IN VITRO SCREENING OF FUNGICIDES AND PLANT EXTRACTS AGAINST PATHOGENIC FUNGI ASSOCIATED WITH INFECTED FRUITS OF CARICA PAPAYA L.

REZUANA BINTE HELAL¹ AND SHAMIM SHAMSI*

Department of Botany, University of Dhaka, Dhaka - 1000, Bangladesh

ABSTRACT

Five fungicides *viz.*, Autostin 50 WDG, Dithane M 45, Greengel 72 WP, Ridomil Gold MZ 68 WG and Tilt 250 EC exclusively were tested against three pathogenic fungi *viz.*, *Colletotrichum gloeosporioides* (Penz.) Sacc., *Fusarium nivale* (Fr.) Ces. and *Fusarium* sp. associated with *Carica papaya* L. following poisoned food technique. At 100 ppm Autostin showed complete growth inhibition of *C. gloeosporioides*, *F. nivale* and *Fusarium* sp., while Tilt 250 EC showed complete growth inhibition of *C. gloeosporioides* at the same concentration. These fungicides at 400 and 500 ppm showed complete growth inhibition of *F. nivale*. Tilt 250 EC showed complete growth inhibition of *Fusarium* sp. at 300, 400 and 500 ppm. Eight plants extract, *viz.*, *Adhatoda vasica*, *Azadirachta indica*, *Curcuma longa*, *Ocimum sanctum*, *Lantana camara*, *Tagetes erecta*, *Thuja orientalis* and *Vitex negundo* were also tested for their efficacy at 5, 10, 15 and 20% concentrations against these pathogenic fungi. Out of the eight plant extracts, *Ocimum sanctum* showed highest (88.57%) radial growth inhibition of *C. gloeosporioides* at 20% concentration. *Curcuma longa* showed highest (88.55%) radial growth inhibition of *F. nivale* and *Adhatoda vesica* showed highest (80.02%) radial growth inhibition of *Fusarium* sp. at the same concentrations. The present investigation suggests Autostin 50 WDG and Tilt 250 EC as best inhibiting chemical fungicides for *C. gloeosporioides*, *F. nivale* and *Fusarium* sp. Extracts of *O. sanctum*, *C. longa* and *Adhatoda vesica* were found to be superior to other plant extracts tested.

Keywords: In vitro screening, Fungicides, Plant extracts, Pathogenic fungi, Carica papaya L.

INTRODUCTION

Papaya (Carica papaya L.) plant is known to uterotonic, nephroprotective, inflamatory and anti-tumor properties (Milind and Gurditta 2011). Papaya is a good source of nutrient and vitamin A and C (Duke 1984). Glucose is the main sugar in papaya but sucrose content increases during ripening and can reach up to 80% of the total sugars (Parni and Verma 2014). The availability of the fruit is reduced due to high level of post-harvest loss (Mondal et al. 1995). Sawant and Gawri (2011) reported that fungal infection reduced the nutritional value of the fruits. Approximately 41 % postharvest loss of papaya was recorded in Bangladesh (Hassan 2010). Various fungi cause rots in fruits of papaya.

Due to environmental concerns of controlling fungal disease by toxic chemicals, researchers have focused their efforts on developing alternative methods of controlling fungal diseases (Baker 1987, Cook 1993 and Heydari and Pessarakli 2010). One is use of biological antagonists and plant extracts. Plant extracts can be successfully exploited in modern agriculture which has recently attracted attention of several workers. Plant constituents have been reported to be successful fungitoxicants because of low phytotoxicity, systemicity, easy biodegradability and favorable effects for the growth of the host (Fawcett and Spencer 1970 and Panday et al. 1983). Extracts obtained from many plants have recently been studied for their antifungal

¹ The experiment was conducted in the laboratory of Mycology and Plant Pathology, Department of Botany, University of Dhaka, Dhaka – 1000, Bangladesh

activities (Monoharachary and Reddy 1978, Ahmed and Sultan 1984, Miah *et al.* 1990, Hosen *et al.* 2016).

The present investigation was undertaken to evaluate the toxicity of some chemical and plant extracts against test pathogens.

