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In-vitro antibacterial activity of herbal aqueous extract against multi-drug resistant Klebsiella sp. isolated from human clinical samples

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ABSTRACT

The studies were carried out to evaluate antibacterial activity of 35 aqueous herbal extracts against a total of 20 clinical *Klebshiella* sp. isolates. The maximum antibacterial activity was found as 90% in crude extracts of *Syzygium aromaticum* (leaf) and *Citrus limon* L. (fruit) followed by 85% in *Spondias pinnata* (leaf). Sensitivity of these isolates was also evaluated for eight commercial antibiotic discs following disc diffusion assay where most of the isolates found to develop resistance against multiple commercial antibiotics. 85% of isolates exhibited resistant to chloramphenicol and erythromycin and 80% were resiatant to sulfamethoxazole and cephradine. The isolates showed their resistance between 55-60 % to the other four antibiotic discs, *viz*; gentamycin, streptomycin, ciprofloxacin and azithromycin. Among 35 herbal extracts tested, 19 herbal extrats were found to possess antimicrobial activity in all multi-drug resistant isolates. Therefore these herbal extracts could be used in future direction as alternative therapeutic agents for the treatment of human diseases caused by *Klebsiella* sp.

Key Words: Klebshiella sp., herbal extract, multidrug resistant, antibacterial activity.

INTRODUCTION

Among the human pathogenic bacteria Klebsiella sp. are very notable. They are gram-negative, nonmotile, encapsulated, lactose fermenting, facultative anaerobic and rod shaped bacterium found in the normal flora of the mouth, skin, and intestines (Ryan and Ray, 2004). Species of Klebsiella are ubiquitous, found naturally in the soil, water and vegetables. In humans, they cause pneumonia (inflammatory illness of the lungs), urinary tract infections (UTI), septicemia, soft tissue infections and abdominal infections (Podschun and Ullmann, 1998). They are opportunistic pathogen and under certain conditions may cause serious infections (Wei et al., 2008). Antibiotics are often used against diseases caused by Klebsiella sp. But these pathogens are becoming increasingly antibiotic-resistant, so that many are now labeled as "Multidrug-resistant (MDR) Klebshiella sp. (Yan et al., 2001; Ktari et al.,

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2006). Thus it is an important task for the researcher to find out alternative medicine. Antimicrobials of plant origin have enormous therapeutic potential. Advantages of herbal drugs include true improvement of disease condition. Even in some medical practice herbal treatments are being used. So that Plants based antimicrobials represent a vast untapped source for medicines and further exploration of plant antimicrobials needs to occur. Herbs can be very effective in programs for resolving urinary tract infections and typhoid fever (Tambekar, 2010). Thus, herbal treatment would promise a greater viable solution for effective treatment of diseases caused by bacteria (Khan et al., 2007; Rahman and Hossain, 2010). The efficacy of herbal extract over the multidrug resistant (MDR) Klebshiella sp. isolates has been investigated in the present study.

MATERIALS AND METHODS

Collection of Bacterial Isolates

Human clinical isolates *Klebsiella* sp. were collected from Popular Diagnostic Center, Sylhet branch and Sylhet Osmani Medical College and aseptically transferred to the USDA Project Laboratory of the

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Table 1: List of medicinal plants used to evaluate antibacterial activity.

