

## **ORIGINAL ARTICLE**

# Characterization of seahorse (*Hippocampus comes* L.) extracts originating from culture and nature in Pesawaran, Lampung, Indonesia

Trisnawati Mundijo<sup>1,2</sup> , Franciscus Dhyanagiri Suyatna<sup>3</sup> , Agung Eru Wibowo<sup>4</sup> , Agus Supriyono<sup>4</sup> , Yurnadi Hanafi Midoen<sup>5</sup>

#### **ABSTRACT**

**Objective:** Indonesia is an archipelagic country with a mega biodiversity, among others, in the marine area. Seahorses (*Hippocampus* spp.) are a marine fish known to have biocompounds used in traditional medicine "Jamu," such as *Hippocampus comes* L. (HCL). The present study aims to analyze and compare the chemical contents of cultured seahorse (CS) and natural seahorse (NS) extracts.

**Materials and Methods:** The CS and NS were identified morphometrically. After freeze-drying, the seahorses were ground into powder with a grinder. The seahorse powder was extracted with ethanol and a water solvent. The extract contained biocompounds, proximate, amino acids, and steroids with high-performance liquid chromatography.

Results: The study found unique characteristics of HCL. The highest yield was obtained in NS using a water solvent (18.6%). The biocompounds in seahorses consist of alkaloids and triterpenoids. The highest proximate of water content (11.03%) and ash content (42.50%) was found in NS. In addition, other compounds were also detected in CS, such as fat (7.48%) and protein (47.67%). Both of HCL's different sources had all essential and nonessential amino acids in which the highest concentration were in NS, i.e., L-arginine (56,537.22 mg/kg), L-lysine (17,794.17), glycine (113,649.80 mg/kg), L-proline (47,056.15), and L-alanine (43,451.81). The analysis of the steroid compound of the extract suggested the presence of steroid glycosides.

**Conclusion:** The highest yield of the seahorse extract with a water solvent is about 18.6% and protein content of 47.67% in CS. The crude extract has alkaloids, triterpenoids, and glycine (113,649.80 mg/kg) in NS with water, suggesting the presence of steroid glycosides.

#### **ARTICLE HISTORY**

Received February 24, 2022 Revised July 21, 2022 Accepted July 21, 2022 Published November 19, 2022

#### **KEYWORDS**

Hippocampus comes L.; amino acid; steroid; cultured; nature



© The authors. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License (http://creativecommons.org/licenses/by/4.0)

#### Introduction

Seahorse is a marine fish with great potential as a fishing commodity, in the fishing industry, and as a global food [1,2]. There are more than 50 species of seahorses worldwide [3], and 33 species are found in Indonesia [4]. One of the seahorse species in Indonesia is the *Hippocampus comes* L. (HCL), which is the most used in traditional medicine because of its medicinal value [5–7]. In Indonesia,

HCL is used in traditional medicine as a "Jamu" [4] because it has biocompounds including the aphrodisiac effect, respiratory system, immune system, and antioxidant, antifatigue, and anti-inflammatory properties [8,9]. The bioactive compounds in seahorse extract are steroids, amino acids, protein, taurine, fatty acids, cholesterol, and trace elements [10,11].

**Correspondence** Yurnadi Hanafi Midoen ⊠ yurnadi.kes@ui.ac.id ☐ Department of Medical Biology, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia.

How to cite: Mundijo T, Suyatna FD, Wibowo AE, Supriyono A, Midoen YH. Characterization of seahorse (*Hippocampus comes* L.) extracts originating from culture and nature in Pesawaran, Lampung, Indonesia. J Adv Vet Anim Res 2022; 9(4):610–616.

<sup>&</sup>lt;sup>1</sup>Doctoral Programme Biomedical Science, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia

<sup>&</sup>lt;sup>2</sup>Department of Medical Biology, Faculty of Medicine, Universitas Muhammadiyah Palembang, Palembang, Indonesia

<sup>&</sup>lt;sup>3</sup>Department of Pharmacology and Therapeutics, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia

<sup>&</sup>lt;sup>4</sup>Research Center for Pharmaceutical Ingredients and Traditional Medicine, National Research and Innovation Agency, Tangerang, Indonesia

<sup>&</sup>lt;sup>5</sup>Department of Medical Biology, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia

To date, research on seahorses in Indonesia commonly uses ethanol as a solvent. Several reports described the extraction with ethanol [5], water [12], ether acetate [13], and methanol [14]. However, no previous research has compared HCL extraction solvents to ethanol and water.

