EFFECTS OF 2, 3, 5-TRIIODOBENZOIC ACID ON THE ANATOMICAL CHANGES OF CHICKPEA VAR. BARI CHOLA-7

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The use of synthetic plant growth regulators to regulate plant growth has become an important tool in many agricultural and horticultural practices. Among the synthetic plant growth regulators, TIBA, a polar auxin transport inhibitor is widely used. At low concentration, it is capable of checking the excessive vegetative growth and lodging tendency, reducing the abscission of flowers and immature pods, and modifying the crop canopy to improve the productivity of crops⁽¹⁾.

Physiological modifications as well as improved yield and yield attributes caused by TIBA have been reported in various economically important crop plants *viz.* soybean⁽²⁾, groundnut⁽³⁾, tomato⁽⁴⁾ and chickpea⁽⁵⁾. While the effects of TIBA on the external appearance of plants are well known, limited information is available as to its effects upon the internal structure⁽⁶⁾.

Chickpea (*Cicer arietinum* L.) is one of the major pulse crops in Bangladesh and is the third most important food legume grown in the world after beans and peas. It is one of the important sources of protein in the dietary uptake of the people of Bangladesh. It is ranked fifth in area but second in consumption priority⁽⁷⁾. The present investigation was undertaken to study the effects of TIBA on the anatomical structure of root and shoot in chickpea.

A field experiment was carried out at the research field of the Department of Botany, University of Dhaka during Rabi (December-March) season of 2017-2018. Seeds of BARI Chola-7 were collected from BARI (Bangladesh Agricultural Research Institute), Joydebpur, Gazipur. The experimental soil was analyzed and low levels of potassium and very low levels of nitrogen and phosphorus were recorded⁽⁸⁾. Cowdung, urea, TSP and MP were applied at basal amount during final land preparation. The experiment was laid out in randomized block design with four replications. The unit plot size was 2 m × 1.1 m. Seeds were sown on 29th November, 2017 in rows having a gap of 40 cm. Plants in rows were maintained 15 cm apart by thinning seedlings at 15 days after sowing. Intercultural operations were done as and when necessary. Five foliar treatments, *viz.* water spray (control), 10, 20, 50 and 100 ppm TIBA, respectively, were tested against

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chickpea var. BARI Chola-7. The foliar spray was done at 45 days after sowing by using separate sprayers. Three weeks after treatment, control plants and 20 ppm TIBA treated plants which showed positive physiological effects were collected and preserved in FAA (formalin-acetic acid-alcohol) solution for anatomical study. Free hand cross sections of both root and shoot were taken and stained in safranin. Root sections were taken 0.5 cm below the root collar and stem sections were taken 4 cm above the soil line. Photomicrograph of the stained specimens were taken using Trinocular Res. microscope with digital camera system (Axiocam ERc 5s, Zeiss, Germany).

Effects on anatomy of stem: 20 ppm TIBA treated plants showed an increase (Table 1, Fig. 1A, B) in diameter of the stem compared to the control though pith diameter showed a marginal decrease. Similar results were also reported by Berova and Zlatev⁽⁹⁾ in tomato and by Tsegaw et al.⁽¹⁰⁾ in potato. Due to 20 ppm TIBA, significant changes occurred in the number and size of xylem vessels. In TIBA treated plants, number of vessels in stem increased considerably though their size or average diameter (0.50 \pm 0.30) of the cavity decreased compared to the control plants. In addition, the xylem vessels are more uniformly distributed throughout the diameter of the stem of TIBA treated plants than those of control. Research workers reported different results in different plant species with respect to TIBA and other auxin transport inhibitor induced stem anatomical modifications. Kroll and Moore⁽⁶⁾ reported decrease in the number of xylem vessels in soybean stem following TIBA application at 70 g/h. However, Robnett and Morey⁽¹¹⁾ observed an increased number of xylem elements in 2,4-D and 2,4-T treated plant stems. Mattsson et al.(12) noticed increased number of vascular tissues in the NPA (1-Nnaphthylphtalamic acid)-treated plants. Krause⁽¹³⁾ reported production of small vessels in soybean following TIBA application.

| Diameter | Control plants | Treated plants |
|------------------|-----------------|-----------------|
| Stem (mm) | 2.08 ± 0.20 | 2.22 ± 0.30 |
| Stem pith (µm) | 5.80 ± 0.70 | 5.60 ± 0.60 |
| Stem vessel (µm) | 0.60 ± 0.20 | 0.50 ± 0.30 |
| Root (mm) | 3.50 ± 0.30 | 4.13 ± 0.30 |
| Root pith (µm) | 3.40 ± 0.40 | 4.20 ± 0.20 |
| Root vessel (µm) | 0.80 ± 0.30 | 0.70 ± 0.30 |

Table 1. Effect of TIBA on stem and root characteristics. Mean value of four observations ± standard deviation.

Effects on anatomy of root: In plants receiving 20 ppm TIBA, diameter of both root and pith increased (Table 1, Fig. 2A, B) noticeably compared to the control. Moreover, diameter of root was greater than that of stem in both control and TIBA treated plants.



Fig. 1. A, B. Transverse section (50× magnification) of the stem of the control (A) and 20 ppm TIBA treated plants (B) of *Cicer arietinum* L. var. BARI Chola-7. P = pith, XV = xylem vessel, E = epidermis.



Fig. 2. A-F. Transverse section of the root of the control (A,C,E) and 20 ppm TIBA treated plants (B, D, F) of *Cicer arietinum* L. var. BARI Chola-7. P = pith, XV = xylem vessel, E = epidermis. A, B (50× magnification); C, D (100× magnification); E, F (400× magnification).

Increased root diameter following growth retarding chemical application was also reported in other plants, *viz.* soybean⁽¹⁴⁾ and potato⁽¹⁰⁾. Burrows *et al.*⁽¹⁵⁾ found reduced stem diameter but roots with increased diameter in chrysanthemum due to similar growth retarding chemical PBZ. The most significant changes that occurred due to 20 ppm TIBA treatment was in the number and sizes of the xylem vessels. The xylem vessel diameter in TIBA-treated roots decreased compared to the control (Fig. 2C-F). But, 20 ppm TIBA induced the formation of more secondary xylem vessels in root and they were evenly distributed within the radii because of continuous production. This result is also in consistent with the findings of Tsegaw *et al.*⁽¹⁰⁾ in potato.

Galston⁽¹⁶⁾ suggested that TIBA antagonizes and may completely negate the effect of IAA. It may do this by competing for a carrier or by accelerating the destruction of IAA. Kamien and Skoog⁽¹⁷⁾ and Hay⁽¹⁸⁾ concluded that low concentrations of TIBA primarily affected the polarity of auxin, rather than the destruction of it. Aberg⁽¹⁹⁾ indicated that at high concentration, TIBA antagonized the effect of externally applied auxin and apparently also the natural auxin and caused a growth inhibition. However, at low amounts TIBA had weak auxin effects which might be due to a synergistic action upon residual amounts of the native auxin. Therefore, depending on the plant species and the concentration applied, TIBA may either stimulate or inhibit plant growth, thereby affecting morphological and anatomical modifications. The study confirms that TIBA treatment can induce similar anatomical modifications in a wide range of crops. The effect of TIBA on the induction of anatomical changes may be mediated by changing the hormonal balance of the plant.

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