

ORIGINAL RESEARCH ARTICLE

OPEN ACCESS

Antioxidant and cytotoxic potential of methanol extract of *Tabernaemontana divaricata* leaves

Nowshin Nowaz Rumzhum, *Md. Mostafizur Rahman, Md. Khalequzzaman Kazal

Laboratory of Pharmacognosy and Pharmacology, Department of Pharmacy, Stamford University Bangladesh, Dhaka-1217, Bangladesh

Abstract

The methanolic extract obtained from the leaves of *Tabernaemontana divericata* (Family: Apocynaceae) was evaluated for *in vitro* antioxidant potential by determination of total antioxidant capacity, assay of nitric oxide scavenging activity and reducing power test. The extract was also screened for its cytotoxic effect using brine shrimp lethality bioassay. The results revealed potent antioxidant property in all antioxidant assays compared to the reference antioxidant, ascorbic acid in a dose dependent manner. Further, the methanolic extract of *Tabernaemontana divericata* showed significant cytotoxic effect (LC₅₀: 3.12µg/ml) compared with positive control, Vincristine Sulphate (LC₅₀: 0.331µg/ml).

Key Words: Tabernaemontana divaricata, Apocynaceae, Antioxidant, Cytotoxicity.

INTRODUCTION

Human beings have been utilizing plants for basic preventive and curative health care since time immemorial. It has been reported that phytochemicals, non-nutritive chemicals present in fruits and herbs may protect human from a host of diseases for their biological activities (Argal and Pathak, 2006). Plant produces these chemicals to protect itself but recent research demonstrates that many phytochemicals can protect humans against diseases. Natural products, either as pure compounds or as standardized plant extracts, provide unlimited opportunities for new drug leads because of the unmatched availability of chemical diversity (Newman and Cragg, 2007). Evaluation of the local flora exploited in traditional medicine for various biological activities is a necessary first step in the isolation and characterization of the active principle and further leading to drug development (Rahman et al., 2011). A large number of plants have been screened as a viable source of natural

Contact No.: +880 1717 551 617

antioxidants including tocopherols, vitamin C, carotenoids and phenolic compounds which are responsible for maintenance of health, to help the human body reduce oxidative damage and protection from coronary heart diseases and cancer (Yanga *et al.*, 2002). In view of these *Tabernaemontana divaricata* was studied for its potential antioxidant and cytotoxic effects.

Tabernaemontana divaricata (Bengali name - Togor; Family - Apocynaceae) is a beautiful evergreen shrub, about 54cm high, with large shiny leaves, crepe jasmine flowers, may appear sporadically all year. T. divaricata, garden plant in tropical countries, is a rich source of alkaloids with various pharmacological properties. It has been used in the folk medicine for anti-infection, anti-inflammation, analgesic, anti-tumour, antioxidative effect and the (Ghani, effect in neuronal activity 2003; Pratchayasakul et al., 2008). In this study, the antioxidant and cytotoxic properties of the methanolic extract are being reported to validate the traditional use of the crude drug through in vitro evaluation.

© 2012 Rumzhum et al.; licensee Saki Publishing Club. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by-nd/3.0/), which permits unrestricted use (including commercial use), distribution and reproduction of the work in any medium, provided the original work is properly cited and remain unaltered.

^{*}Corresponding Author:

Md. Mostafizur Rahman, Assistant Professor

Department of Pharmacy, Stamford University Bangladesh 51, Siddeswari Road, Dhaka-1217, Bangladesh

E-mail: hasanmostafiz@yahoo.com

EXPERIMENTAL METHODS

Plant materials

Tabernaemontana divaricata was collected from Dhaka in March 2008 and was identified by Bangladesh National Herbarium, Mirpur, Dhaka, where a voucher specimen (DCAB accession no.: 32069) has been deposited. The leaves of the plant were first sun dried and then ground into coarse powder.

