	22.70 ± 0.13	23.54 ± 0.07	25.06 ± 0.05	$22.86^{b} \pm 0.18$	< 0001	<.0001	0.0014
Cooking Loss	60	18.90 ± 0.26	22.48 ± 0.05	22.84 ± 0.06	25.01 ± 0.02	$22.31^{\circ} \pm 0.10$	<.0001		
(%)	Mean	$20.07^d \pm 0.42$	$22.91^{\circ} \pm 0.07$	$23.57^{b} \pm 0.06$	$24.98^a \pm 0.04$		=		
	0	0.33 ± 0.03	0.43 ± 0.03	0.49 ± 0.01	0.56 ± 0.02	$0.45^{\circ} \pm 0.03$			
FFA (%)	30	0.79 ± 0.06	0.81 ± 0.01	0.87 ± 0.01	0.87 ± 0.02	$0.84^{b} \pm 0.03$	<.0001	<.0001	<.0001
	60	0.87 ± 0.04	0.87 ± 0.02	1.51 ± 0.03	2.11 ± 0.02	$1.34^a \pm 0.03$	<.0001		
	Mean	$0.67^{c} \pm 0.04$	$0.70^{\circ} \pm 0.02$	$0.96^{b} \pm 0.02$	$1.18^a \pm 0.02$				
	0	0.83 ± 0.02	0.83 ± 0.01	0.89 ± 0.02	1.30 ± 0.02	$0.96^{c} \pm 0.02$			
POV	30	0.88 ± 0.01	1.01 ± 0.02	1.06 ± 0.01	1.50 ± 0.01	$1.11^{b} \pm 0.01$	<.0001	<.0001	0.0347
(meq/kg)	60	1.07 ± 0.08	1.04 ± 0.01	1.06 ± 0.01	1.52 ± 0.01	$1.17^a \pm 0.03$	<.0001		
	Mean	$0.93^{\circ} \pm 0.03$	$0.96^{bc} \pm 0.01$	$1.01^{b} \pm 0.01$	$1.44^{a} \pm 0.01$		=		
TBARS	0	0.06 ± 0.01	0.16 ± 0.01	0.19 ± 0.01	0.26 ± 0.01	$0.17^{c} \pm 0.01$			
	30	0.18 ± 0.01	0.41 ± 0.01	0.50 ± 0.02	0.62 ± 0.02	$0.43^{b} \pm 0.02$	<.0001	<.0001	<.0001
(mg- MDA/kg)	60	0.21 ± 0.01	0.57 ± 0.01	0.86 ± 0.03	1.14 ± 0.02	$0.69^{a} \pm 0.02$			<.0001
MIDA/Kg)	Mean	$0.15^{d} \pm 0.01$	$0.38^{c} \pm 0.01$	$0.52^{b} \pm 0.02$	$0.67^a \pm 0.02$	1 1 1 1			

Mean values with different superscript in same row for the treatment and in same column for the days interval varies significantly at p < 0.05. T_0 =Control group, T_1 = 1 KGy irradiated group, T_2 = 2 KGy irradiated group T_3 = 3 KGy irradiated group, DI=Days of Intervals, T= Treatment, T*DI=Interaction of Treatment and Days of Intervals

Statistically significant (p<0.05) changes were found of cooking loss in non-irradiated and irradiated groups. Cooking loss was gradually increased with increasing irradiation dose. Irradiation, as well as storage time

decreased muscle fiber that was the cause of increased cooking losses. Increase in cooking loss of irradiated meat samples could be due to the degradation of myofibrillar and structural proteins were found by Sweetie *et al.* (2015) in irradiated meat samples which are similar with the present study.

Biochemical properties

Table 3 shows the range of different treatments for FFA, PV and TBARS were 0.67 to 1.18, 0.93 to 1.14, and 0.15 to 0.67%, respectively. The range values of different days of intervals for FFA, POV and TBARS were 0.45 to 0.1.34, 0.96 to 1.17 and 0.17 to 0.69%. FFA value was significantly (p< 0.05) increased with irradiation level as well as longer storage time. Similarly Quattara *et al.* (2002) showed that gamma irradiation increased lipid oxidation in ground beef samples. In general terms, irradiation accelerates the lipid oxidation process, which is highly significant in foods with a high content of fats and much unsaturated fatty acids, in which numerous free radicals are formed due to this oxidation (O'Bryan *et al.*, 2008).

POV value was significantly (p<0.05) increased with irradiation level as well as with storage time. Chengliang *et al.* (2017) and Al-Bachir and Zeinou (2009) reported that an increase in oxidation activity and lipid per oxidation as a result of both radiation level of treatment and storage time on meat and meat products which was similar with the present findings. TBARS value was significantly (p<0.05) increased with irradiation level of as well as with storage time. Kim *et al.* (2012) found that TBARs increased significantly with storage time which is in agreement with the present study. There was positive and significant interaction between treatments and days of intervals among all treatments for all biochemical parameters (Table 3).