| Sl. No. | Local name | English Name | Botanical name | Parts of plant used | | | |
|---------|------------|-----------------------|----------------------------|---------------------|--|--|--|
| 01. | Amloki | Amla | Phyllanthus emblica L. | Fruit | | | |
| 02. | Arjun | Arjuna | Terminalia arjuna | Leaf | | | |
| 03. | Amra | Hog plum | Spondias pinnata | Leaf | | | |
| 04. | Lobongo | Clove | Syzygium aromaticum | Fruit | | | |
| 05. | Lebu | Lemon | Citrus limon L. | Fruit | | | |
| 06. | Daruchini | Cinnamon | Cinnamomus Zeylanicum | Bark | | | |
| 07. | Eucalyptus | Eucalyptus | Eucaplytus globules | Leaf | | | |
| 08. | Kalijira | Black cumin | Nigella sativa L. | Seed | | | |
| 09. | Rosun | Garlic | Allium sativum | Bulb | | | |
| 10. | Arjun | Arjuna | Terminalia arjuna | Bark | | | |
| 11. | Jolpai | Olive | Olea europaea L. | Leaf | | | |
| 12. | Peyaj | Onion | Allium cepa | Bulb | | | |
| 13. | Dumur | Common fig | Ficus carica | Leaf | | | |
| 14. | Paan | Betel leaf | Piper betle L. | Leaf | | | |
| 15. | Jarul | Banaba Plant | Lagerstroemia speciosa | Leaf | | | |
| 16. | Chalta | Chalta | Dillenia indica L. | Leaf | | | |
| 17. | Mehedi | Henna | Lawsonia inermis L. | Leaf | | | |
| 18. | Ada | Ginger | Zingiber officinale | Rhizome | | | |
| 19. | Lojjaboti | Mimosa | Mimosa pudica | Leaf | | | |
| 20. | Tejpata | Indian bay leaf | Cinnamomum tamala | Leaf | | | |
| 21. | Bel | Bael | Aegle marmelos | Leaf | | | |
| 22. | Lebu | Lemon | Citrus limon L. | Leaf | | | |
| 23. | Pepe | Papaya | Carica papaya | Leaf | | | |
| 24. | Helencha | Spinach | Atternatherna phloxeroides | Leaf | | | |
| 25. | Chatni | Palm-leaf Marshmallow | Althaea cannabina | Leaf | | | |
| 26. | Allamanda | Allamanda | Allamanda schottii | Leaf | | | |
| 27. | Thankuni | Asiatic Pennywort | Centella asiatica | Leaf | | | |
| 28. | Dhonia | Coriander | Coriandrum sativum L. | Leaf | | | |
| 29. | Casava | Casava | Manihot esculenta | Leaf | | | |
| 30. | Tulsi | Tulsi /HolyBasil | Ocimum sanctum | Leaf | | | |
| 31. | Horitoki | Black Myrobalan | Terminalia Chebula | Leaf | | | |
| 32. | Joba | China rose | Hibiscus rosa-sinensis L. | Leaf | | | |
| 33. | Tetul | Tamarind | Tamarindus indica L | Leaf | | | |
| 34. | Cha | Tea | Camellia sinensis | Leaf | | | |
| 35. | Bhringraj | False Daisy | Eclipta alba (L.) Hassk. | Leaf | | | |

Department of Genetic Engineering and Biotechnology, Shahjalal University of Science and Technology for further studies. Twenty clinical isolates of presumptive *Klebsiella* sp. were collected during March, 2011. Presumptive *Klebsiella* sp. isolates were obtained from urine samples of patiences who claimed fever, pneumonia and urinary tract infection (UTI). All presumptive *Klebsiella* sp. isolates were collected in chromogenic media plates, as blue-purple mucoid colony. After collection of all the isolates, were labeled, sub cultured and stored at -20°C for further use.

Identification of Klebsiella sp.

A series of morphological, physiological and biochemical tests were performed to identify the suspected *Klebsiella* sp isolates. The test included Gram staining, motility, oxidase activity, catalase production, acid production in glucose, oxidation-fermentation (OF) test (glucose, lactose and sucrose fermentation), Voges-Proskauer Test (VP) and Hydrogen Sulfide Production. All tests were conducted according to The Bergey's Manual of Determinative Bacteriology (Bergey and John, 1994).