Extraction of plant materials

About 100gm of powered material was taken in a clean, flat-bottomed glass container and soaked in 500ml of 90% methanol. The container with its contents was sealed and kept for a period of 7 days accompanying occasional shaking and stirring. The whole mixture then underwent a coarse filtration by a piece of clean, white cotton material. The filtrate (methanol extract) obtained was evaporated under ceiling fan and in a water-bath until dried. It rendered a gummy concentrate of blackish color. The gummy concentrate was designated as crude extract of methanol. To get preliminary idea about the active constituents present in the plant leaves extracts different chemical tests were performed and showed the presence of alkaloid, flavonoids and tannins (Evans, 1989).

Antioxidant property

Qualitative assay

A suitably diluted stock solutions (sample solutions) were spotted on pre-coated Silica gel TLC (Thin layer chromatography) plates and the plates were developed in solvent systems of different polarities (polar, medium polar and non-polar) to resolve polar and non-polar components of the extract and to choose the solvent system in which stock solutions run well. The plates were dried at room temperature and were sprayed with 0.02% DPPH in ethanol. Bleaching of DPPH by the resolved bands was observed for 10 minutes and the color changes (yellow on purple background) were noted (Sadhu *et al.*, 2003).

Quantitative assay

Free radical scavenging activity of the methanol extract was evaluated by determination of total antioxidant capacity, assay of nitric oxide scavenging activity and reducing power test. In all methods ascorbic acid was used as standard.

Determination of total antioxidant capacity

The antioxidant activity of the extract was evaluated by the phosphomolybdenum method according to the procedure of Prieto and colleagues (Prieto et al., 1999). The assay is based on the reduction of Mo(VI)-Mo(V) by the extract and subsequent formation of a green phosphate/Mo(V) complex at acid pH. 0.3 ml extract was combined with 3ml of reagent solution (0.6M sulfuric acid, 28mM sodium phosphate and 4mM ammonium molybdate). The tubes containing the reaction solution were incubated at 95°C for 90 min. Then the absorbance of the solution was measured at 695nm using a spectrophotometer (Hach, DR-4000U) against blank after cooling to room temperature. Methanol (0.3ml) in the place of extract was used as the blank. The antioxidant activity is expressed as the number of equivalents of ascorbic acid.

Assay of Nitric oxide scavenging activity

The procedure is based on the method, where sodium nitroprusside in aqueous solution at physiological pH spontaneously generates nitric oxide, which interacts with oxygen to produce nitrite ions that can be estimated using Greiss reagent. Scavengers of nitric oxide compete with oxygen leading to reduced production of nitrite ions. For the experiment, sodium nitroprusside (10mM) in phosphate buffered saline was mixed with different concentrations of methanolic extract of T. divaricata dissolved in methanol and incubated at room temperature for 150 min. The same reaction mixture without the methanol extract but the equivalent amount of methanol served as the control. After the incubation period, 0.5ml of Griess reagent (1% sulfanilamide, 2% H₃PO₄ and 0.1% N-(1naphthyl) ethylene-diamine-dihydrochloride was added. The absorbance of the chromophore formed was read at 546 nm (Sreejayan and Rao, 1997).

Reducing power test

The reducing power of the extract was determined according to the method of Oyaizu (Oyaizu, 1986). Different amounts of extracts (50-250mg) in 1ml of methanol were mixed with phosphate buffer (2.5ml, 0.2mol/l, pH 6.6) and potassium ferricyanide [K₃Fe(CN)₆] (2.5ml, 1%). The mixture was incubated at 50°C for 20 min. A portion (2.5 ml) of trichloroacetic acid (10%) was added to the mixture, which was then centrifuged (650 x g at room

 Table 1: Total antioxidant capacity of methanolic extract
 of Tabernaemontana divaricata.

