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- Short Communication

## COMPARATIVE STUDY OF NON POLAR AND POLAR SOLVENT EXTRACTS OF *GLOCHIDION VELUTINUM* WIGHT IN RESPECT OF ANTIOXIDANT AND ANTIBACTERIAL ACTION

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Antioxidant and antimicrobial activities of non-polar and polar extracts of *Glochidion velutinum* Wight have been studied. The phenolic content was determined by Folin-Ciocalteu reagent and colorimetric method was used to quantify flavonoids. Antioxidant activity was also measured by DPPH free radical scavenging assay and disk diffusion method was applied to test antimicrobial activity. Of the hexane, chloroform, ethanol and methanol extracts of *Glochidion velutinum*, higher total phenolic and flavonoid contents were found in methanol and ethanol extracts. Moreover, ethanol and methanol extracts also showed better antimicrobial and antioxidant activities than the other two extracts. The presence of various phytochemicals e.g. alkaloids, tannins, phenols etc in these extracts also substantiated the observed antioxidant and antimicrobial activities.

## Key words: *Glochidion velutinum*, Antioxidant, Antimicrobial, Phytochemicals, Phenolic content, Flavonoid

Plants have been the best source of remedies for curing a variety of diseases and have played key roles worldwide for maintenance of health. Natural products of higher plants are important sources of therapeutic agents. Therefore, many research groups are currently screening various plants for different biological activities (Mothana *et al.* 2010, Mulabagal *et al.* 2007, Leu *et al.* 2006).

*Glochidion velutinum* Wight (Family-Euphorbiaceae) is a small tree or large shrub upto 9 m with coraceous, pinnate venation leaves, yellow male flowers and globose and depressed fruits branches and leaves. This plant is locally known as matachhar and widely distributed in China, Pakistan, India, Nepal, and Bangladesh (Madhavachetty *et al.* 2008). Several triterpenoids, triterpene glycosides and alkaloids are known to be the constituents of this plant (Sandhya *et al.* 2010). The stem bark of the plant has been reported to be moderately bactericidal (Karuppusamy and Rajasekaran, 2009).

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Traditionally, this plant is used for treating cancer, diabetes, diarrhoea, inflammation and for the healing of wounds (Madhavachetty *et al.* 2008). Literature survey revealed that leaves of G. *velutinum* are effective against type 2 diabetes (Are *et al.* 2011). In addition, there are also reports in support of antioxidant and cytotoxic potential of this plant (Sandhya *et al.* 2011, Hasan *et al.* 2016). However, no study appears to have been conducted on antioxidant and antimicrobial activities of different extracts of the leaves of this plant, yet. The objective of this work was to compare the antimicrobial and antioxidant properties of four different extracts of the leaves of G. *velutinum*. A great interest in the antioxidant activity of plant extracts exists because of free radicals (e.g., reactive oxygen species) that can be responsible for several diseases, for example, heart disease, stroke, arteriosclerosis and cancer, as well as the aging process (Hoye *et al.* 2008).

The plant *Glochidion velutinum* was identified by the expert of Bangladesh National Herbarium, Mirpur, Dhaka where a voucher specimen bearing the accession number DACB-43211 has been deposited. The leaves of the plant were collected from the forest range of Rajendrapur, Gazipur, Dhaka, Bangladesh during the  $2^{nd}$  week of January 2015. The cleaned leaves were dried in shade for 7 days followed by drying in an oven at 40°C for 2 hours. The leaves were then ground to a coarse powder with the help of a mechanical grinder.

The dried powder (400gm) was taken in four porous bags and placed in the chamber of four soxhlet extractors. The powder was first defatted with n-hexane for 28 hours. The defatted powder was then extracted with chloroform, ethanol and methanol successively as before, each for 28 hours. The extracts thus obtained were separately collected and denoted as hexane extract (HGV), chloroform extract (CGV), methanol extract (MGV) and ethanol extract (EGV).

Preliminary phytochemical screenings of G. *velutinum* extracts were carried out using standard procedures as described by Trease and Evans (1978). All the extracts HGV, CGV, MGV and EGV revealed the presence of flavonoids, carbohydrate and alkaloids. The presence of tannins, steroids and saponins were observed only in CGV, MGV and EGV.

The antioxidant activity was determined by DPPH free radical assay by standard protocol (Braca *et al.* 2001). The scavenging of DPPH free radical is indicated by the deep violet color being turned pale yellow or colorless. Various concentrations (1.56 to 800  $\mu$ g/mL) of crude extracts were used to assess free radical inhibitory ability and ascorbic acid (1.56 to 50  $\mu$ g/mL) was taken as standard. The absorbance of crude extracts/ standard was measured at 517 nm with UV-Visible spectrophotometer

(Shimadzu) and free radical inhibitory activity was determined using the following equation:

% inhibition = 
$$(C-T)/C \times 100$$

where, C = Absorbance of control, T = Absorbance of crude extracts/standard.