Antibiotic sensitivity of the isolates

The antibiotic sensitivity test was performed

following the disc diffusion technique as described by Rahman and Hossain 2010. The nutrient broth culture containing desirable bacterial culture was incubated at 37°C for 24-48 hours in a shaking incubator. After obtaining suitable growth of bacterial culture by observation of turbidity, 50µl of broth culture were dropped in the nutrient agar plate. Culture was spread by sterile "L" shaped glass rod for the preparation of spread plate culture to set the antibiotic discs. The anti-microbial discs were dispensed onto the surface of the inoculated agar plate. Eight commercially prepared antibiotics discs viz., gentamicin (10µg/disc), ciprofloxacin (20µg/disc), streptomycin (10µg/disc), chlorampheazithromycin (25µg/disc) (30µg/disc), sulphamethoxazole (25µg/disc), erythromycin (15µg/disc), and cephradine (30µg/disc) manufactured by Oxoid Ltd. and gentamycin (10µg/disc) manufactured by Becton Disc kinson & company were placed on the surface of the medium with sterile forceps and pressed gently to ensure good contact with the surface of the medium. The plates were inverted and placed in an incubator at 37°C within 15 minutes after the discs were applied. After 16 to 18 hours of incubation, each plate was examined for the determination of the zone of inhibition of the antibiotics. The anti-microbial activities of discs were determined by measuring the zone of inhibition expressed in millimeter.

Antibacterial activity of herbal extract to Klebsiella sp.

A total of 35 herbal extracts prepared from 33 plants were used in this study to screen their antibacterial activity to the Klebsiella sp (Table 1). Most of the herbs were collected from different parts of Sylhet district. The fresh parts of plants such as young leaves, bark, bulb, root, flower, rhizome or petiole were collected and washed several times with distilled water. The plant parts were cut into small pieces and paste was made by using mortar-pestle. Approximately 10µl of individual herb extract was inoculated onto spread plate culture of the Klebshilla sp. isolates. The plate was then allowed to incubate at 37°C for overnight. After 12-24 h of incubation, the herb extract was noted for zone of inhibition for each Klebsiella sp. isolates. The diameter of the herb extracts and the diameter of the zone of inhibitions were measured by measuring scale in millimeter (mm). The ratio between the diameters was calculated.

Table 2: Biochemical characteristics of Presumptive *Klebshiella* sp.

| Biochemical characteristics | Isolate No. K1-K20 | | | | | | |
|-----------------------------|--------------------|--|--|--|--|--|--|
| of isolates | | | | | | | |
| Gram staining | - | | | | | | |
| Grow Aerobically | + | | | | | | |
| Grow Anaerobically | + | | | | | | |
| O-F | F | | | | | | |
| Motility | - | | | | | | |
| Lactose fermentation | + | | | | | | |
| Sucrose fermentation | + | | | | | | |
| Glucose fermentation | + | | | | | | |
| Indole test | - | | | | | | |
| H ₂ Sproduction | - | | | | | | |
| MR | - | | | | | | |
| VP | + | | | | | | |

Note: + = Positive, - = Negative, F= Fermentative

RESULTS AND DISCUSSION

The study was performed to investigate the antibacterial properties of some medicinal plant extracts to *Klebshiella* sp. isolates. All of the presumptive isolates that exhibited the colony characteristics similar to *Klebsiella* sp. isolates were sub-cultured and assessed for their morphological, physiological and biochemical characteristics. All bacterial isolates were Gram negative, facultative anaerobes, and negative in Indole test. The isolates were nonmotile, lactose fermenter, positive in VP test and negative in MR test and able to ferment glucose and sucrose but did not produce H₂S in TSI. Based on all of the above tests isolates were identified to belong to the genus *Klebshiella* (**Table 2**).