Qian et al. [15] reported the extraction with water, methanol, and ethanol used in *Hippocampus kuda* L. Sanaye et al. [16] also used methanol in the extraction of *Hippocampus kuda L.* [8]. However, methanol is not recommended because it can be toxic for humans [17].

So far, studies are yet to document the findings of HCL analysis from both culture and nature. There was only one study, but it utilized a different species. There is a discrepancy between the existing data and the secondary metabolite level of HCL, which is the most common species in Indonesia. Previous studies in Indonesia by Sari et al. [5] and Safryna [10] revealed that HCL extracted with ethanol solvents contained amino acids and steroids. Using a qualitative test, the study identified the possible presence of steroids. The objective of this study is to quantify the steroids using the high-performance liquid chromatography (HPLC) test. In the present study, we quantify the steroids and other compounds of HCL from different habitats using HPLC with different solvents. This study provides a scientific reason for using HCL to help people's health.

#### **Materials and Methods**

#### Ethical approval

The ethics committee of the Faculty of Medicine, Universitas Indonesia, approved this study with protocol number KET-101/UN2.F1/ETIK/PPM.00.02/2021.

# Morphological identification

Morphological identification was made according to the guidelines for seahorse identification [18], obtained from meristic measurements and the unique morphology [10,11].

## Samples

Male and female HCL seahorses were obtained from culture and nature. The cultured seahorse (CS) was collected from the Marine Cultivation Fisheries Center, Marine and Fisheries Department, Lampung, Indonesia. The CS was obtained from Juwana, a tampon tank with aeration. During the juvenile phase, the CS was fed zooplankton like *copepods, Nauplii*, and *Artemia* sp. Shrimps, *Artemia* sp., and *Nauplii* were given after the next development phase, when the size of CS was more than 2 cm. The feedings were done three times a day. The minimal length of an adult seahorse used in the study was 12 cm. Only adult seahorses are commonly used for traditional medicine [19].

The natural seahorse (NS) was collected from the fishers by Karya Usaha Bersama, Karya Laut, Pesawaran, and Lampung, with supervision from the Marine Cultivation Fisheries Center, Marine and Fisheries Department, Lampung, Indonesia.

# Sample collection

The seahorses were stabilized with fish stabilizer (3 ml in 12 l of water), washed with water, and frozen with a freeze dryer (Heto FD4 Diagnostic) for 48 h at -45°C.

The freeze-dried samples were weighed and crushed into powder using a Retsch grinder. The seahorse powder was analyzed for proximate, water content, ash, fat, protein, and carbohydrates, according to the National Standardization Agency, Indonesia [20].

The extraction was done with 5 gm of powder from CS and NS using ethanol and water as the solvent to find the yield, crude extract compounds, amino acids, and steroids. The results for the yield were divided by the total weight (%) [10].

Identifying alkaloids, steroids, triterpenoids, saponin, flavonoids, and tannin was conducted using reagents Dragendrof, Mayer, and Wagner. The steroid and triterpenoid were analyzed using the Liebermann–Buchard method. Saponin and tannin contents were analyzed using magnesium (Mg) and flavonoids were analyzed using chloride acid (HCl) and amyl alcohol [5]. Detection and quantification of amino acids were conducted using the HPLC detector fluorescence and HPLC PDA. Steroids were analyzed using the UV-VIS C-18 column [5,10,21,22].

#### **Ethanol extraction**

Ethanol (96%) was used in the maceration of 5 gm seahorse powder in 20 ml of solvent (w/v) [5]. Maceration was done for 24 h and filtered with Whatman 42. Next, the macerated HCL was processed using the rotary evaporator Heidolph, Buchi Vacuum Pump V-700, 20°C–25°C.

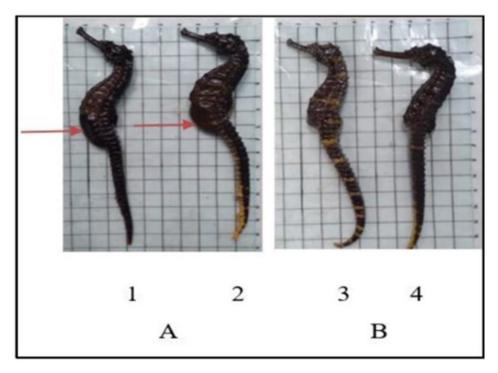
## Water extraction

Five grams of macerated seahorse powder were extracted with 50 ml of phosphate buffer for 24 h and stirred at 500 rpm for 2 h [15]. The mixture was centrifuged [11], and the supernatant was frozen for 48 h at -45°C and stored at -80°C [21].

