



Original Article

Stromal CD10 as a Tumor Microenvironment Immunomarker in Colorectal Carcinoma: Association with Clinicopathological Parameters

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Abstract

Background: Colorectal carcinoma (CRC) is the third most common malignancy worldwide, posing a major public health challenge. Tumor-stromal interactions are increasingly recognized as potential therapeutic targets. CD10 has been implicated in tumor aggressiveness, with altered expression linked to the development of various cancers. This study aimed to evaluate stromal CD10 expressions in colorectal carcinoma and its association with clinicopathological parameters. **Materials and Methods:** A cross-sectional observational study was conducted on 50 colorectal carcinoma cases at the Department of Pathology, Dhaka Medical College, from March 2022 to February 2024. Purposive sampling was used, and immunohistochemistry for stromal CD10 expression was performed. Formalin-fixed, paraffin-embedded tissue from histologically confirmed cases was analyzed. Data was entered, cleaned, and analyzed using SPSS version 25.0. **Results:** Tumors were most frequently located in the left colon (58%), with a mean size of 5.1±2.8 cm; 72% showed exophytic growth, and most were staged as T3 (42%). Nodal metastasis and lymphovascular invasion were observed in 50% and 28% of cases, respectively, while 70% were low grade. Stromal CD10 expression was detected in 48% of tumors. Higher CD10 positivity was seen among patients aged 41-60 years, females, and smokers. Although no significant associations were found with most tumor parameters, CD10 expression was significantly associated with lymphovascular invasion (71.4% vs. 38.9%, $p<0.05$). **Conclusion:** The findings of this study suggest that stromal CD10 may have a role in colorectal carcinoma progression, particularly in relation to lymphovascular invasion.

Keywords: Clinicopathological parameters, CD10, Colorectal cancer, Stromal expression.

Received: August 01, 2025; **Accepted:** September 15, 2025

doi <https://doi.org/10.3329/emcj.v11i1.89992>



Introduction

Colorectal carcinoma (CRC) ranks as the third most frequently identified cancer globally and stands as the second leading cause of cancer-associated fatalities worldwide¹. The anticipated rise in the global incidence of colorectal carcinoma is projected to elevate from 1.8 million new cases annually to 2.2 million by the year 2030². In Bangladesh, colorectal cancer ranks 7th in prevalence and 10th in cancer-related mortality³. CRC commonly presents with altered bowel habits, blood in stool, abdominal discomfort, unexplained weight loss, fatigue, and a persistent sense of incomplete evacuation². Its risk increases with age, particularly after 50, and is further influenced by genetic predispositions (e.g., Lynch syndrome, FAP), a personal history of inflammatory bowel disease, and familial occurrence. Lifestyle factors-

including a diet rich in red and processed meats, low fiber intake, physical inactivity, obesity, and use of tobacco or alcohol- also significantly contribute to disease susceptibility⁴. CRC is classified based on cellular origin and tumor characteristics. Adenocarcinoma accounts for over 90% of cases, while less common types include carcinoid tumors, gastrointestinal stromal tumors (GISTs), lymphomas, and sarcomas⁵. The World Health Organization (WHO) currently classifies colorectal adenocarcinoma into low-grade (formerly well to moderately differentiated) and high-grade (formerly poorly differentiated) tumors. This simplified system is clinically relevant, as low-grade tumors generally carry a more favorable prognosis, while high-grade tumors exhibit aggressive behavior and poorer outcomes⁶. Previously, WHO categorized

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colorectal adenocarcinoma into three histological grades based on glandular differentiation: Grade 1 (well-differentiated), with >95% gland formation and close resemblance to normal epithelium, associated with favorable prognosis; Grade 2 (moderately differentiated), showing 50-95% glandular differentiation with partial architectural distortion; and Grade 3 (poorly differentiated), characterized by <50% gland formation, marked atypia, and aggressive clinical behavior⁷.

At the time of diagnosis, over 20% of patients present with metastatic (stage IV) CRC.⁸ The exact mechanisms underlying cancer metastasis remain elusive, yet research indicates that it arises from intricate interactions between tumor cells and their surrounding tumor microenvironment⁹. Many molecules are secreted by tumor cells or by stromal cells facilitate tumor invasion and metastasis¹⁰. One such molecule is Matrix Metalloproteinase (MMP), that has influence in cancer progression and in defining the role of stromal microenvironment in invasiveness and metastatic potential¹¹. MMPs play a multifaceted role in cancer progression by facilitating extracellular matrix degradation, inhibiting apoptosis, promoting angiogenesis, and modulating the immune response, thereby supporting tumor invasion and metastasis¹². Cluster of Differentiation 10 (CD10) is a membrane-bound, zinc-dependent metalloprotease expressed in epithelial cells of the prostate, colon, liver, stomach, breast, lung, endometrium,¹³⁻¹⁵ renal tubular brush borders and various tumor & stromal cells¹⁶.