Materials	Concentration (µg/ml)	Equivalent to ascorbic acid
Methanol extract of Tabernaemontana divaricata	10	0.065±0.09
	25	0.102±013
	50	0.243±0.17
	125	0.636±0.10
	250	1.034 ± 0.06
	500	1.991±0.12

temperature) for 10 min. The upper layer of solution (2.5ml) was mixed with distilled water (2.5ml) and FeCl₃ (0.5ml, 0.1%), and the absorbance was measured at 700nm. Increased absorbance of the reaction mixture indicated increased reducing power.

Cytotoxicity study

Brine shrimp lethality bioassay (Mclaughlin, 1982; Persoone, 1988) technique was applied for the determination of cytotoxic property of methanolic extract of *T. divaricata*.

Preparation of positive control group

Vincristine sulphate was used as the positive control. Measured amount of the vincristine sulphate was dissolved in DMSO to get an initial concentration of 20μ g/ml from which serial dilutions were made using DMSO to get 10μ g/ml, 5μ g/ml, 2.5μ g/ml, 1.25μ g/ml, 0.625μ g/ml, 0.3125μ g/ml, 0.15625μ g/ml, 0.078125μ g/ml and 0.0390μ g/ml. Then positive control solutions were added to the pre-marked vials containing ten living brine shrimp nauplii in 5ml simulated sea water to get the positive control groups.

Preparation of negative control group

100µl of DMSO was added to each of three premarked glass vials containing 5ml of simulated sea water and 10 shrimp nauplii to use as control groups. If the brine shrimps in these vials show a rapid mortality rate, then the test was considered as invalid as the nauplii died due to some reason other than the cytotoxicity of the compounds.

Counting of nauplii

After 24h, the vials were inspected using a magnifying glass and the number of survived

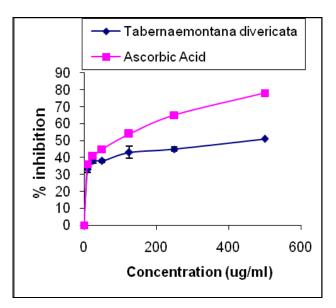


Figure 1: Nitric oxide scavenging activity of methanolic extract of *Tabernaemontana divaricata*.

nauplii in each vial was counted. From this data, the percent (%) of lethality of the brine shrimp nauplii was calculated for each concentration.

RESULTS

Antioxidant property

Qualitative assay

The color changes (yellow on purple background) on the TLC plates were observed due to the bleaching of DPPH by the resolved bands.

Quantitative assay

Total antioxidant capacity

Total antioxidant capacity exerted by the extract is concentration dependent. It was observed that the extract was likely to have the capacity of reduction of Mo (VI) to Mo (V) by the antioxidant principle and the formation of a green phosphate/Mo (V) complex with a maximal absorption at 695nm. The antioxidant activity is expressed as the number of equivalents of ascorbic acid (**Table 1**).

Nitric oxide scavenging activity

From **Figure 1**, it is observed that the extract is likely to have concentration dependent nitric oxide scavenging activity. The leaves may have the property to counteract the effect of NO formation and in turn may be of considerable interest in preventing the ill effects of excessive NO generation in the human body. Further, the scavenging activity may also help to arrest the chain of reactions

Table 2: LC₅₀ data of the test sample of *Tabernaemontana divaricata* in brine shrimp lethality bioassay.

Sample	LC50 (µg/ml)
Vincristine Sulphate	0.331±0.13
Methanolic extract of	3.12±0.11
Tabernaemontana Divaricata	

initiated by excess generation of NO that are detrimental to the human health. Nitric oxide is also implicated for inflammation, cancer and other pathological conditions (Moncada *et al.*, 1991).

Reducing power

Reduction ability of the extract has been investigated from the Fe+++ to Fe++ transformation using the method followed by Oyaizu (Oyaizu, 1986). Earlier authors (Tanaka et al. 1988; Duh, 1998) have observed a direct correlation between antioxidant activity and reducing power of certain plant extracts. The reducing properties are generally associated with the presence of reductones (Duh, 1998) which have been shown to exert antioxidant action by breaking the free radical chain by donating a hydrogen atom (Gordon, 1990). Reductones are also reported to react with certain precursors of peroxide, thus preventing peroxide formation. Figure 2, demonstrates the reduction ability of *T. divaricata*.