Fig. 1 shows the percentage of DPPH free radical inhibition produced by four extracts of G. *velutinum*. There were significant differences in antioxidant activity of HGV ( $IC_{50}=30\mu g/ml$ ), CGV ( $IC_{50}=25.4\mu g/ml$ ), MGV ( $IC_{50}=0.755\mu g/ml$ ) and EGV ( $IC_{50}=2.6 \mu g/ml$ ). Antioxidant activity of MGV was much higher than those of the other extracts. From the results, it is also seen that n-hexane extract possesses less antioxidant activity.



Fig. 1. DPPH free radical scavenging activity of Glochidion velutinum

The total phenol content in the extracts was determined using Folin-Ciocalteu reagent (FCR) based on colorimetric method at 765nm by following standard procedure (Harborne *et al.* 2009). Gallic acid was used as standard to produce calibration curve. The phenolic content was expressed as mg of gallic acid equivalents (GAE) /gm of extract. Total flavonoid content in the extracts was determined using the colorimetric method involving aluminum chloride (Chang *et al.* 2002). Quercetin was used as a reference standard and flavonoid contents of extracts were calculated as mg of QE/ gm of extract. The mean and standard errors of the phenolic and flavonoid compounds of the extracts of G. *velutinum* have been shown in Figs. 2 and 3 respectively. Methanolic and ethanolic extracts of GV had greater amount of phenolics (29.89  $\pm$  1.22 & 17.9  $\pm$  1.26 mg/gm, GAE) as well as flavonoids (48.12  $\pm$  2.28 & 30.22  $\pm$  2.84 mg/gm, QE) as compared to chloroform and *n*-hexane extracts. However, no remarkable differences were found in phenolic and flavonoid content of CGV and HGV.



Fig. 2: Total phenolic content of extracts of *Glochidion velutinum* Fig. 3: Total flavonoid content of extracts of *Glochidion velutinum* 

The antibacterial activity of the extracts was determined by the disk diffusion method using nutrient agar medium (Baur *et al.* 1966). Gram positive and Gram negative bacterial species were collected as pure cultures from the Department of Microbiology, Gono Bishwabidyalay. The sterile Whatman-1 filter paper disks (6mm), impregnated with 500  $\mu$ g of hexane, chloroform, methanol and ethanol extract, were placed gently on the previously marked zones on the agar plates seeded in the test organisms. The plates were incubated at 37°C for 24 hour. The zones of inhibition produced by the extracts against different microorganism were measured. From the results of antibacterial activity shown in Table 2, it is evident that the methanol and ethanol extracts showed mild sensitivity to *Escherichia coli*. Hexane, chloroform, and ethanol extracts did not show any sensitivity to *Escherichia coli*. No zone of inhibition has been observed in case of negative control (NC).

Microbes	Diameter of inhibited zone(mm)				
	HGV	CGV	MGV	EGV	NC
Staphylococcus aureus	7	9	18	17	nzd
Bacillus subtilis	8	11	21	14	nzd
Pseudomonas aeruginosa	8	11	17	16	nzd
Shigella dysenteriae	7	7	15	11	nzd
Escherichia coli	-	-	6	-	nzd
Streptococcus pyogenes	8	7	19	16	nzd

Table-2: Antibacterial activity of different solvent extracts of Velutinum

\*nzd: no zone detected

Phenols are the omnipresent metabolites in plants and responsible to wide range of therapeutic uses such as antioxidant, antimutagenic, anticarcinogenic, free radical and scavenging activities. The scavenging ability of phenols is due to the presence of phenolic hydroxyl group (Ayesha Siddiqua *et al.* 2010). Antioxidants have important roles in preventing free radical damage to blood, cells, and tissues (Saleh *et al.* 2010).

The present study displayed that (Fig. 1) ethanol and methanol extracts had the highest antioxidant activity as compared to hexane and chloroform extracts. According to a study by Erkan *et al.*, (2008), there is a close relationship between antioxidant activity and total phenolic compounds of natural extracts. So the present study clearly demonstrates that all the extracts of experimental plant possess antioxidant activity. The presence of secondary metabolites has been reported to be responsible for their antibacterial properties (Adeshina *et al.* 2010). The ethanol and methanol extracts exhibited moderate antimicrobial (greater zone of inhibition) activity against microbes used in this study as compared to the hexane extract. This may be due to alkaloids and saponins being largely present in the methanol extract. The finding is consistent with the reports that nitrogen containing naturally occurring alkaloids have antimicrobial properties due to their ability to intercalate with the DNA of microorganisms (Kasolo *et al.* 2010).

From the present experiments, it is clear that antioxidant and antibacterial activities of *Glochidion velutinum* varied significantly depending on the extracting solvents. Nevertheless, further research is needed for isolation and identification of active substance present in the extracts, which could be rational for medicinal uses.