It plays a pivotal role in peptide metabolism through the degradation of key signaling molecules such as enkephalins, bradykinin, and selected cytokines¹⁷. Initially identified in leukemia, CD10 has since emerged as a significant immunohistochemical marker in various malignancies, including breast and colorectal carcinomas^{18,19}. In CRC, CD10 expression- particularly within stromal cells- has been linked to tumor progression, aggressiveness, and differentiation status²⁰. Despite surgical resection being the mainstay of CRC treatment, high recurrence and metastasis rates persist, with nearly 60% of patients affected postoperatively and 20% dying from disease recurrence²¹. Current therapies largely target epithelial tumor cells, often neglecting the genetically stable stromal compartment, which may contribute to therapeutic failure and relapse¹⁰. Given CD10's expression in tumor-associated stroma, presents a promising diagnostic and therapeutic target. The anti-CD10 monoclonal antibody JMAM-1 has demonstrated antitumor activity in mesothelioma, suggesting potential applications in CRC²². This study investigates stromal CD10 expression in CRC and its association with clinicopathological features, aiming to

elucidate its role in tumor invasiveness and to explore its viability as a therapeutic target.

Materials and Methods

Study design and settings: This cross-sectional observational study was conducted to evaluate clinicopathological and immunohistochemical parameters in colorectal adenocarcinoma. The study period spanned March 2022 to February 2024 and included 50 patients who underwent colorectal tumor resection at the Department of Surgery, Dhaka Medical College Hospital (DMCH) and were histologically confirmed as colorectal adenocarcinoma. Patients who had received preoperative chemotherapy or radiotherapy, or who presented with completely necrotic or non-carcinomatous tumors, were excluded.

Data collection procedure: Histopathological analysis was performed at the Department of Pathology, Dhaka Medical College, while immunohistochemical (IHC) staining was carried out at Primate Diagnostic Center, Dhaka. Clinical data- including age, gender, education, and family history were collected using structured proformas through patient or attendant interviews and review of hospital records. Tumor-related variables assessed included size, location, growth pattern, grade, depth of invasion, nodal metastasis, and lymphovascular invasion (LVI). Tumor location was classified as right colon (caecum to transverse colon) or left colon (descending colon to rectum). Tumor size was categorized as ≤3 cm, 3.1-6 cm, and >6 cm. Gross morphology was classified as exophytic or endophytic. Tumor grading followed WHO criteria: low grade (well to moderately differentiate) and high grade (poorly differentiated). Depth of invasion was staged T1-T4 according to penetration into surrounding structures. Nodal involvement was recorded as metastatic or non-metastatic, and LVI was documented as present or absent.

Paraffin-embedded tumor blocks were retrieved from archival pathology samples. Sections (4 μm) were stained with hematoxylin and eosin for routine histopathology. For CD10 IHC, antigen retrieval was followed by HRP-based EnVision staining. Stromal CD10 expression was semi-quantitatively scored, with ≥5% positivity considered positive. Tonsil tissue served as the positive control.

Statistical analysis: Data was entered, cleaned, and analyzed using SPSS version 25.0. Descriptive statistics were expressed as frequencies and percentages. The Chi-square (χ^2) test was used to assess associations. A p-value <0.05 at a 95% confidence interval was considered statistically significant.

Ethical approval: Informed written consent was obtained from all participants. Data confidentiality was maintained, and unauthorized access was strictly prohibited. The Ethical approval was obtained from the Ethical Review Committee of Dhaka Medical College, and written informed consent was secured from all participants (Reference: ERC-DMC/ECC/2022/397).

Results

In the Table-I, the study population ranged in age from 14 to 83 years (mean: 50.3 ± 14.3 years) and was stratified into three age groups, with the 41-60-year category representing the majority (54.0%). A slight female predominance was observed (M:F=1:1.2). A positive family history was reported in 10.0% of cases, while 32.0% had a history of smoking.