Cytotoxic property

Following the procedure of Mayer and colleagues (Meyer *et al.*, 1982) the lethality of all the crude extracts to brine shrimp were determined on *A. salina*. The LC₅₀ obtained from the best-fit line slope was found to be LC₅₀ $3.12\pm0.11\mu$ g/ml, Vincristine sulphate as positive control (**Table 2**). This clearly indicates the presence of potent bioactive principles in this crude extract of which might be very useful as antiproliferative, antitumor, pesticidal and other bioactive agents (Meyer *et al.*, 1982).

DISCUSSION

There is a growing interest in the investigation of natural antioxidant compounds from plants, since they contain secondary metabolites with structural diversity (Joseph and Priya, 2011). In comparison with the synthetic compounds that are currently available, a good natural antioxidant will have a

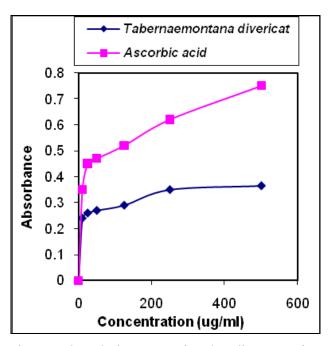


Figure 2: The reducing power of methanolic extract of *Tabernaemontana divaricata*.

higher potency and lower toxicity. A number of known antioxidants as well as yet unknown antioxidants are supposedly present in plants (Haripyaree *et al.*, 2010). These antioxidants are going to do a lot of good to human health by sequestering the hazardous free radicals which are generated due to physiological errors in the cells. Hence, there is currently a strong interest in plants as pharmaceuticals, especially from edible plant parts, because these compounds play an important role, preventing free radical induced diseases such as cancer and atherosclerosis.

There is a close association between cytotoxic properties of natural compounds with their anticancer effect (Mans *et al.*, 2005). Mounting evidence supports that the natural bioactive compound are good candidate for anticancer drugs.

CONCLUSION

Our present study demonstrates the antioxidant and cytotoxic potentialities of *T. divaricata*, which would improve our understanding to the biological role of the plant and future avenue to develop new anticancer therapeutics.

ACKNOWLEDGMENT

Authors wish to thank the authority of Stamford University Bangladesh and the Chairman, Department of Pharmacy of the same for extending their cordial support to perform these investigations.

REFERENCES

Argal, A., Pathak, A.K. (2006). CNS activity of Calotropis gigantean roots. J Ethnopharmacology, 106: 142-145. http://dx.doi.org/10.1016/j.jep.2005.12.024 PMid:16446065

Duh, P.D. (1998). Antioxidant activity of burdock (ArctiumlappaLinne): its scavenging effect on free radical and active oxygen. Journal of the American Oil Chemist's Society, 75(4): 455-461. [DOI]

Evans, W.C. (1989). Trease and Evan's Text book of Pharmacognosy, University Press, Cambridge, UK. 13th Ed. Pp. 546.

Ghani, A. (2003). Medicinal Plants of Bangladesh. The Asiatic Society of Bangladesh, Dhaka. 2nd Ed. pp. 7, 176-177.

Gordon, M.H. (1990). The mechanism of antioxidant action in vitro. Elsevier Applied Science, London. In BJF: Hudson (Ed.) pp. 1–18.

Haripyaree. A., Guneshwor, K., Damayanti, M. (2010).
Evaluation of Antioxidant Properties of Some Medicinal Plants by Sulfur Free Radical Reactivity with Curcumin as Reference. Electronic Journal of Environmental, Agriculture and Food Chemistri, 9(2): 337-344.