MATERIALS AND METHODS

Infected ripe fruits of Carica papaya L. having characteristic symptoms were collected from five different markets of Dhaka City, during April and May, 2016 in separate sterile polyethylene bags, labeled properly and brought to the laboratory. The associated fungi were isolated following Tissue planting method on PDA medium (Anonymous 1968). Identification of the fungi were done following standard literature. Pathogenecity of the isolated fungi were done following fruit inoculation technique. Among the isolated fungi Colletotrichum gloeosporioides, Fusarium nivale and Fusarium sp. were found to be pathogenic to papaya fruit (Helal 2017). Five fungicides with different active ingredients, viz., Autostin WDG, Dithane M 45, Greengel 72 WP, Ridomil Gold MZ 68 WG and Tilt 250 EC were collected from the Krishi Upokoron Biponi Kendro at Khamarbari, Farmgate, Dhaka (Table 1). Of the five fungicides used in the present investigation, Autostin 50 WDG and Dithane M 45 are systemic while Tilt 250 EC and Greengel 72 WP are protectant fungicides and Ridomil gold MZ 68 WG is both systemic and protectant fungicide. For each fungicide, a stock solution having the concentrations of 10,000 ppm was prepared. The calculated amount of the stock solution of a fungicide was supplemented with sterilized PDA medium to get the conc. of 100, 200, 300, 400 and 500 ppm. In the control set required amount of sterile water instead of fungicide solution was added to the PDA medium.

A total of eight plants were selected for evaluating their effect on the radial growth of C.

gloeosporioides, F. nivale and Fusarium sp. follwing Rai and Bashar (1991). The desired parts of each plant were thoroughly washed in tap water, air dried and then used for fresh extract preparation. The pulverized mass of a plant part was squeezed through four folds of fine cloth and the extracts were centrifuged at 3000 rpm for 20 minutes to remove particulate matter. The supernatants were filtered through Whatman filter paper and the filtrate was collected in 250 mL Erlenmeyer flasks. In this method, the requisite amount of the filtrate of each plant extract was mixed with PDA medium to get 5, 10, 15 and 20% concentratios.

Both the sets i.e. medium with fungicides or plant extracts thus prepared were poured into sterilized Petri plates and was allowed to solidify. Each Petri plate was inoculated centrally with a 5 mm agar disc cut from the margin of actively growing 7 day culture of the test pathogens. In the control set, a Petri plate containing PDA medium with the requisite amount of distilled water instead of a fungicide or a plant extract was added then agar disc with mycelia of each test pathogen was inoculated in the centre of Petri plate. Three replications were maintained for both the experiments and control sets. The inoculated Petri plates were incubated at 25±2°C. The radial growth of the colonies of the test pathogens was measured after 5 days of incubation.

The fungitoxicity of the fungicides and plant parts extracts in terms of percentage inhibition of mycelial growth was calculated by using the following formula:

$$I = \frac{C - T}{C} \times 100$$

where, I= Per cent growth inhibition, C= Growth in control, T= Growth in treatment.

The results were statistically analyzed by "t" test following Steel and Torrie (1960).

RESULTS AND DISCUSSION

Three pathogenic fungi Colletotrichum gloeosporioides (Penz.) Sacc., Fusarium nivale (Fr.) Ces., and Fusarium sp. were isolated from infected papaya fruits. Five fungicides viz., Autostin 50 WDG, Dithane M 45, Greengel 72 WP, Ridomil Gold MZ 68 WG and Tilt 250 EC were selected to evaluate in vitro efficacy at 100, 200, 300, 400 and 500 ppm concentrations against three pathogenic fungi.

ppm and 500 ppm concentrations. Greengel 72 WP and Ridomil 68 WG showed complete growth inhibition of the fungus at 500 ppm respectively. The toxicity of these fungicides against *C. gloeosporioides* at 100 ppm in descending order was Autostin 50 WDG=Tilt 250 EC> Greengel 72 WP> Ridomil Gold MZ 68 WG> Dithane M 45 (Table 1).