Among all colorectal carcinoma cases, tumors were most commonly located in the left colon (58%), including the descending colon, sigmoid colon, and rectum. Tumor size ranged from 1.5 to 13.0 cm, with a mean diameter of 5.1 ± 2.8 cm. More than half (54%) measured 3.1-6.0 cm, while smaller tumors (≤ 3 cm) occurred in 24% and larger tumors (> 6.0 cm) in 22%, indicating relatively fewer cases at the extremes of size. Exophytic growth was the predominant pattern (72%). Most tumors were staged as T3 (42%), followed by T2 (34%) and T4 (24%), with no T1 lesions detected. Nodal metastasis was present in 50% of cases, and lymphovascular invasion in 28%. The majority of tumors were low grade (70%). Stromal CD10 expressions were observed in 48% of cases. (Table-II) The present study examined the association between stromal CD10 expression and clinico-demographic variables. Higher CD10 positivity was observed among patients aged 41-60 years (59.3%), females (51.9%), and smokers (62.5%), whereas lower expression rates were noted in younger individuals, males, and those with a positive family history (Table-III).

Table-IV shows that most associations were statistically non-significant ($p > 0.05$). CD10 expression was evaluated in relation to various tumor parameters in colorectal carcinoma. Higher CD10 positivity was observed in left colonic tumors (51.7%) compared with proximal colonic lesions (42.9%). Tumors measuring 3.1-6.0 cm demonstrated the highest expression (55.6%), while larger tumors (> 6.0 cm) showed lower expression (36.4%) ($p = 0.495$). Growth pattern variations were not associated with CD10 expression ($p = 0.650$). Although CD10 expression was most frequent in T4 lesions (58.3%), the difference across stages was not significant ($p = 0.670$). Cases with nodal metastasis showed slightly higher expression (52.0%) compared with node-negative tumors (44.0%). A

statistically significant association was observed with lymphovascular invasion, with 71.4% positivity in LVI-positive cases versus 38.9% in LVI-negative cases ($p = 0.039$). While expression was higher in high-grade tumors (53.3%), no significant association was found across WHO grades ($p = 0.621$).

Table-I: Socio-demographic outlines of cases (n=50)

Characteristics	Frequency (n)	Percentage (%)
Age (years)		
≤40	10	20.0
41-60	27	54.0
>60	13	26.0
Mean±SD	50.3 ±14.3	
Gender		
Male	23	46.0
Female	27	54.0
Family history		
Positive	5	10.0
Negative	45	90.0
History of smoking		
Yes	16	32.0
No	34	68.0

Table-II: Cases according to their tumor parameters (n=50)

Traits	Frequency (n)	Percentage (%)
Tumor location		
Left colon	29	58.0
Right colon	21	42.0
Tumor size		
≤ 3.0 cm	12	24.0
3.1-6.0 cm	27	54.0
> 6.0 cm	11	22.0
Mean ± SD	5.1±2.8	
Growth pattern		
Exophytic	36	72.0
Endophytic	14	28.0
Extent of tumor		
T2	17	34.0
T3	21	42.0
T4	12	24.0
Lymph node metastasis		
Yes	25	50.0
No	25	50.0
Lymphovascular invasion		
Yes	14	28.0
No	36	72.0
Grade of tumor		
Low grade	35	70.0
High grade	15	30.0
Expression of CD10		
Positive	24	48.0
Negative	26	52.0

Table-III: Association of CD10 expression with socio-demographics of the cases (n=50)

Socio-demographic factors of the cases (n=30)			
Attributes	CD10 expression		p-value
	Positive	Negative	
	n (%)	n (%)	
Age			
≤40	3 (30.0)	7 (70.0)	0.245
41-60	16 (59.3)	11 (40.7)	
>60	5 (38.5)	8 (61.5)	
Gender			
Male	10 (43.5)	13 (56.5)	0.555
Female	14 (51.9)	13 (48.1)	
Family history			
Positive	2 (40.0)	3 (60.0)	1.000
Negative	22 (48.9)	23 (51.1)	
Smoking history			
Yes	10 (62.5)	6 (37.5)	0.159
No	14 (41.2)	20 (58.8)	

p-values were obtained by Fisher's exact test

Table-IV: Association of CD10 expression with different clinicopathological parameters of tumor (n=50)