Joseph, B., Priya, R.M. (2011). Bioactive Compounds from Endophytes and their Potentials in Pharmaceutical Effect: A Review. American Journal of Biochemistry and Molecular Biology, 1(3): 291-309. http://dx.doi.org/10.3923/ajbmb.2011.291.309

Mans, D.R.A., Rocha, A.B.D., Schwartsmann, G. (2005). Anti-Cancer Drug Discovery and Development in Brazil: Targeted Plant Collection as a Rational Strategy to Acquire Candidate Anti-Cancer Compounds. The Oncologist, 5:185-198. http://dx.doi.org/10.1634/theoncologist.5-3-185 PMid:10884497

Mclaughlin, J.L. (1982). Brine shrimp: a convenient general bioassay for active constituents. Planta. Med, 45: 31-32. http://dx.doi.org/10.1055/s-2007-971236 PMid:17396775

Meyer, B.N., Ferringni, N.R., Puam, J.E., Lacobsen, L.B., Nichols, D.E., McLaughlin, J.L. (1982). Brine shrimp: a convenient general bioassay for active constituents. Planta Medica. 45: 31-32. http://dx.doi.org/10.1055/s-2007-971236 PMid:17396775 Moncada, A., Palmer, R.M.J., Higgs, E.A. (1991). Nitricoxide: physiology, pathophysiology and pharmacology. Pharmacological Reviews, 43(2): 109–142. PMid:1852778

Newman, D.J., Cragg, G.M. (2007). Natural products as sources of new drugs over the last 25 years.J. Nat. Prod, 70 (3): 461-477. http://dx.doi.org/10.1021/np068054v PMid:17309302

Oyaizu, M. (1986). Studies on product of browning reaction prepared from glucose amine. Japanese Journal of Nutrition, 44(6): 307-315. http://dx.doi.org/10.5264/eiyogakuzashi.44.307

Persoone, G. (1988). Proceedings of the international symposium on brine shrimp, Artemiasalina. University Press, Wittern, Belgium. Pp. 1-3.

Pratchayasakul, W., Pongchaidecha, A., Chattipakorn, N., Chattipakorn, S. (2008). Ethnobotany & ethnopharmacology of Tabernaemontana divaricata. Indian J Med Res, 127 : 317-335. PMid:18577786

Prieto, P., Pineda, M., Aguilar, M. (1999). Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: Specific application to the determination of vitamin E. AnalBiochem, 269(2): 337-341. http://dx.doi.org/10.1006/abio.1999.4019 PMid:10222007

Rahman, M.M., Habib, M.R., Hasan, S.M.R., Sayeed, M.A., Rana, M.S. (2011). Antibacterial, Cytotoxic and Antioxidant potential of Methanolic extract of Phyllanthus Acidus L. Int J. Drug Dev. & Res, 3(2): 154-161.

Sadhu, S.K., Okuyama, E., Fujimoto, H., Ishibashi, M. (2003). Separation of Leucasaspera, a Medicinal Plant of Bangladesh, Guided by Prostaglandin Inhibitory and Antioxidant Activities. Chem Pharm Bull, 51 (5): 595-598. http://dx.doi.org/10.1248/cpb.51.595

Sreejayan, N. Rao, M.N.A. (1997). Nitric oxide scavenging by curcuminoids. J. Pharm. Pharmacol. 49(1): 105-107. http://dx.doi.org/10.1111/j.2042-7158.1997.tb06761.x PMid:9120760

Tanaka, M., Kuie, C.W., Nagashima, Y., Taguchi, T. (1988). Applications of antioxidative Maillard reaction products from histidine and glucose to sardine products. Bulletin of the Japanese Society of Scientific Fisheries, 54(8): 1409– 1414. http://dx.doi.org/10.2331/suisan.54.1409

Yanga, J.H., Linb, H.C., Maub, J.L. (2002). Antioxidant properties of several commercial mushrooms. Food Chem, 77: 229-235. [DOI]