On the other hand, complete inhibition of the growth of Fusarium nivale was observed with

Table 1. Per cent inhibition of radial growth of *Colletotrichum gloeosporioides* at different concentrations of fungicides

Name of fungicides	% inhibition of radial growth at different concentrations (ppm)					
-	100	200	300	400	500	
Autostin 50 WDG	100 ^a	100 ^a	100 ^a	100 ^a	100 ^a	
Dithane M-45	51.21 ^b	56.91 ^b	67.47 ^a	100^{a}	100^{a}	
Greengel 72 WP	60.97 ^b	66.67 ^a	68.29 ^a	78.04^{a}	100^{a}	
Ridomil Gold MZ 68 WG	55.28 ^b	60.97 ^b	70.73 ^a	81.30 ^a	100 ^a	
Tilt 250 EC	100 ^a	100^{a}	100^{a}	100^{a}	100^{a}	

a, b and c indicate significance of 't' value at p = 0.001 and 0.01 and 0.05, respectively.

In a row, figures with same letter do not differ significantly, whereas figures with dissimilar letter differ significantly (as per DMRT), NS = Not significant. Efficiency gradient of fungicides against *Colletotrichum gloeosporioides* at 100 ppm concentration: Autostin 50 WDG=Tilt 250 EC> Greengel 72 WP> Ridomil Gold MZ 68 WG> Dithane M 45.

In the present investigation, complete inhibition of the radial growth of *C. gloeosporioides* was observed with Autostin 50 WDG and Tilt 250 EC at all the concentrations used. Among the fungicides, Dithane M 45 showed complete growth inhibition of *C. gloeosporioides* at 400

Autostin 50 WDG at all the concentrations used. Tilt 250 EC showed 100% growth inhibition of the fungus at 400 ppm and 500 ppm concentrations. Ridomil gold MZ 68 WG, Dithane M 45 and Greengel 72 WP showed highest 78.02%, 71.42% and 70.32% growth

Table 2. Per cent inhibition of radial growth of *Fusarium nivale* at different concentrations of fungicides

Name of fungicides	% innibition of radial growth at different concentrations (ppm)					
-	100	200	300	400	500	
Autostin 50 WDG	100 ^a	100 ^a	100 ^a	100 ^a	100 ^a	
Dithane M-45	58.24 ^b	60.43 ^a	64.83 ^a	69.23 ^a	71.42 ^a	
Greengel 72 WP	51.64 ^b	56.04 ^b	58.24 ^a	64.83 ^a	70.32^{a}	
Ridomil Gold MZ 68 WG	59.34 ^b	62.63 ^a	63.29 ^a	67.03 ^a	78.02 ^a	
Tilt 250 EC	86.90 ^a	93.78 ^a	98.20 ^a	100 ^a	100 ^a	

a, b and c indicate significance of 't' value at p = 0.001, 0.01 and 0.05, respectively.

In a row, figures with same letter do not differ significantly, whereas figures with dissimilar letter differ significantly (as per DMRT), NS = Not significant. Efficiency gradient of fungicides against *Fusarium nivale* at 100 ppm concentration: Autostin 50 WDG =Tilt 250 EC>Ridomil Gold MZ 68 WG> Dithane M-45>Greengel 72 WP.

inhibition of *Fusarium nivale*, respectively at 500 ppm. The toxicity of these fungicides against *Fusarium nivale* at 100 ppm in descending order was Autostin 50 WDG> Tilt 250 EC> Ridomil Gold MZ 68 WG> Dithane M 45> Greengel 72 WP (Table 2).

Complete inhibition of the growth of *Fusarium* sp. was observed with Autostin 50 WDG, at all concentrations used. Tilt 250 EC showed 100% growth inhibition of the test fungi at 300 ppm, 400 ppm and 500 ppm concentrations. Dithane M 45 showed highest 99.50% growth inhibition of *Fusarium* sp. at the 500 ppm concentration. At 500 ppm concentration Ridomil 68 WG and Greengel 72 WP showed complete inhibition of *Fusarium* sp. The toxicity of these fungicides against *Fusarium* sp. at 100 ppm in descending order was Autostin 50 WDG> Tilt 250 EC> Dithane M-45> Greengel 72 WP> Ridomil Gold MZ 68 WG (Table 3).

due to selection of different test pathogens. Efficacy gradients observed in the present study expressed that Autostin 50 WDG, Tilt 250 EC, Dithane M-45, Greengel 72 WP and Ridomil 68 WG were the best inhibiting agent against the in vitro growth of the test pathogens. In contrast to the present study, Chakraborty et al. (2009) reported that, at 0.5% dose, Bavistin happened to be the most efficient with the highest inhibition (83.7%) of growth of Fusarium solani causing wilt of brinjal in vitro. Bashar (1992) while reporting the results of laboratory evaluation of some pesticides on F. oxysporum f. sp. Ciceri causing wilt of chick pea found that Bavistin checked the growth of the pathogen completely at 100 ppm. Bashar also noted that Dithane M 45

