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Antioxidant and cytotoxic activities of Ageratum conyzoides stems

Fatema Nasrin

Department of Pharmacy, Southeast University, Banani, Dhaka-1213, Bangladesh

ABSTRACT

Modern civilization is facing more than hundreds of disorders associated with free radicals and natural antioxidants from non-edible plants are gaining importance to fight these disorders. The intention of this report is to evaluate a well known medicinal weed *Ageratum conyzoides* stems for its antioxidant and cytotoxic Effects. Antioxidant potentiality of the crude methanolic extract of the *Ageratum conyzoides* (AC) stems was investigated on DPPH scavenging activity, reducing ability, total antioxidant capacity as well as total phenolic contents. Cytotoxic study was done by brine shrimp lethality bioassay and vincristin sulphate was used as standard. The total phenols and total antioxidant capacity of AC was found to be 38.125 ± 2.01 mg/g equivalent of gallic acid and 333.37 ± 4.22 mg/gm equivalent of ascorbic acid, respectively. The percentage (%) scavenging of DPPH free radical of the extract was found to be concentration dependent with IC₅₀ value 46.01 ± 2.23µg/ml while IC₅₀ value of standard ascorbic acid was found to be 29.56 ± 0.11 µg/ml. The reducing power of AC was found to be concentration dependent. The cytotoxicity exhibited by AC was found promising with LC₅₀ value 1.32µg/ml, comparing with the LC₅₀ (0.689µg/ml) values of vincristin sulphate. The present investigation suggests that *Ageratum conyzoides* possesses remarkable antioxidant and cytotoxic property.

Key Words: Free radicals, AC, DPPH, total phenolic contents. IC₅₀, brine shrimp lethality bioassay.

INTRODUCTION

Medicinal plants constitute an effective source of both traditional and modern medicines. Herbal medicine has been revealed to have valid utility and about 80% rural population depends on its efficacy for their primary health care. Scientist from divergent fields in a similar efforts are investigating flora a new with an eye to their therapeutic worth. A sense of urgency accompanies the search as the pace of species extinction continues. Over the years, the WHO advocated that countries should interact with traditional medicine with a view to identifying and exploiting aspects that provide safe and effective remedies for ailments of both microbial and nonmicrobial origins (WHO 1978). As a part of such study we focused on an annual herb, Ageratum conyzoides L. (Family-Astreracae).

Ageratum conyzoides is erect, 30-80cm tall with fine white hairs on the stem and pink flowers (Kaul and

Corresponding Author:

Fatema Nasrin

Lecturer Department of Pharmacy, Southeast University Banani, Dhaka-1213, Bangladesh E-mail: *nasrin_0209@yahoo.com* Contact No.: +8801913714999 Neelangini, 1989). It is a weed, commonly called Billygoat-weed, Goatweed etc., generally found in cultivated fields and other ecosystems such as pastures, grasslands, wastelands and even forest areas (Batish et al., 2006). The plant is known to have originated from tropical America and now spread to various tropical and subtropical parts of the world (Juliana et al., 2010). This plant is widely utilized in traditional medicine wherever it grows (Bioka et al., 1993). It has been long known in herbal or folk medicine as a remedy for diverse ailments in Africa (Almagboul et al., 2001) and worldwide. Various pharmacological investigations have verified its antibiotic efficacy (Durodola, 1977), analgesic effect in rats (Menut et al., 1993), antioxidative effect (Jagetia et al., 2003), hepatoprotective effects (Ita et al., 2009) and as a blood booster (Ita et al., 2007). The phytochemical screening showed that A. conyzoides contains alkaloids, resins, saponins, tannins, glycosides and flavonoids (Kamboj and Saluja, 2008). Many different compounds have been isolated and identified in A. conyzoides; such as kaempferol and glycoside (rhamnoside); quercetin, scutellarein, eupalestin, chromene, stigmas-7-en-3ol, sitosterol, stigmasterol, fumaric acid, caffeic acid, saponin, pyrrolizidinic alkaloids, ageratochromene

© 2013 Nasrin; licensee Saki Publishing Club. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by-nd/3.0/), which permits unrestricted use (including commercial use), distribution and reproduction of the work in any medium, provided the original work is properly cited and remain unaltered. derivatives and alkane (Nyuna et al., 2010). Furthermore, Ageconyfavones Α, Β, and C; hexametoxyflavone; three coumarinic compounds, including 1-2 benzopirone; 1,2- desifropirrolizidinic and licopsamine have been identified in A. conyzoides (Kamboj and Saluja, 2008; Ladeira et al., 1987). The whole plant is traditionally used in management of wounds, burns and bacterial diseases (Ming, 1999). No studies on cytotoxic and antioxidant effect of A. conyzoides have so far been undertaken. Taking this in view and as a part of our ongoing research on Bangladeshi medicinal plants, the present study aimed to evaluate the antioxidant and cytotoxic activity of the methanolic extract of A. conyzoides stems.