## **Results**

# Morphology of a seahorse

The HCL from Pesawaran Lampung, has a unique morphology, as shown in Figure 1.



**Figure 1.** HCL from a private collection. A: Male; B: Female; 1 and 4: NS; and 2 and 3: CS. Arrows: Brood pouch in male seahorses.

#### **Proximate tests**

The proximate analysis is shown in Figure 2, of which water (11.03%) and ash content (42.50%) were the highest constituents in NS. Fat (7.48%) and protein content (47.67%) were the highest in CS.

### **Extraction**

The extraction details are shown in Table 1. The results show that the highest yield in water solvent for CS and NS extractions was, respectively, 15.4% and 18.6%. The alkaloid and triterpenoid content of the extract were also high.

Essential and nonessential amino acids were detected in both seahorses (Table 2), in which NS had the highest contents of L-arginine (56,537.22 mg/kg), L-lysine (17,794.17), glycine (113,649.80 mg/kg), L-proline (47,056.15), and L-alanine (43,451.81).

The analysis of steroids showed no similar steroids to the standard. The peak chromatographic details are shown in Table 3.

## **HPLC** for steroid

The HPLC and absorbance profile in Table 3 show that the peaks of the analyzed extract were almost as high as the standards. In addition, the retention time of the extract chromatogram appeared earlier than the standard. The NS extract with a water solvent had the best results compared with other extracts as indicated by the profile similarity

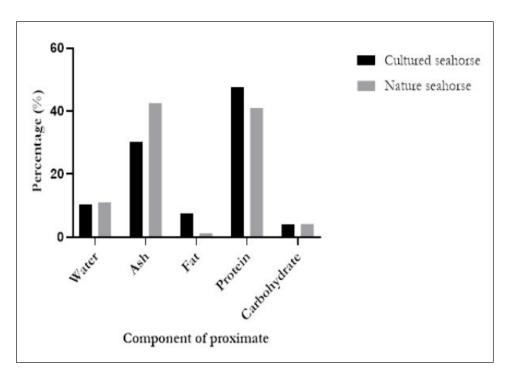
and maximum absorbance according to electroVolt absorbance, i.e., 750 mAU for ethinyl estradiol, 1.000 mAU for methyl testosterone, and 500 mAU for progesterone. We also confirmed the presence of steroid glycosides with the Keller–Killiani assay (Fig. 3).

# **Discussion**

In the present study, we used adult HCL from Pesawaran, Lampung, at least 12 cm in length. Similar to previously reported studies, we found a unique morphology in the seahorses with double spine cheeks, patches on the body, and a reddish yellow color on the tail [4,5,9–11,21].

In our study, we used a seahorse, at least 12 cm in length, which is an adult and used for traditional medicine [19]. In addition to that, the Convention on International Trades in Endangered Species (CITES) recommends that the length of a seahorse be at least 10 cm [23] to avoid extinction in wild nature [24].

The novelty in our study is the characterization of the extracts in which we compare the NS and CS with different solvents. We found that the yield from a water solvent was the highest percentage in NS (18.6%). In proximate analysis, fat content (7.48%) and protein content (47.67%) were higher in CS, but the water (11.03%), ash (42.50%), and carbohydrate contents (4.18%) were higher in NS. The protein content, the highest constituent in proximate analysis, was similar to that in previous studies [5,10,25]. Our



**Figure 2.** The proximate test of HCL. Fat content (7.48%) and protein content (47.67%) were highest in CS; on the other hand, water content (11.03%), ash content (42.50%), and carbohydrate (4.18%) were mostly high in NS.