Table 3. Per cent inhibition of radial growth of Fusarium sp. at different concentrations of fungicides

Name of fungicides	% inhibition of radial growth at different concentrations (ppm)					
_	100	200	300	400	500	
Autostin 50 WDG	100 ^a	100 ^a	100 ^a	100 ^a	100 ^a	
Dithane M-45	78.30^{a}	84.90 ^a	86.79 ^a	90.00^{a}	99.5 ^a	
Greengel 72 WP	75.47 ^a	81.13 ^a	82.07 ^a	82.07 ^a	100^{a}	
Ridomil Gold MZ 68 WG	46.22 ^b	74.52 ^a	81.13 ^a	98.70 ^a	100 ^a	
Tilt 250 EC	95.80 ^a	99.90 ^a	100 ^a	100 ^a	100 ^a	

a, b and c indicate significance of 't' value at p = 0.001, 0.01 and 0.05, respectively. In a row, figures with same letter do not differ significantly, whereas figures with dissimilar letter differ significantly (as per DMRT), NS = Not significant. Efficiency gradient of fungicides against *Fusarium* sp. at 100 ppm concentration: Autostin 50 WDG> Tilt 250 EC> Dithane M-45>Greengel 72 WP>Ridomil Gold MZ 68 WG

Efficacy of various fungicides against the three test fungi pathogens indicates that Autostin 50 WDG, Tilt 250 EC, Dithane M-45, Greengel 72 WP and Ridomil 68 WG showed promising results. The same fungicides also showed different effects on three test pathogens in the present investigation. This variation might be

failed to check the growth of the pathogen completely even at 3,000 ppm.

All the fungicides tested caused partial or complete inhibition of *C. gloeosporioides*, *F. nivale* and *Fusarium* sp. at 500 ppm concentration (Tables 1-3). Similar observations have also been reported for other fungicides by

Vishwakarma and Chaudhury (1992) and Bashar (1992). Singh and Singh (1970) observed that reaction of *Fusarium* spp. to fungicides varies from species to species and sometimes even from isolate to isolate of the same species. It was reported by Chowdhury *et al.* (2015) that Dithane, Ridomil and Bavistin at 400 and 500 ppm showed complete inhibition of radial growth of *F. moniliforme*.

Results of plant extracts on the radial growth of *Colletotrichum gloeosporioides*, *Fusarium nivale* and *Fusarium* sp. are presented in Tables 4-6. All the plant extracts showed varied degree

followed by Curcuma longa (84.00%),Azadirachta indica (82.79%), Adhatoda vasica (80.99%), Thuja orientalis (71.00%), Lantana camara (67.32%), Tagetes erecta (65.59%) and Vitex nigundo (42.77%). The inhibition of the with pathogen increases increase concentration of plant extracts in culture medium. The order of effectiveness against C. gloeosporioides at 20% concentration was Ocimum sanctum> Curcuma longa> Azadirachta indica> Adhatoda vasica> Thuja orientalis> Lantana camara> Tagetes erecta> Vitex negundo (Table 4).

Table 4. Effects of plant extracts on the radial growth of Colletotrichum gloeosporioides at different concentrations

Name of plants	% inhibition of radial growth of the pathogen at different concentrations (%)					
	5	10	15	20		
Adhatoda vasica	51.78 ^b	62.77 ^b	78.23 ^a	80.99 ^a		
Azadirachta indica	54.78 ^b	70.45 ^a	77.84 ^a	82.79 ^a		
Curcuma longa	50.00 ^b	52.50 ^b	67.50 ^a	84.00 ^a		
Lantana camara	40.34 ^c	48.99 ^b	56.20 ^b	67.32 ^b		
Ocimum sanctum	62.23 ^b	50.14 ^b	71.28 ^a	88.57 ^a		
Tagetes erecta	31.18 ^c	49.46 ^b	48.38 ^b	65.59 ^b		
Thuja orientalis	27.66 ^c	32.65 ^b	65.22 ^a	71.00^{a}		
Vitex nigundo	18.98 ^{NS}	36.70°	39.24 ^b	42.77 ^b		

a, b and c indicate significance of 't' value at p = 0.001, 0.01 and 0.05, respectively. In a row, figures with same letter do not differ significantly, whereas figures with dissimilar letter differ significantly (as per DMRT), NS = Not significant. Efficiency gradient of plant extracts against *Colletotrichum gloeosporioides* at 20% concentration: cimum sanctum> Curcuma longa> Azadirachta indica> Adhatoda vasica> Thuja orientalis> Lantana camara> Tagetes erecta> Vitex negundo.