The Klebshiella sp. isolates showed variable result in their antibiotic sensitivity pattern against eight commercial antibiotic discs tested. Most of the isolates exhibited resistant to multiple commercial antibiotics and refereed as "Multidrug-resistant organisms" (MDROs). Antibiotics often used against diseases caused by Klebsiella sp. But multidrugresistant (MDR) strains of Klebsiella sp. are now encountered frequently and the rates of multiple drug-resistances have increased considerably in recent years. In reviewed study it is available that the frequency of ESBL producing Klebshiella sp. were found to be resistant against ciprofloxacin (74.8%) cephalosporin (23%) (Podschun et al., 1998; Alipourfard and Yeasmin, 2010). In the present study, 85% of Klebsiella sp. isolates were resistant to chloramphenicol and erythromycin followed by 80% to

Table 3: Antimicrobial activity of aquas herbal extract against Klebshiella sp. isolates.

| Name of harden | Name of isolates with their sensitivity pattern (K1-K20) | | | | | | | | | | | | | | | | | | | |
|--------------------------|--|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| Name of herbs | K1 | K2 | КЗ | K4 | K5 | K6 | K7 | K8 | K9 | K10 | K11 | K12 | K13 | K14 | K15 | K16 | K17 | K18 | K19 | K20 |
| Spondias pinnata | 2.90 | 1.44 | 2.42 | 2.21 | 2.8 | - | 2.85 | 2.75 | 2.95 | 2.75 | 2.29 | 2.29 | 2.75 | 2.88 | 2.45 | - | 2.60 | 2.00 | 2.90 | - |
| Allium sativum | 2.55 | - | - | 2.75 | - | - | 2.80 | 1.95 | - | 2.65 | 1.88 | - | 2.00 | 2.25 | 2.00 | - | 2.29 | 2.77 | 2.52 | 2.05 |
| Terminalia arjuna (leaf) | 1.44 | - | - | 2.75 | - | 1.80 | - | 1.73 | - | 1.95 | 1.45 | 1.95 | 1.75 | 1.36 | 1.20 | 1.20 | 1.55 | 2.00 | 1.36 | 1.70 |
| Citrus limon L. (fruit) | 3.00 | 4.00 | 4.60 | 4.20 | 3.80 | 4.00 | - | 4.40 | 4.00 | 4.40 | 3.40 | 2.90 | 2.90 | 3.00 | - | 3.60 | 3.60 | 3.60 | 3.20 | 2.40 |
| Tamarindus indica L | - | 2.16 | 2.36 | - | 2.43 | 2.25 | - | - | - | 2.30 | 2.40 | 2.33 | - | 2.25 | 2.14 | 2.76 | 2.45 | 2.47 | 2.33 | 2.20 |
| Phyllanthus emblica L. | 2.44 | 2.25 | - | 2.44 | 2.40 | - | - | 2.90 | 2.30 | 2.36 | 2.44 | 2.70 | - | 2.40 | 2.70 | 2.20 | 2.70 | 2.00 | 3.22 | 2.50 |
| Syzygium aromaticum | 2.50 | 2.20 | 2.38 | 2.44 | - | 2.30 | 2.44 | - | 2.50 | 2.30 | 2.50 | 2.95 | 2.40 | 2.35 | 2.40 | 2.70 | 2.30 | 2.35 | 2.50 | 2.45 |
| Olea europaea L. | - | - | 2.50 | 3.00 | 2.90 | 1.40 | - | - | 1.50 | 2.25 | 2.75 | - | - | 2.90 | 2.75 | 2.95 | 2.45 | 2.60 | 1.90 | 2.85 |
| Terminalia arjuna (bark) | 2.50 | - | - | 2.90 | - | - | - | - | - | - | - | 1.65 | 1.40 | 2.33 | - | 1.85 | 1.95 | - | - | 2.05 |
| Eucaplytus globules | 2.44 | 2.58 | - | 2.87 | 2.33 | - | - | - | 2.33 | 1.33 | 2.40 | - | - | 2.44 | 2.56 | 2.44 | 2.30 | 2.35 | 2.45 | 2.44 |
| Nigella sativa L. | 2.50 | - | 2.44 | 2.57 | 2.33 | 2.59 | 2.44 | 1.75 | - | 2.45 | 2.51 | 2.25 | - | 2.55 | - | 1.95 | - | 2.58 | 1.80 | 2.73 |
| Cinnamomus Zeylanicum | - | 1.45 | 1.55 | 1.39 | - | 1.20 | - | 1.80 | 1.35 | 1.25 | - | - | - | - | - | 1.33 | - | - | 1.30 | 1.45 |
| Allium cepa | - | - | 1.55 | 1.70 | - | 1.25 | - | - | - | 1.28 | 1.36 | 1.45 | 1.50 | 1.20 | - | - | 1.30 | 1.45 | - | 1.40 |
| Cinnamomum tamala | 2.42 | 2.20 | 2.40 | 2.44 | - | - | - | 2.13 | - | 2.40 | 2.20 | 2.15 | - | - | - | - | - | - | - | - |
| Camellia sinensis | 3.5 | 2.25 | 2.22 | 2.15 | - | 2.15 | - | - | 2.25 | - | - | - | - | 2.55 | 1.65 | - | 2.5 | - | - | 1.25 |
| Citrus limon L. (leaf) | - | - | 1.95 | - | - | 1.15 | - | - | - | 1.25 | - | - | - | - | - | - | - | 1.70 | - | - |
| Polygonum tomentosum | - | 1.45 | - | 1.33 | - | 1.88 | - | - | - | 1.33 | 1.25 | - | 1.05 | 1.19 | - | 1.25 | - | - | - | - |
| Ficus carica | - | - | - | - | - | 2.00 | - | - | - | - | - | - | 1.95 | - | - | 1.85 | - | - | - | - |
| Lagerstroemia speciosa | 1.33 | - | - | - | - | - | - | - | - | 1.41 | - | - | - | - | - | 1.19 | - | 1.41 | 1.33 | - |