**Table 1.** The characterization of the HCL extract.

Downston	cs		NS	
Parameters	Ethanol solvent	Water solvent	Ethanol solvent	Water solvent
Yield (%)	7.0	15.4	11.4	18.6
Biocompounds				
Alkaloid	+	+	+	+
Flavonoid	-	-	-	_
Saponin	-	-	-	-
Steroid	-	-	-	-
Triterpenoid	+	+	+	+
Tannin	-	-	-	-
Steroids				
Ethinyl estradiol	-	-	-	-
Methyl testosterone	-	_	_	-
Progesterone	-	_	_	_

The NS extract has a high yield with a water solvent (18.6%); only has alkaloids and triterpenoids; and there is none of the same steroids with the standards.

study found alkaloids and triterpenoids, which are different from other studies conducted in Indonesia [5,10].

In our study, we found that amino acids L-arginine (56,537.22 mg/kg), L-lysine (17,794.17), glycine (113,649.80 mg/kg), L-proline (47,056.15), and L-alanine (43,451.81) were the highest in NS. These data suggested

a favorable condition for the NS to have a better survival capacity in their living environment than CS. The NS must be able to adapt to changes in the marine environment consisting of various climate changes, reclamation, pollution, etc. [2]. This study found that the presence of high amounts of amino acids in HCL may potentially be used

<sup>(+):</sup> Identified, (-): Not identified.

for nutraceutical natural products from marine sources. Marine organisms are natural products with structure and bioactivity that are highly different from those on the mainland. This study also increases our interest in exploring drug development opportunities from natural products [23,26].

**Table 2.** Amino acids in seahorses (the levels are presented from the highest to the lowest in NS).

Commound name	Level of amino acids (mg/kg)		
Compound name	cs	NS	
Essential			
L-Arginine	54,743.61	56,537.22	
L-Phenylalanine	27,695.61	23,213.67	
L-Threonine	24,034.43	22,491.30	
L-Leucine	26,880.58	21,904.59	
L-Valine	20,007.96	19,287.34	
L-Lysine	17,355.09	17,794.17	
L-Histidine	14,368.55	12,725.72	
L-Isoleucine	13,671.32	11,335.90	
L-Methionine	3,645.71	3,357.61	
L-Tryptophane	2,281.78	1,300.00	
Nonessential			
Glycine	90,909.58	113,649.80	
L-Proline	39,633.42	47,056.15	
L-Glutamate Acid	46,796.58	45,812.52	
L-Alanine	40,175.12	43,451.81	
L-Aspartate Acid	29,895.15	29,559.04	
L-Serine	26,273.32	23,958.78	
L-Tyrosine	17,523.82	13,003.43	
L-Sistine	1,161.92	863.57	

The HCL seahorse extracts have essential and nonessential amino acids, with the highest levels in L-arginine (56,537.22 mg/kg), L-lysine (17,794.17), glycine (113,649.80 mg/kg), L-proline (47,056.15), and L-alanine (43,451.81) in NS.

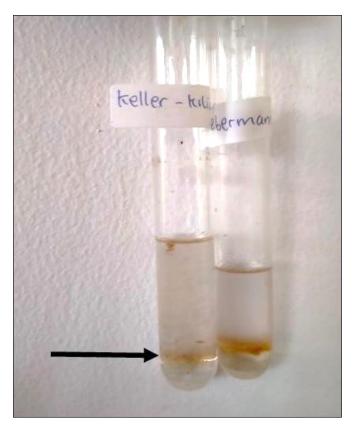
We observed that the amino acids in our results differ from those of Sun et al. [21], who reported that the highest level of amino acids came from CS. The difference might be partly due to the studied species and the body tissue of the seahorse used for extraction.

Our study is the first in Indonesia to use the HPLC method to find the steroid in HCL. The chemical components in HCL indicated the presence of steroid glycosides in both HCL and solvents. The analyzed quantitative steroids with liquid chromatography-mass spectrometry (LC-MS) showed that the four compounds with Mw of 240, 619, 717, and 755 were not ethinyl estradiol, methyltestosterone, or progesterone, which were used as standards. Indeed, we detected the compounds as peaks in the HPLC with an earlier appearance of retention time and gave positive results with Keller-Killiani reactions. The positive results with the Keller-Killiani reaction suggested the presence of steroid glycosides in HCL with sugar (glycoses) and nonsugar (aglycones) side chains. Aglycones are commonly a secondary metabolite from steroids, alkaloids, and saponin compounds that can influence biological activity. During extreme conditions, some fish can produce this metabolite, such as diosgenin and aglycone compounds. Steroid glycosides in marine organisms have a different structure in comparison with terrestrial organisms, which contain steroid aglycones and carbohydrates. Exploration of steroid glycosides from HCL and marine organisms is needed in the future, considering that 70% of the world is covered by the sea [27–30].