of growth inhibition of the pathogens at different concentrations. Out of the eight plant extracts, *Ocimum sanctum* (88.57%) showed best results for inhibition of radial growth of *Colletotrichum gloeosporioides* at 20% concentration which was

Chowdhury et al. (2015) reported that ethanol extract of A. indica showed complete radial growth inhibition of at 10 and *T*. concentration whereas erecta showed inhibition complete 20% growth at concentrations. Prasad and Anamika (2015)

reported that extracts of *L. camara* were found most effective for the control of the *C. gloeosporioides*. Imtiaj *et al.* (2005) found that plant extracts such as *Curcuma longa*, *T. erecta* and *Zingeber officinale* were also highly effective against *C. gloeosporioides*.

Out of the eight plant extracts, *Curcuma longa* showed highest (88.55%) radial growth inhibition of *Fusarium nivale* at 20% concentration which was followed by *Ocimum sanctum* (87.65%), *Azadirachta indica* (75.64%), *Tagetes erecta* (69.09%), *Lantana camara* (57.78%), *Adhatoda vasica* (56.79%), *Vitex negundo* (49.03%) and *Thuja orientalis* (47.00%). The inhibition of the pathogen increases with the increase of the concentration of the plant extracts in culture medium (Table-5).

The order of effectiveness against Fusarium nivale at 20% concentration was Curcuma longa> Ocimum sanctum> Azadirachta indica> Tagetes erecta> Lantana camara> Adhatoda vasica> Vitex negundo> Thuja orientalis (Table 5). Out of the eight plant extracts, Azadirachta indica showed highest (90.19%)

radial growth inhibition of Fusarium sp. at 20% concentration which was followed by Adhatoda vasica (80.02%), Ocimum sanctum (79.89%), Thuja orientalis (69.34%), Lantana camara (67.99%), Cucurma longa (67.64), Tagetes erecta (65.93%) and Vitex negundo (58.75%). The inhibition of the pathogen increases with the increase of the concentration of the plant extracts in culture medium. The order of effectiveness against Fusarium sp. at 20% concentration was Azadirachta indica> Adhatoda vasica> Ocimum sanctum> Thuja orientalis> Lantana camara> Cucurma longa>Tagetes erecta> Vitex negundo (Table 6). In contrast to the present study, Bashar and Chakma (2014) reported that the plant extract of A. indica showed 62.03% growth inhibition of F. oxysporum at 20% concentration. The same plant extracts also showed different effects on different pathogens in the present investigation. This variation might be due to selection of different test pathogens. Singh et al. (1993) reported the effectiveness of aqueous extracts of A. indica in the control of disease development in banana. Plant

Table 5. Effects of plant extracts on the radial growth of Fusarium nivale at different concentrations

% inhibition of radial growth of the pathogen at different concentrations (%) Name of plants 5 10 15 20 Adhatoda vasica 32.78^{c} 45.66^b 45.66^b 56.79^b 40.45^c 55.67^b 65.54a 75.64^{a} Azadirachta indica Curcuma longa 69.44^b 72.22^a 79.00^{a} 88.55^a 56.44^b 57.78^b Lantana camara 26.90^{c} 30.33^{c} 38.27^c 67.90^b 70.37^{a} 87.65^a Ocimum sanctum Tagetes erecta 36.36^c 63.63^b 56.81^b 69.09^b 30.12^{NS} 45.50^c 47.00^{c} Thuja orientalis 35.22^c 18.75^{NS} 49.03^b Vitex negundo 31.25° 47.50^{c}

a, b and c indicate significance of 't' value at p = 0.001, 0.01 and 0.05, respectively. In a row, figures with same letter do not differ significantly, whereas figures with dissimilar letter differ significantly (as per DMRT), NS = Not significant. Efficiency gradient of plant extracts against *Fusarium nivale* at 20% concentration: *Curcuma longa> Ocimum sanctum> Azadirachta indica> Tagetes erecta> Lantana camara> Adhatoda vasica> Vitex negundo> Thuja orientalis*.