MATERIALS AND METHODS

Plant material

The plant *A. conyzoides* were collected in the month of March 2012 from Jamalpur, Bangladesh. A voucher specimen for this collection has been maintained in the Bangladesh National Herbarium (Voucher Specimen No - DACB 37866), Dhaka, Bangladesh.

Preparation of the extract

The stems of the plant were first washed with water to remove adhering dirt and then dried at 45°C for 36 hrs in an electric oven, then powdered with a mechanical grinder, passing through sieve # 40, and stored in a tight container. The dried powdered material (1kg) was taken in a clean, flat bottomed glass container and soaked in methanol for seven days. The whole mixture then underwent a coarse filtration by a piece of clean, white cotton material. The total filtrate was concentrated to dryness, *in vacuo* at 40°C to render the methanol extract (390g) of brownish red color.

Drugs and chemicals

The active drugs Diazepam were the generous gift samples from Square Pharmaceuticals Ltd., Bangladesh. Ammonium molybdate (Merck, Germany), Sodium phosphate (BDH, England), Potassium ferri-cyanide (K₃[Fe(CN)₆]) Trichloroacetic acid (CCl₃COOH), Folin–Ciocalteu reagent, DPPH free radical, ascorbic acid, gallic acid was obtained from Merck, Germany. All chemicals used were of analytical reagent grade.

In vitro antioxidant activity

The amount of phenolic compounds

The total phenolic content of AC was determined using Folin–Ciocalteu reagent (Yu *et al.*, 2002). The content of total phenolic in the extract of AC was calculated from regression equation of the calibration curve (y = 0.0138x + 0.1275, $R^2 = 0.988$) and is expressed as Gallic acid equivalents (GAE).

Determination of total antioxidant capacity

The antioxidant activity of the extract was evaluated by the phosphomolybdenum method (Prieto *et al.*, 1999). The assay is based on the reduction of Mo (VI)–Mo (V) by the extract and subsequent formation of a green phosphate/Mo (V) complex at acid pH. The total antioxidant of AC was calculated from regression equation of the calibration curve (y = 0.0043x + 0.1503, R² = 0.8874) and is expressed as ascorbic acid equivalents (AAE)

Free radical scavenging activity measured by 1, 1diphenyl-2-picryl-hydrazyl (DPPH)

The free radical scavenging activity of the extract, based on the scavenging activity of the stable 1, 1diphenyl-2- picrylhydrazyl (DPPH) free radical was determined by the method described by Braca (Braca et al., 2001). The 0.1 mmol/L solution of DPPH in methanol was prepared and 1 ml of this solution was added to 3 mL of extract's solution at different concentrations (5, 10, 25 and 50 µg/mL). After 30 min, absorbance was measured at 517 nm. Vitamin C (ascorbic acid) was used as a reference drug. The percentage inhibition activity was calculated from [(A0-A1)/A0] x 100, where A0 is the absorbance of the control, and A1 is the absorbance of the extract/ standard. Median inhibitory concentration (IC50) value was calculated from the equation of line obtained by plotting a graph of concentration (µg/ml) versus % inhibition.

Reducing power activity

The reducing power of AC was determined according to the method described by Oyaizu (Oyaizu, 1986). For the measurement of the reductive ability, transformation of Ferric ion to Ferrous ion was investigated in the presence of extracts. Increased absorbance of the reaction mixture indicated increased reducing power. Ascorbic acid was used as the standard. Phosphate buffer (pH 6.6) was used as blank solution. The absorbance of the final



Figure 1: Free radical scavenging activity of different concentrations of methanolic extract of *A. conyzoides* and ascorbic acid by DPPH radicals.

reaction mixture of two parallel experiments was taken and is expressed as mean \pm standard deviation. Increased absorbance of the reaction mixture indicated increased reducing power.

Cytotoxic activity

Brine shrimp lethality bioassay (Rahman and Rashid, 2008) technique was applied for the determination of general toxic property of AC. Here, in vivo lethality test has been carried out using brine shrimp nauplii eggs (Artemia salina). Vincristin sulphate was used as a positive control in the bioassay. Eggs were placed in one side of a small tank divided by a net containing 3.8% NaCl solution for hatching. In the other side of the tank was placed a light source to attract the nauplii. After 2 days of hatching period the nauplii were ready for the experiment. Four milligrams of the complexes was accurately measured and dissolved in DMSO to get a concentration of varying concentrations 100, 50, 25, 12.50, 6.25, 3.125, 1.563, 0.781, 0.39, 0.19 µg/ml. Ten brine shrimp nauplii were then placed in each vial. For the control test of each vial, one vial containing the same volume of DMSO plus water up to 5 ml was used. After 24 hour of incubation, the vials were observed using a magnifying glass and the number of survivors in each vial were counted and noted. From these data, the percentage of mortality of the nauplii was calculated for each concentration and the 50% leathal concentration (LC50) values were determined.