Microbiological assessment

From Table 4 shows the range of TVC, TCC and TYMC among different treatments was 4.81 to 3.22, 1.66 to 0.75 and 1.75 to 0.74, respectively. The range of TVC,

TCC and TYMC among different days interval was 3.81 to 4.26, 1.10 to 1.32 and 0.94 to 1.28, respectively. The results clearly showed that TVC was decreased significantly (p<0.05) with higher irradiation doses among all treatments group but decreased with storage period (Table 4). T₃ showed significantly lower bacteria than other treatments group. Similar results were found by Henriques et al. (2013). The low radiation doses can be efficiently used to control pathogens in chicken meat which is in accordance with Torgby et al. (2014). The present results clearly showed that TCC was decreased significantly (p<0.05) with higher irradiation doses among all treatments group and also increased with storage period. Similarly, Marta et al. (2016) found that cobalt-60 gamma irradiation process was effective in eliminating E. coli and found that lowest dose is enough to abolish this enter pathogen from the evaluated samples. Vereschako et al. (2016) proved that reduction in E. coli concentration has a linear relationship by the radiation doses. Non-irradiated group showed higher level of TYMC than irradiated groups.

The results clearly showed that TYMC was decreased significantly (p<0.05) with higher irradiation doses among all treatments group. During radiation, DNA molecules undergo swelling and break alongside the chain, preventing them from functioning normally. Fallah et al. (2010b) reported that the low doses irradiation reduced the initial counts of TYMC, while high doses were found below the detection levels of TYMC during 6 days of storage. There was positive and significant interaction between treatments and days of interval for TVC and TYMC (Table 4). Irradiation significantly improved the microbiological quality of aerobically packaged ready-to-cook (RTC). Iranian barbecued chicken by reducing the microbial floras without undesirable and detrimental effects on the sensory acceptability (Fallah et al., 2010b) which is in accordance with the present study except flavor.

Table 4. Effect of different doses of irradiation on microbial population (mean \pm SE) of indigenous chicken meat samples compared at different storage period

Parameters	DI	Treatments (T)	Mean ± SE	Level of significance					
		T_0	T_1	T_2	T ₃	_	T	DI	T*DI
	0	4.61 ± 0.16	3.88 ± 0.02	3.68 ± 0.04	3.08 ± 0.05	$3.81^{b} \pm 0.07$			
TVC	30	4.78 ± 0.11	3.67 ± 0.06	3.02 ± 0.01	3.03 ± 0.01	$3.62^{c} \pm 0.05$	<.0001	<.0001	<.0001
(log CFU/g)	60	5.02 ± 0.01	4.63 ± 0.03	3.83 ± 0.08	3.58 ± 0.04	$4.26^a \pm 0.04$			
	Mean	$4.81^{a} \pm 0.09$	$4.06^{b} \pm 0.04$	$3.51^{\circ} \pm 0.04$	$3.22^{d} \pm 0.03$		_		
	0	1.56 ± 0.05	0.99 ± 0.01	0.88 ± 0.02	0.58 ± 0.03	$1.01^{\circ} \pm 0.03$	<.0001	<.0001	
TCC	30	1.63 ± 0.04	1.07 ± 0.03	0.97 ± 0.01	0.71 ± 0.04	$1.09^{b} \pm 0.03$			0.2241
(log CFU/g)	60	1.79 ± 0.03	1.27 ± 0.03	1.25 ± 0.01	0.97 ± 0.04	$1.32^a \pm 0.03$			0.2241
	Mean	$1.66^{a} \pm 0.04$	$1.11^{b} \pm 0.02$	$1.03^{\circ} \pm 0.01$	$0.75^{d} \pm 0.04$				
	0	1.63 ± 0.03	0.91 ± 0.01	0.62 ± 0.04	0.60 ± 0.01	$0.94^{\circ} \pm 0.03$			
TYMC (log CFU/g)	30	1.74 ± 0.03	0.97 ± 0.02	0.79 ± 0.01	0.68 ± 0.02	$1.04^{b} \pm 0.02$	/ 0001	<.0001	0.0153
	60	1.88 ± 0.02	1.27 ± 0.03	1.00 ± 0.01	0.95 ± 0.02	$1.28^a \pm 0.02$			0.0133
	Mean	$1.75^a \pm 0.03$	$1.05^{b} \pm 0.02$	$0.80^{\circ} \pm 0.02$	$0.74^{d} \pm 0.03$	•			

Mean values with different superscript in same row for the treatment and in same column for the days interval varies significantly at p < 0.05. T_0 =Control group, T_1 = 1 KGy irradiated group, T_2 = 2 KGy irradiated group T_3 = 3 KGy irradiated group, DI=Days of Intervals, T= Treatment, T*DI=Interaction of Treatment and Days of Intervals.

Conclusion

The study revealed that gamma irradiation had significant effect on indigenous chicken meat quality and safety. Among the treatments, irradiation dose 2.0 kGy showed the best results in terms of sensory evaluation, biochemical analysis and microbial assessment and the shelf life extension of indigenous chicken meat. It may be concluded that gamma irradiation will enable to deliver the larger amount of high quality indigenous chicken meat with extended shelf life.

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