Table 6. Effects of plant extracts on the radial growth of Fusarium sp. at different concentrations

Name of plants	% inhibition of radial growth of the pathogen at different concentrations ($%)$					
	5	10	15	20		
Adhatoda vasica	66.50 ^b	69.45 ^b	75.78 ^a	80.02 ^a		
Azadirachta indica	58.92 ^b	75.27 ^b	82.25 ^a	90.19 ^a		
Cucurma longa	58.82 ^b	61.76 ^a	64.70 ^a	67.64 ^a		
Lantana camara	29.77°	34.46 ^b	58.92 ^a	67.99 ^a		
Ocimum sanctum	43.15°	53.68 ^b	64.21 ^b	79.89^{a}		
Tagetes erecta	24.57 ^b	34.74 ^b	54.91 ^a	65.93 ^a		
Thuja orientalis	32.14 ^c	45.67 ^b	56.88 ^b	69.34 ^a		
Vitex negundo	27.02 ^b	37.50 ^a	46.25 ^a	58.75 ^a		

a, b and c indicate significance of 't' value at p = 0.001, 0.01 and 0.05, respectively. In a row, figures with same letter do not differ significantly, whereas figures with dissimilar letter differ significantly (as per DMRT), NS = Not significant. Efficiency gradient of plant extracts against Fusarium sp. at 20% concentration: Azadirachta indica> Adhatoda vasica> Ocimum sanctum> Thuja orientalis> Lantana camara> Cucurma longa> Tagetes erecta> Vitex negundo

parts and their constituents of some higher plants have already been reported to be of successful nature of fungitoxicants, lesser phytotoxicity, systemicity, easy biodegradability and favourable effects of the growth of the host (Fawcett and Spencer 1970, Panday *et al.* 1983). Chakraborty *et al.* (2009) reported the efficacy of various cell free extracts of the plants against the growth inhibition of the pathogen. The effectiveness of extracts varied significantly with dosage, where 100% inhibition of the pathogen was achieved both with neem and garlic extracts.

ACKNOWLEDGEMENT

The authors wish to express gratitude to the Ministry of Science and Technology, People's Republic of Bangladesh for providing financial assistance through NST fellowship awarded to Rezwana Binte Helal. The authors also wish to thank the Centre for Advanced Research in Sciences (CARS), Dhaka University for providing support to the research during the tenure of the research (2016-2017).

REFERENCES

Ahmed, N. and K. Sultana. 1984. Fungitoxic effect of garlic on treatment of jute seed. Bangladesh J. Bot. **13**(2): 130-136.

Anonymous. 1968. *Plant Pathologist Pocket Book*. The Commonwealth Mycological Institute, England.pp. 267.

Baker, K. F. 1987. Evolving concepts of biological control of plant pathogens. Annu. Rev. *Phytopathol.* **25**: 67-85

Bashar, M.A. and B, Rai. 1991. Antifungal activity of extracts of some plant parts against *Fusarium oxysporum* f. sp. *ciceri. Bangladesh J. Bot.* **20**: 219-222.

Bashar, M.A. 1992. Laboratory evaluation of some pesticides on *Fusarium oxysporum* f. sp. *ciceri* causing wilt of chickpea. *Bangladesh J. Bot.* **21**(1): 157-159.

Bashar, M.A. and M. Chakma 2014. *In vitro* control of *Fusarium solani* and *F. oxysporum* the causative agent of brinjal wilt. *Dhaka Univ. J. Biol. Sci.* **23**(1): 53-60.

Chakraborty, M.R., N.C. Chatterjee and T.H. Quimio. 2009. Integrated management of fusarial wilt of eggplant (*Solanum melongena*) with soil solarization. *Micologia Aplicada International*.

Chowdhury, P., M..A. Bashar and S. Shamsi. 2015. *In vitro* evaluation of fungicides and plant extracts against pathogenic fungi of two rice varieties. *Bangladesh J. Bot.* **44**(2): 251-259.

Cook, R.J. 1993. Making greater use of microbial inoculants in agriculture. *Amrn. Rev. Phytopathol.* **31**: 53-80.