Another study found steroids and cholesterol to be from different species of seahorses. Other authors found oxacycloheptadecan-2-one, chrysophanol, cholesteryl benzoate, eicosapentaenoic acid, and docosahexaenoic acid in

**Table 3.** Data on HPLC with retention time and wavelength values of standard compounds and compounds in seahorse extracts.

Standard compounds/extract	Retention time	UV absorbance (nm)	mAU	
Ethinyl estradiol	4.100	248.79; 279.98; 356.13	300	
Methyl testosterone	3.500	208.13; 240.93; 282.68; 374.48; 486.99; 521.75; 654.97; 749.18	300	
Progesterone	7.408	202.78; 242.29; 361.05	50	
	3.005	239.4; 347.76	110	
Ethanol extract of CS	2.705	485.83; 550.37; 577.35; 666.93; 738.28.	300	
	2.555	-	200	
	2.549	248.84; 275.66	350	
Water extract of CS	2.693	249.13; 273.64; 485.76; 514.11; 581.14; 631.80; 665.5; 756.46	600	
	2.566	248.61; 274.86	350	
	2.510	240.74; 251.75; 350.73	70	
Ethanol extract of NS	2.672	193.36; 240.2; 424.99; 485.39; 581.03; 663.85	150	
	3.057	239.65; 250.69; 355.01	50	
	2.472	252.81; 274.91	750	
Water extract of NS	2.707	195.51; 252.39; 467.55; 581.54; 631.77; 655.6; 748.03	1,000	
	2.580	253.64; 274.48	500	



**Figure 3.** Keller–Killiani test shows a ring (arrow). It clearly shows a positive result with a brown ring, indicating the presence of steroid compounds.

Hippocampus erectus [14]. Wu et al. [31] found cholest-4-en-3-one, 3-β-hydroxycholest-5-en-7-one, cholest-5-ene-3β,  $7\beta$ -diol, cholest-5-ene-3β,  $7\alpha$ -diol in Hippocampus trimaculatus.

From these data, the results of the study consisting of yield value, proximate content, biocompounds, amino acids, and steroid glycosides conclude that NS with a water solvent has more optimal results than CS with an ethanol solvent. Therefore, HCL seahorses in nature with a water solvent are feasible to explore and to investigate its value in traditional medicine.

## **Conclusion**

The HCL seahorse extracts from Pesawaran, Lampung, Indonesia, has a high yield in CS with a water solvent of about 18.6%, protein of 47.67%, and only identified alkaloids and triterpenoids. The most elevated amino acids are shown in NS with a water solvent and an indicated steroid glycoside. The results of this study will be helpful in further reviewing data on seahorses, especially in Indonesia, which potentially can be used as a natural product for medicinal use.

## List of abbreviations

HCL. *Hippocampus comes* L.; CS, Cultured seahorse; NS, Nature seahorse; HPLC, High-performance liquid chromatography; LC-MS, Liquid Chromatography-Mass Spectrometry; UV, Ultraviolet; NSA, National Standardization Agency; CITES, Convention on International Trades in Endangered Species.

# **Acknowledgments**

The authors would like to thank Mr. Ali Hafiz Al Qodri, A.Pi, from the Ministry of Maritime Affairs and Fisheries of Indonesia, Directorate General of Aquaculture, Lampung, for collecting the seahorse. They also thank Universitas Indonesia for funding this research through a PUTI grant with contract number NKB-578/UN2.RST/HKP.05.00/2020. The authors are very grateful to Mr. Nuralih, S.Si., M.Si., Mr. Nurhadi, S.Si., and Mr. Hasan for guiding the extraction process at the National Research and Innovation Agency, Indonesia. They also thank Mr. Aryo Tedjo, S.Si., M.Si. for guiding the analysis.

#### **Conflicts of interest**

The authors have no conflicts of interest to declare.

## **Authors' contributions**

TM conducted the study, collected the data, and wrote the original draft under the supervision of YHM, FDS, and AEW. YHM designed and conceptualized the research; acquired the funding; conducted the experiment; administered the data, methodology, and project; investigated the process and resources; supervised and analyzed the data, validation, and visualization; and reviewed the writing and editing. FDS designed and conceptualized the research; acquired the funding; conducted the experiment; administered the data, methodology, and project; investigated the process and resources; supervised and analyzed the data, validation, and visualization; and reviewed the writing and editing. AS designed and conceptualized the research; supervised the methodology and resources; analyzed the data and validation. YHM, FDA, AEW, and AS approved the final manuscript.