Tumor (n=56)		CD10 expression		P-value
Attributes	Positive	Negative		
	n (%)	n (%)		
Tumor location				
Left colon	15 (51.7)	14 (48.3)	0.536	
Right colon	9 (42.9)	12 (57.1)		
Tumor size (in cm)				
≤3.0	5 (41.7)	7 (58.3)	0.495	
3.1-6.0	15 (55.6)	12 (44.4)		
> 6.0	4 (36.4)	7 (63.6)		
Tumor growth pattern				
Exophytic	18 (50.0)	18 (50.0)	0.650	
Endophytic	6 (42.9)	8 (57.1)		
Tumor extent				
T2	7 (41.2)	10 (58.8)	0.670	
T3	10 (47.6)	11 (52.4)		
T4	7 (58.3)	5 (41.7)		
Lymph node metastasis				
Yes	13 (52.0)	12 (48.0)	0.571	
No	11 (44.0)	14 (56.0)		
Lymphovascular invasion				
Yes	10 (71.4)	4 (28.6)	0.039	
No	14 (38.9)	22 (61.1)		
WHO grading				
Low grade	16 (45.7)	19 (54.3)	0.621	
High grade	8 (53.3)	7 (46.7)		

p-values were obtained by Chi Square test

Discussion

This cross-sectional study was conducted to evaluate stromal CD10 expression in colorectal adenocarcinoma and its association with clinicopathological variables. The mean age of patients was 50.3±14.3 years, with 54% in the 41-60 year group, which also demonstrated the highest

CD10 positivity (59.3%). These findings are comparable to studies in India, Austria, and Iraq^{23,24}, but contrast with higher mean ages reported in Iran (60.2±13.9) and Greece (70.9±9.3)²⁵. Female patients constituted 54% of the sample and exhibited greater CD10 expression (51.9%) than males (43.5%). Comparable trends were reported in another study in Africa²⁶, while opposite patterns were observed in UK and Pakistani cohorts^{27,28}. Although family history and smoking were not significantly associated with CD10 positivity, a slight reduction in expression was noted among those with a positive family history²⁹. Tumor in left colon showing higher CD10 expression (51.7%) compared to proximal ones (42.9%), aligning with findings by Granados-Romero et al. (2017).³⁰ The mean tumor size was 5.1±2.8 cm, with CD10 positivity peaking in tumors sized 3.1-6.0 cm, similar to finding the studies^{31,32}.

Current study found positivity of stromal CD10 expression higher (50%) in tumors with exophytic growth patterns while lower (42.9%) in tumor with endophytic growth pattern, suggesting a correlation between CD10 and protruding morphology^{33,34}. Stromal CD10 expression increased with tumor depth, highest in T4 lesions (58.3%), and was more frequent in cases with lymph node metastasis (52%) and lymphovascular invasion (71.4%, p= 0.039). CD10 was positive in 48% of cases. Its absence in adjacent non-neoplastic mucosa suggests a role in tumor microenvironment remodeling, cellular signaling disruption³⁴ and disease progression⁷, indicating its potential prognostic significance in colorectal carcinoma.

Conclusion

Stromal CD10 expression was observed in nearly half of colorectal adenocarcinoma cases and showed a significant association with lymphovascular invasion, suggesting a potential role in tumor aggressiveness. No significant correlation was found with other clinicopathological parameters. Assessment of stromal CD10 may aid in identifying tumors with higher invasive potential. Larger, multicenter studies are recommended to further validate its prognostic significance and potential as a therapeutic target.

Acknowledgments

The authors are thankful to all the participants and hospital authorities for their earnest cooperation.

Funding

This study did not receive any funds.

Conflict of interest

The authors declared that they have no conflicts of interest.