sulfamethoxazole and cephradine. 55-60% of *Klebsiella* sp. isolates were resistance to streptomycin, azithromycin, gentamicin and ciprofloxacin **(Figure 1)**.

The prevalence of resistant Klebsiella sp. is significant and deserves more consideration. To overcome all of these constrains now we are taking shelter to our ancestor's medicinal practice. According to former customs we are taking interest about herbs. Reviewed studies stated that enormous work has done to screen the antibacterial activity of herbs against human pathogen (Jahan et al., 2007; Khan et al., 2007; Rahman et al., 2008; Misra et al., 2009). Among the 35 samples tested in the present study, 19 crude herb extracts were found to exibit their antibacterial properties against Klebshiella sp. isolates. viz. Spondias mombin L., Allium sativum, Terminalia arjuna (leaf), Citrus limon L. (fruit), Tamarindus indica L, Phyllanthus emblica L., Syzygium aromaticum, Olea europaea L., Terminalia arjuna (bark), Eucaplytus globules, Nigella sativa L., Cinnamomus zeylanicum, Allium cepa, Cinnamomum Camellia sinensis, Citrus limon L. (fruit), Polygonum tomentosum, Ficus carica and Lagerstroemia speciosa. The maximum antibacterial activity were found up to 90% in Citrus limon L. (fruit) and Syzygium aromaticum (leaf) followed by 85% activity were found in Spondias pinnata (leaf) (Table 3).

In recent years, work is being done with organic solvent to find out which solvent is more efficient. Phytochemical studies are also going on with important medicinal herbs to analyze their pharmacological properties. There is strong need to develop alternate antimicrobial drugs for the treatment of diseases caused by *Klebsiella* sp. Because of huge emergence of multi-drug resistant (MDR) bacteria it is an urgent need to discover new therapeutics that

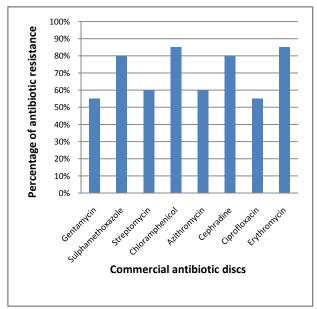


Figure 1: Percentage of Resistance of Klebshiella sp. isolates to commercial antibiotics.

would be effective against MDR strain. This study suggested that herbs with unique chemical compounds that can either inhibit the growth of pathogens or kill them considered as potential candidates for developing new antimicrobial drugs.

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