- Duke JA 1984.Borderline Herbs, CRS Press, Boca Roton Florida,USA.
- Fawcett, C.H. and D.M. Spencer. 1970. Plant chemotherapy with natural products. *Ann. Rev. Phytopathol.* **8**: 403-418.
- Hassan, M,K. 2010. A guide to postharvest handling of fruits and vegetables, Department of Horticulture. Bangladesh Agricultural University, Mymensingh.
- Helal, R.B. 2017. Mycoflora associated with papaya (Carica papaya L.) and their management. MS thesis. Department of Botany University of Dhaka. Bangladesh pp. xi+182
- Hosen, S., S Shamsi and M.A. Bashar. 2016. *In vitro* efficacy of fungicides and plant extracts on the growth of *Colletotrichum gloeosporioides* (Penz.) Penz. & Sacc. And *Sclerotium rolfsii* Sacc. the causal organisms of anthracnose and soft rot of jute. *Dhaka Univ. J. Bio. Sci.* 25(2): 195-199.
- Imtiaj, A., S.A. Rahman, S. Alam, R. Parvin, K. M., Farhana, S.B. Kim, T. S. Lee. 2005. Effect of Fungicides and Plant Extracts on the Conidial Germination of Colletotrichum gloeosporioides causing mango anthracnose. Mycobiology. 33(4): 200-205.
- Heydari, A. and M. Pessarakli, 2010. A review on biological control of fungal plant pathogens using microbial antagonists. *Journal of Biological Sciences* **10**(4): 203-290.
- Islam, M. A., S. Shamsi, S. Hosen and M. A. Bashar. 2017. In vitro effect of five plant extracts and five fungicides on Fusarium oxysporum Schlecht. and F. solani (Mart.) Sacc. causal agent of brinjal (Solanum melongena L.) wilt. Dhaka Univ. J. Bio. Sci. 26(1): 39-44.
- Miah, M.A., T. Ahmed, N. R. Sharma, A. Ali and S.A. Miah. 1990. Antifungal activity of some plant extracts. *Bangladesh J. Bot.* **19**: 5-10.

Milind, P. and Gurditta. 2011. Busketful benefits of papaya. *International Research Journal of Pharmacy*. **2**(7): 6-12.

- Mondal, M.F., M.A. Rahman and M.A.J. Pramanik. 1995. Effect of different post harvest treatments on physio chemical changes and shelf life of mango. *Bangladesh horticulture*. **23**(1&2): 1-5.
- Monoharachary, C. and M.M. Reddy. 1978. Efficacy of some leaf extracts on *Botryodiplodia theobromae* Pat. *Geobios*. **5**(4): 190-191.
- Panday, D.K., H. Chandra, N.N. Tripathi and Dixit 1983. Fungitoxicity of some higher plants with special reference to the synergistic activity among some fungitoxicants. *Phytopathol. Z.* **106**: 226-232.
- Parni, B. and Y. Verma. 2014. Biochemical properties in peel, pulp and seeds of *Carica papaya*. *Plant Archives* **14**(1): 565-568.
- Prasad, R,R. and Anamika. 2015. Effects of plant leaf extract against *Colletotrichum gloeosporioides* (Penz) Sac. Causing postharvest Disease of Papaya. *Journal of Agricultural Science* **7**(5): 195-198.
- Sawant, S.G. and D.U. Gawai. 2011. Effect of fungal infections on nutritional value of papaya fruits, *Curr. Bot.* 2(1): 43-44.
- Singh N and Singh RS 1970. Development of wilt causing species of *Fusarium* in fungicide treated soils. Indian Phytopath. **23**: 545-552.
- Singh HNP, Prasad MM and Sinha KK 1993. Efficacy of leaf extracts of some medicinal plants against disease development in banana. Letters in Applied Microbiology **17**: 269-271.
- Steel. R. G. D. and J. H. Torrie. 1960. *Principles and Procedures of Statistics*. MacGraw Hill Book Co., New York. pp. xvi + 481.
- Vishwakarma, S. N. and K. C. B. Chaudhury. 1992. Laboratory evaluation of some fungicides against some root rot disease pathogens of gram. *Phytopath*. **35**: 624-627.

(Received revised manuscript on 23 September 2018)