References

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin.* 2021; 71 (3): 209-49. doi: 10.3322/caac.21660.
2. Arnold M, Sierra MS, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global patterns and trends in colorectal cancer incidence and mortality. *Gut.* 2017; 66 (4): 683-91. doi: 10.1136/gutjnl-2015-310912.
3. Warnakulasuriya S. Global epidemiology of oral and oropharyngeal cancer. *Oral Oncol.* 2009; 45 (4-5): 309-16. doi: 10.1016/j.oraloncology.2008.06.002.
4. Ribeiro Franco PI, Rodrigues AP, de Menezes LB, Pacheco Miguel M. Tumor microenvironment components: Allies of cancer progression. *Pathol Res Pract.* 2020; 216 (1): 152729. doi: 10.1016/j.prp.2019.152729.
5. Reynolds I, Healy P, Mcnamara DA. Malignant tumours of the small intestine. *Surgeon.* 2014; 12 (5): 263-70. doi: 10.1016/j.surge.2014.02.003.
6. Ueno H, Kajiwarra Y, Shimazaki H, Shinto E, Hashiguchi Y, Nakanishi K, et al. New criteria for histologic grading of colorectal cancer. *Am J Surg Pathol.* 2012; 36 (2): 193-201. doi: 10.1097/PAS.0b013e318235edee.
7. Fleming M, Ravula S, Tatishchev SF, Wang HL. Colorectal carcinoma: Pathologic aspects. *J Gastrointest Oncol.* 2012; 3 (3): 153-73. doi: 10.3978/j.issn.2078-6891.2012.030.
8. Farouk S, Khairy A, Salem AM, Soliman AF, Bader El Din NG. Differential Expression of miR-21, miR-23a, and miR-27a, and Their Diagnostic Significance in Egyptian Colorectal Cancer Patients. *Genet Test Mol Biomarkers.* 2020; 24 (12): 825-34. doi: 10.1089/gtmb.2020.0184.
9. Basbınar Y, Calıbası-Kocal G. Cancer Metastasis. *Intech Open*; 2018. Available at: <http://dx.doi.org/10.5772/intechopen.74633>. [Accessed on August 12, 2023]
10. Ulaganathan S. An Analysis of Correlation of Stromal CD10 Expression in Carcinoma Breast NOS Type with ER, PR and HER2/Neu. *Int J Adv Res.* 2018; 10 (7): 212-7.
11. Curran CS, Keely PJ. Breast tumor and stromal cell responses to TGF- β and hypoxia in matrix deposition. *Matrix Biol.* 2013; 32 (2): 95-105. doi: 10.1016/j.matbio.2012.11.016.
12. Quintero-Fabián S, Arreola R, Becerril-Villanueva E, Torres-Romero JC, Arana-Argáez V, Lara-Riegos J, et al. Role of Matrix Metalloproteinases in Angiogenesis and Cancer. *Front Oncol.* 2019; 9: 1370. doi: 10.3389/fonc.2019.01370.
13. Wang S, Xiao Y, An X, Luo L, Gong K, Yu D. A comprehensive review of the literature on CD10: its function, clinical application, and prospects. *Front Pharmacol.* 2024; 15: 1336310. doi: 10.3389/fphar.2024.1336310.
14. Maguer-Satta V, Besançon R, Bachelard-Cascales E. Concise review: neutral endopeptidase (CD10): a multifaceted environment actor in stem cells, physiological mechanisms, and cancer. *Stem Cells.* 2011; 29 (3): 389-96. doi: 10.1002/stem.592.
15. Song J, Aumüller G, Xiao F, Wilhelm B, Albrecht M. Cell specific expression of CD10/neutral endopeptidase 24.11 gene in human prostatic tissue and cells. *Prostate.* 2004; 58 (4): 394-405. doi: 10.1002/pros.10345.
16. Jana SH, Jha BM, Patel C, Jana D, Agarwal A. CD10-A new prognostic stromal marker in breast carcinoma, its utility, limitations and role in breast cancer pathogenesis. *Indian J Pathol Microbiol.* 2014; 57: 530-6. doi: 10.4103/0377-4929.142639.
17. Iwaya K, Ogawa H, Izumi M, Kuroda M, Mukai K. Stromal expression of CD10 in invasive breast carcinoma: a new predictor of clinical outcome. *Virchows Arch.* 2002; 440 (6): 589-93. doi: 10.1007/s00428-002-0639-4.
18. Kadota K, Buitrago D, Lee MC, Villena-Vargas J, Sima CS, Jones DR, et al.. Tumoral CD10 expression correlates with high-grade histology and increases risk of recurrence in patients with stage I lung adenocarcinoma. *Lung Cancer.* 2015; 89 (3): 329-36. doi: 10.1016/j.lungcan.2015.06.003.
19. Makretsov NA, Hayes M, Carter BA, Dabiri S, Gilks CB, Huntsman DG. Stromal CD10 expression in invasive breast carcinoma correlates with poor prognosis, estrogen receptor negativity, and high grade. *Mod Pathol.* 2007; 20 (1): 84-9. doi: 10.1038/modpathol.3800713.
20. Jang TJ, Park JB, Lee JI. The Expression of CD10 and CD15 Is Progressively Increased during Colorectal Cancer Development. *Korean J Pathol.* 2013; 47 (4): 340-7. doi: 10.4132/KoreanJPathol.2013.47.4.340
21. Miyata Y, Kumagai K, Nagaoka T, Kitaura K, Kaneda G, Kanazawa H, et al.. Clinicopathological significance and prognostic value of Wilms' tumor gene expression in colorectal cancer. *Cancer Biomark.* 2015; 15 (6): 789-97. doi: 10.3233/CBM-150521.
22. Mizutani N, Abe M, Kajino K, Matsuoka S. A New CD10 Antibody Inhibits the Growth of Malignant Mesothelioma. *Monoclon Antib Immunodiagn Immunother.* 2021; 40 (1): 21-7. doi: 10.1089/mab.2020.0033.
23. Steele G Jr, Posner MR. Adjuvant treatment of colorectal adenocarcinoma. *Curr Probl Cancer.* 1993; 17 (4): 223-69.