#### References

- [1] Setyono DED. Biological characteristics of seahorses (*Hippocampus* spp.) as a basic knowledge of their cultivation. Oseana 2020; 45(1):70–81; https://doi.org/10.14203/oseana.2020. Vol.45No.1.57
- [2] Koning S, Hoeksema BW. Review: diversity of seahorse species (*Hippocampus* spp.) in the international aquarium trade. Diversity 2021; 13:187; https://doi.org/10.3390/d13050187
- [3] Saher NU, Kamal M, Rodini M, Naz F, Narejo AH. First accord and morphological identification of the seahorse *Hippocampus kelloggi* great seahorse Jordan and Snyder (1902) in coastal waters of Pakistan. Oceanogr Fish 2021; 13(2):1–7; https://10.19080/ OFOAJ.2021.13.555859

- [4] Nurilmala M, Nurhayati KH, Butet NA, Sudrajat AO. Molecular marker based on 16S rRNA gene for seahorse (*Hippocampus* spp.) from Bintan Island-Indonesia. In The 3rd EIW IOP Conference Series: Earth Environmental Science, 2019, Vol. 278, pp 1–10; https://10.1088/1755-1315/278/1/0120541
- [5] Sari EM, Nurilmala M, Abdullah A. Amino acid, and bioactive compounds of seahorse (*Hippocampus comes*). J Trop Marine Sci Technol 2017; 9(2):605–17; http://dx.doi.org/10.29244/jitkt. v9i2.19295
- [6] Chen X, Yi Y, You X, Liu J, Shi Q. High-throughput identification of putative antimicrobial peptides from multi-omics data of the lined seahorse (*Hippocampus erectus*). Mar Drugs 2020; 18:30; https:// doi.org/10.3390/md18010030
- [7] Xie J, Bu L, Jin S, Wang X, Zhao Q, Zhou S, et al. Outbreak of vibriosis caused by Vibrio harveyi and Vibrio alginolyticus in farmed seahorse Hippocampus kuda in China. Aquaculture 2020; 523:735168; https://doi.org/10.1016/j.aquaculture.2020.735168
- [8] Kang N, Kim SY, Rho S, Ko JY, Jeon YJ. The anti-fatigue activity of a mixture of seahorse (*Hippocampus abdominalis*) hydrolysate and red ginseng. Fish Aquat Sci 2017; 20:1–8; https://doi. org/10.1186/s41240-017-0048-x
- [9] Zhang YY, Bo-Mi R, Qian ZJ. A review-biology: aquaculture and medical use of seahorse (*Hippocampus spp*). Ann Res Rev Biol 2017; 14(5):1–12; https://doi.org/10.9734/ARRB/2017/34152
- [10] Safryna DA. Characteristics of the seahorse (Hippocampus comes) and hydrolysate. Thesis, Bogor Agricultural Institute, Bogor, Indonesia, 2020.
- [11] Guo Z, Lin D, Guo J, Zhang Y, Zheng B. In vitro antioxidant activity and in vivo anti-fatigue effect of seahorse (Hippocampus). Peptides Mol 2017; 22(482):1–24; https://doi.org/10.3390/molecules22030482
- [12] Adam K, Fitria L, Satro M. The effect of the protein fraction of seahorse extract (*Hippocampus kuda* Bleeker, 1852) on increasing hemoglobin levels of mice (*Mus musculus* L). Indones Pharm J 2014; 4(2):83–90.
- [13] Xu DH, Wang LH, Mei XT, Li BJ, Lu JL, Xu SB. Protective effects of seahorse extracts in a rat castration and testosterone-induced benign prostatic hyperplasia model and mouse oligospermia's model. Environ Toxicol Pharm 2014; 37:679–88; http://dx.doi. org/10.1016/j.etap.2014.02.001
- [14] Li K, Yan L, Zhang Y, Yang Z, Zhang C, Li Y, et al. Seahorse treatment improves depression-like behavior in mice exposed to CUMS through reducing inflammation/oxidants and restoring neurotransmitter and neurotrophic function. J Ethno Pharm 2020; 250:112487; https://doi.org/10.1016/j.jep.2019.112487
- [15] Qian ZJ, Ryu B, Kim MM, Kim SK. Free radical and reactive oxygen species scavenging activities of the extracts from seahorse, Hippocampus kuda Bleeker. Biotechnol Bioproc Eng 2008; 13:705–15; https://doi.org/10.1007/s12257-008-0093-5
- [16] Sanaye SV, Pise NM, Pawar AP, Parab PP, Sreepada RA, Pawar HB, et al. Evaluation of antioxidant activities in captive-bred cultured yellow seahorse, *Hippocampus kuda* (Bleeker, 1852). Aquaculture 2014:631291; https://doi.org/10.1016/j.aquaculture.2014.08.007