## REFERENCE

- Adeshina, G., J. Ehinmidu, J. Onaolapo, and L. Odama. 2010. Phytochemical and antimicrobial studies of the ethyl acetate extract of Alchornea cordifolia leaf found in Abuja, Nigeria. *Journal of Medicinal Plant Research* 4(8): 649-658.
- Are, P.C., R.R.R Adidala and G. Puchchakayala. 2011. Hypoglycemic and Antidiabetic Activity of *Glochidion velutinum* on Streptozotocin-Nicotinamide Induced Type 2 Diabetic Rats. *European Journal of Biological Sciences* 3(4): 126-130.
- Ayesha Siddiqua, K.B. premakumari, Rokeya Sultana, Vithya and Savitha. 2010. Antioxidant activity and estimation of total phenolic content of Muntingia calabura by colorimetry. *International J.Chem. Tech. Res.* 2: 205-208.
- Baur. AW, WM Kirby, JC Sherris and M.Turck. 1966. Antibiotic susceptibility testing by a standard single disk method. *Am J Clin Path* **45**: 493-496.
- Braca, A., N.D. Tommasi, L.D. Bari, C. Pizza, M. Politi and I. Morelli. 2001. Antioxidant principles from Bauhinia terapotensis. *Journal of Natural Products* 64: 892-895
- Chang, C.C., M.H. Yang, H.M. Wen and J.C. Chern. 2002. Estimation of total flavonoid content in Propolis by two complementary colorimetric methods. *J. Food Drug Analysis* **10**: 179-180.
- Erkan, N., G. Ayranci and E. Ayranci. 2008. Antioxidant activities of rosemary extract, blackseed essential oil, camosic acid, rosmarinic acid and sesamol. *Food Chem.* **110**(1): 76-82.
- Harbourne, N., E. Marete, J. C. Jacquier, and D. O'Riordan. 2009. Effect of drying methods on the phenolic constituents of meadowsweet (Filipendula ulmaria) and willow (Salix alba). LWT -Food Science and Technology 42(9): 1468-1473.
- Hasan, M.R., T. Islam, M.I.H. Shameem, M.A. Awoal. 2016. Evaluation of Cytotoxic Potentiality of Different Extracts of Glochidion Velutinum Wight's Leaves through Brine Shrimp Lethality Bioassay. World Journal of Pharmaceutical Research, 5(1): 172-179.
- Hoye, A.T., J. E. Davoren, P. Wipf, M. P. Fink, and V. E. Kagan. 2008. "Targeting mitochondria," Accounts of Chemical Research, 41 (1): 87-97.

- Karuppusamy, S. and K.M. Rajasekaran. 2009. High Throughput Antibacterial Screening of Plant Extracts by Resazurin Redox with Special Referance to Medicinal Plants of Western Ghats. *Global J. Pharmacol* 3(2): 63-68.
- Kasolo, J., S. Gabriel, L. Bimenya, O. Joseph, and J. Ogwal-Okeng. 2010. Phytochemicals and uses of Moringa oleifera leaves in Ugandan rural communities. *Journal of Medicinal Plants Research*, 4(9): 753-757.
- Leu, S.J., Y. P. Lin, R. D. Lin et al. 2006. "Phenolic constituents of Malus doumeri var. formosana in the field of skin care," *Biological and Pharmaceutical Bulletin* **29**(4): 740-745.
- Madhavachetty, K., K. Sivaji and K. Tulasi Rao. 2008. Flowering Plants of Chitoor Dist, A.P, India, Student offset printers, Thirupathi. 317.
- Mothana, R.A.A; S. A. A. Abdo, S. Hasson, F. M. N. Althawab, S. A. Z. Alaghbari, and U. Lindequist. 2010. "Antimicrobial, antioxidant and cytotoxic activities and phytochemical screening of some Yemeni medicinal plants," *Evidence-Based Complementary and Alternative Medicine*, 7(3): 323-330.
- Mulabagal, V., S. Van Nocker, D. L. Dewitt and M. G. Nair. 2007. "Cultivars of apple fruits that are not marketed with potential for anthocyanin production," *Journal of Agricultural and Food Chemistry*, 55(20): 8165-8169.
- Saleh, MA., S. Clark, B. Woodard, S.A. Deolu-Sobogun. 2010. Antioxidant and free radical scavenging activities of essential oils. *Ethn Dis*, **20**: 78-82.
- Sandhya, S., R.S.N.A.K.K. Chaitanya, K.R. Vinod, K.N.V. Rao and B.D avid. 2010. An updated review on the Genus Glochidion Plant. *Archives of Applied Science Res.* **2**(2): 309-322
- Sandyha, S., R.S.N.K.K Chaitanya, K.R. Vinod, J. Chandrasekhar, and D. Banji. 2011. Antioxidant and Cytotoxic Potentiality of Glochidion Velutinum. *European Journal of Biological Sciences* 3(3): 78-85.
- Trease, GE and WC. Evans 1978. A Textbook of Pharmacognosy. 11<sup>th</sup> ed. London: *Published by Bailliere Tindall*. 530.

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