24. Gill MK, Manjari M, Jain K, Kaur TA. Expression of Her-2/neu in colon carcinoma and its correlation with the histological grades and the lymph nodes status. *J Clin Diagn Res*. 2011; 5 (8): 1564-8.
25. Bebars SM, Mohammed HS, Osman GS, Sakr MA. Immunohistochemical expression and prognostic value of epidermal growth factor receptor 1 and human epidermal growth factor receptor 2/neu in primary colorectal carcinoma. *Egypt J Pathol*. 2021; 41 (1): 41-8.
26. Duduyemi BM, Andoh D, Adankwah E, Nyarko H, Agyemang D. The use of special stains in the detection of vascular invasion in cases of colon cancer in resource-limited settings in Africa. *Ann Trop Pathol*. 2020; 11 (1): 21-4. doi: 10.4103/atp.atp_1_20.
27. Abdulkader M, Abdulla K, Rakha E, Kaye P. Routine elastic staining assists detection of vascular invasion in colorectal cancer. *Histopathology*. 2006; 49 (5): 487-92. doi: 10.1111/j.1365-2559.2006.02533.x.
28. Tahir SK. Accuracy of elastic tissue stain in detecting venous invasion in colorectal cancer. *J Islamabad Med Dent Coll*. 2015; 4 (1): 31-4.
29. Slattery ML, Levin TR, Ma K, Goldgar D, Holubkov R, Edwards S. Family history and colorectal cancer: predictors of risk. *Cancer Causes Control*. 2003; 14 (9): 879-87. doi: 10.1023/b:caco.0000003840.94591.76.
30. Granados-Romero JJ, Valderrama-Treviño AI, Contreras-Flores EH, Barrera-Mera B, Herrera Enríquez M, Uriarte-Ruiz K, et al. Colorectal cancer: a review. *Int J Res Med Sci*. 2017; 5 (11): 4667-76. doi: 10.18203/2320-6012.ijrms20174914.
31. Seo AN, Kwak Y, Kim DW, Kang SB, Choe G, Kim WH, et al. HER2 status in colorectal cancer: its clinical significance and the relationship between HER2 gene amplification and expression. *PLoS One*. 2014; 9 (5): e98528. doi: 10.1371/journal.pone.0098528.
32. Kornprat P, Pollheimer MJ, Lindtner RA, Schlemmer A, Rehak P, Langner C. Value of tumor size as a prognostic variable in colorectal cancer: a critical reappraisal. *Am J Clin Oncol*. 2011; 34 (1): 43-9. doi: 10.1097/COC.0b013e3181cae8dd.
33. Oliveira LA, Artigiani Neto R, Joppert Netto G, Sznirer M, Fernandes LC, Lima FD, et al. Tissue expression of CD10 protein in colorectal carcinoma: correlation with the anatomopathological features of the tumor and with lymph node and liver metastases. *J Coloproctol (Rio J)*. 2012; 32: 34-9. doi: 10.1590/S2237-93632012000100005.
34. Ogawa H, Iwaya K, Izumi M, Kuroda M, Serizawa H, Koyanagi Y, et al. Expression of CD10 by stromal cells during colorectal tumor development. *Hum Pathol*. 2002; 33 (8): 806-11. doi: 10.1053/hupa.2002.125773.

Citation of this article

Rashid HO, Jeba R, Yeamin MA, Zannat T, Sultana S, Tahsin KN, Akter J, Nurunnabi M. Stromal CD10 as a Tumor Microenvironment Immunomarker in Colorectal Carcinoma: Association with Clinicopathological Parameters. *Eastern Med Coll J*. 2026; 11 (1): 14-9.

doi: <https://doi.org/10.3329/emcj.v11i1.89992>