- [17] Pubchem.ncbi.nlm.nih.gov. Compound summary methanol. 2022. Available via https://pubchem.ncbi.nlm.nih.gov/compound/887
- [18] Lourie SA, Foster SJ, Cooper EWT, Vincent ACJ. A guide to the identification of seahorses. Project seahorse and traffic. The University of British Columbia and World Wildlife Fund, Vancouver, Canada, 2004.
- [19] Ministry of Maritime Affairs and Fisheries of Indonesia. Directorate General of Aquaculture, Lampung. Cultivation of seahorse (*Hippocampus* spp.). Marine cultivation technical guidelines. Lampung, Indonesia. 2018; 26:1-18.
- [20] Indonesian National Standard. How to test food and drink. National standardization agency. SNI 01-2891-1992.
- [21] Sun J, Xia S, Xie S, Yang Y, Cui P, Shao P, et al. Biochemical composition of wild and cultured seahorses (*Hippocampus kuda Bleeker*). Aquac Res 2020; 51(4):1542–50; https://doi.org/10.1111/are.14502
- [22] Nasution NA, Nurilmala M, Abdullah A. Hydrolysate of seahorse (*Hippocampus kuda*) an assay of anti-inflammatory activity with protein denaturation inhibition method. Fish J 2019; 21(1):47–51; https://doi.org/10.22146/jfs.43699
- [23] Koldewey HJ, Martin-Smith KM. Review: a global review of seahorse aquaculture. Aquaculture 2010; 302:131–52; https:// doi.org/10.1016/j.aquaculture.2009.11.010
- [24] Foster SJ, Vincent ACJ. Holding governments accountable for their commitments: CITES review of significant trade for a very high-volume taxon. Glob Ecol Conserv 2021; 27:e01572; https:// doi.org/10.1016/j.gecco.2021.e01572
- [25] Lin Q, Lin J, Lu J, Li B. Biochemical composition of six seahorse species, *Hippocampus sp* from the Chinese coast. J World Aquac Soc 2008; 39(2):225–34; https://doi. org/10.1111/j.1749-7345.2008.00159.x
- [26] Meikasari NS, Nurilmala M, Butet NA, Sudrajat AO. PCR-RFLP as a detection method of allelic diversity seahorse *Hippocampus comes* (Cantor, 1849) from Bintan waters, Riau Island. Embryo. IOP Conference Series: Earth Environmental Science, 2020, Vol. 404, p 012046; https://doi.org/10.1088/1755-1315/404/1/012046
- [27] Ivanchina NV, Kicha AA, Stonik VA. Review: steroid glycosides from marine organisms. Steroids 2011; 76:425–54; https://doi. org/10.1016/j.steroids
- [28] Li K, Cai J, Su Z, Yang B, Liu Y, Zhou X, et al. Glycosylated natural products from marine microbes. Front Chem 2020; 7:879; https:// doi.org/10.3389/fchem.2019.00879
- [29] Kalinin VI, Ivanchina NV, Krasokhin VB, Makarieva TN, Stonik VA. Review: glycosides from marine sponges (*Porifera, Demospongiae*): structures, taxonomical distribution, biological activities, and biological roles. Mar Drugs 2012; 10:1671–710; https://doi. org/10.3390/md10081671
- [30] Seipp K, Geske L, Opatz T. Review: marine pyrrole alkaloids. Mar Drugs 2021; 19:514; https://doi.org/10.3390/md19090514
- [31] Wu J, Liu Z, Su J, Lu H, Liao D, Song Q. Chemical constituents of the seahorse *Hippocampus trimaculatus* from east China sea. Chem Nat Comp 2017; 53(5):982–3; https://doi.org/10.1007/ s10600-017-2178-x