Figure 2: Reducing power of methanolic extract of A. *conyzoides* and ascorbic acid by spectophotometric detection of Fe³⁺ to Fe²⁺ transformation.

RESULTS

In vitro antioxidant activity

Total phenolic contents

The total phenols content was found to be 38.38 ± 2.01 mg/g plant extract (in GAE) in crude extract of AC.

Total antioxidant capacity

Total antioxidant capacity of AC is expressed as the number of equivalents of ascorbic acid and found to be 333.37 ± 4.22 mg/gm equivalent of ascorbic acid (AAE).

DPPH radical scavenging activity

The percentage (%) scavenging of DPPH free radical was found to be concentration dependent i.e. concentration of the extract between 25-200 μ g/ml greatly increasing the inhibitory activity (Figure 1). The IC₅₀ value was found to be 46.01 ± 2.23 μ g/ml and 29.56 ± 0.11 μ g/ml for AC and standard ascorbic acid, respectively.

Reducing power ability

For the measurement of the reductive ability, we investigated the Fe³⁺ to Fe²⁺ transformation in the presence of AC and compared with standard ascorbic acid (Figure 2). The reducing power of AC was found to be concentration dependent.

Cytotoxic activity

In cytotoxic test activity, % mortality increased gradually with the increase in concentration of the test samples. LC₅₀ values obtained from the best-fit line slope (Figure 3) were 1.32µg/ml and 0.689µg/ml for AC and vincristine sulphate, respectively.



Figure 3: Determination of LC₅₀ values for vincristine sulphate and methanolic extracts of *A. conyzoides* from linear correlation between logarithms of concentration versus percentage of mortality.

DISCUSSION

Recently safety considerations, public's perception and risk reduction of chronic diseases by consumption of fruits and vegetables, have geared interest in the search for natural antioxidants (Dastmalchi et al., 2007). In aerobic environment, all animals and plants require oxygen and hence free radicals (reactive oxygen species) are ubiquitously present. But excess generation of free radicals cause depletion of immune system antioxidants, alter in gene expression and induce abnormal proteins and contribute to more than one hundred disorders in humans including atherosclerosis, arthritis, ischemia and reperfusion injury of many tissues, central nervous system injury, gastritis, cancer and AIDS. Antioxidant can protect the body by preventing the formation of free radicals, by bringing interruption in Free radicals attack, by scavenging the reactive metabolites or by converting them to less reactive molecules (Hegde and Joshi, 2009). We evaluated the antioxidant effect of AC considering these mentioned parameters.

Polyphenolic phytochemicals, flavonoids and phenol have been reported from AC which is suggested to have multiple biological effects, including antioxidant activity (Kahkonen *et al.*, 1999). Phenolic compounds are implicit to induce the cellular antioxidant system by approximately 50% cellular glutathione concentration increasement. Flavonoids are significant in the modulation of γ -glutamylcysteine synthase in both cellular antioxidant defenses and detoxification of xenobiotics (Muchuweti *et al.*, 2007). We have got phenol content 38.38 ± 2.01 mg/g which may be the cause for the antioxidant activity of AC in different models. The proton-donating ability of this extract was evaluated through DPPH assay and reducing ability. IC₅₀ value of AC close to standard ascorbic acid in DPPH assay was found very promising indicating its capacity of having scavenged the free radicals efficiently.

Reducing power of any extract will be given by the amount of reductones present in them. The ability of the hydroxyl groups present in the flavonoids / phenolics to reduce the free radicals by donating their electrons will determine their activity. Dose dependent reducing ability of any extract exerted antioxidant action by breaking the free radical chain by donating hydrogen atom (Duh *et al.*, 1999). So, these antioxidant potentiality of AC are an important approach for the management of oxidative stress ailment.

The brine shrimp lethality bioassay is very useful to assess the bioactivity of the plant extracts which in most cases correlates reasonably well with cytotoxic and anti-tumor properties (McLauglin *et al.*, 1993). LC₅₀ values of AC revealed its considerable cytotoxic potency. Huge amount of phenolics and flavonoids present in AC might be responsible for its promising cytotoxic activity (Okwori *et al.*, 2007; Moreira *et al.*, 2007) and the possible mechanism of cytotoxicity against brine shrimp nauplii due to poisonous effect on cell mitosis.

CONCLUSION

This work has demonstrated that the methanolic extracts of *Ageratum conyzoides* stems possesses promising antioxidant and cytotoxic potentiality, thereby lends support to the traditional use of the plant in infectous and inflammatory disorders. However, further studies are needed to be conducted to understand the exact mechanisms of such actions and to isolate the active principles responsible for the observed activity.

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