

Screening Rice Germplasm against Sheath Blight Disease of Rice and its Integrated Management in Bangladesh

S Parveen^{1*}, M A Ali² and M A Ali³

ABSTRACT

Fifty-seven rice germplasm collected from BRRI Genebank were screened against sheath blight (ShB) by artificial inoculation in field and laboratory conditions in T. Aman 2012. Significant differences on relation to lesion height (RLH) among the germplasm were observed, where the highest (83%) was recorded in susceptible check, BR11 and the lowest (8.33%) was in Orgoja. Severity score of ShB was recorded maximum (9) in Dudhsail, Basi, Chaula mari, Holdemota, Calendamota, Semmua, Kotijira, Halisail, Horakani, Kalisura, Ashfuli, Huglapata and BR11 as highly susceptible to ShB, whereas it was minimum (1) in Orgoja. Gopal ghosh was observed as moderately tolerant with 27.33% RLH and severity score 3, while Kala binni, Khazur chari, Binni, Kalagora, Patjait and Dorkumur found moderately tolerant with severity score 5. In detached sheath inoculation method in test tube, most of the germplasms found highly susceptible, except Orgoja as resistant and Gopal ghosh as moderately tolerant. However, Orgoja showed resistance in both field inoculation and detached sheath inoculation methods. But, Dorkumur was found moderately tolerant in field and highly susceptible in detached sheath inoculation in laboratory. The experiment of Integrated Disease Management (IDM) packages was conducted in the farmer's field with BR11 at Fulpur, Mymensingh during T. Aman 2013. The IDM practices of rice ShB resulted profound effect. Relative lesion height, percent disease index, tiller infection and hill infection were maximum (68%, 69%, 86% and 79% respectively) in T₆ (control) and minimum in T₁ [FDR (removal of floating debris) + 30 July transplanting + Potash (K) fertilizer (202 g decimal⁻¹) + Top dressing of urea (247 kg ha⁻¹) in four equal splits at 15 days interval + single spray of fungicides of Azoxystrobin 10% (0.17 kg ha⁻¹) + Tebuconazole 90% (500 ml ha⁻¹)]. Moreover, the highest number of panicles per m², filled grains per panicle and grains yield were recorded in T₁ (160, 150 and 6.25 t ha⁻¹ respectively) and the minimum in T₆ (227, 120 and 3.6 t ha⁻¹ respectively). Therefore, the best IDM package was T₁ for its effective control of ShB disease as well as yield maximization of rice. Finally, Orgoja could be used in resistance breeding for varietal improvement and the IDM package of T₁ need to be recommended to prescribe in the farmer's field after simulation in different AEZs and seasons with different varieties of Bangladesh.

Key words: Germplasm, resistance, integrated management, sheath blight, rice

INTRODUCTION

Bangladesh agriculture involves food production for 163.65 million people (Salam *et al.*, 2014), where rice is the principal food. This increasing population requires increasing crop yields for stable supply of grain to achieve food security of the country. Consequently, the national average production needs to be increased from 3 to 5 t ha⁻¹ in next 20 years (Mahbub *et al.*, 2001). In Bangladesh, rice production area is 11.01 million hectares of land during 2016-17 (BBS, 2018). However,

36.27 million metric tons of rice is produced in the country during 2017-18 (AIS, 2019). Sheath blight (ShB) of rice was first reported in Japan by Miyakie in 1910. It is caused by *Rhizoctonia solani* Kuhn. It is considered as the most damaging major epidemic disease of rice (Li *et al.*, 2012). ShB is an important disease of rice, especially in intensive rice production systems. The average incidence of ShB in Bangladesh is about 20.3% (Ali *et al.*, 2003). The yield loss caused by ShB in Bangladesh ranged from 14 to 31% under farmer's field (Shahjahan *et al.*, 1986). The presence of one or many factors

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may enhance the severity of ShB beyond economic threshold levels, thereby incurring low to high yield losses.

Incidence and development of ShB of rice depends on climate, host and soil factor (Damicone *et al.*, 1993). Short duration and semi-dwarf cultivars are more susceptible to ShB (Groth and Lee, 2002). During rice ShB epidemics, severe lodging may occur (Wu *et al.*, 2012). Differences in yield loss between very susceptible and moderately resistant cultivars are substantial. On infection by *Rhizoctonia solani*, semi-dwarf varieties show more than twice the reduction in yield and milling quality.

Breeding for resistance against ShB has not been successful due to lack of sources of resistant genes (Rao, 1995; Hashiba and Kobayashi, 1996). Resistance source against ShB disease of rice is not available in Bangladesh and anywhere (Jalal Uddin *et al.*, 2000). Consequently, none of the high yielding varieties is resistant to ShB disease neither in Bangladesh nor elsewhere in the world. Fortunately, rice land races have proven to be highly adaptive to diverse environmental conditions and are believed to harbour a number of valuable genetic resources for crop improvement (Karmakar *et al.*, 2012; Roychowdhury *et al.*, 2013; Ganie *et al.*, 2014). Some of the landraces such as Buhjan, Banshpata, Bhasamanik, Nagra Sail, Raghu Sail are tolerant to rice ShB (Dey, 2014). Therefore, local or land races of rice need to be exploited for getting resistant or moderately resistant or even better tolerant sources for ShB disease.

The control of ShB in the field so far is mainly relied on the use of fungicides, which is not sustainable for its residual effect along with the potential risk of resistant to fungicides overtime. Disease management programme against ShB can concentrate

different approaches such as incorporating cultural practices, exploitation of host resistance, biological control with *Trichoderma harzianum* and *Trichoderma viride* and chemical control. Ashrafuzzaman *et al.* (2005) also reported that emphasis should be given on different management options to control ShB disease of rice. For clean cultivation, burning the crop residues, destroy grasses and other hosts from the field, collecting and burying floating debris after final land preparation may reduce infection foci. Instead of applying excess dose of nitrogen, split application of K fertilizer with last top dress of urea can reduce its infestation. Application of 40 kg MP/ha as top dress in two equal splits and transplanting with 20 cm × 20 cm spacing have affect on ShB (Hossain and Mia, 2001). Large amount of N and phosphate (P) is favourable for ShB disease (Dasgupta, 1992) and high potash (K) or PK is useful for infection (CRRI, 1977). Therefore, the present research programme was planned and designed to develop management technologies of the disease with the aim of recommending suitable control strategies in Bangladesh. The present study was under taken to screen germplasm for their reaction to ShB and to develop an integrated management practice for controlling ShB of rice in Bangladesh.

MATERIALS AND METHODS

Screening of rice germplasm against ShB of rice

Rice germplasm. A total of 57 rice germplasm collected from BRRRI Genebank were screened against ShB disease of rice in the field through hill inoculation method and BR11 was used as susceptible check (Table 1).

Table 1. Primary information of the germplasms used for screening resistance source against sheath blight.

Acc. no.*	Variety	Acc. no.	Variety	Acc. no.	Variety
4111	Gopal ghosh	4794	Kalahati	5221	Kalisura
4112	Chata bazail	4795	Khajur chhori	5222	Akra
4113	Ram dash	4849	Rayeda	5223	Ushi har
4114	Paizra	5121	Jamni	5250	Ashfuli
4118	Kala binni	5122	Chaula maghi	5286	Ranisalut
4149	Beto	5190	Bushi hara (mota)	5289	Buripagli
4155	Chini kani	5192	Lohamugra	5298	Harisankar
4156	Minki	5193	Chaula mari	5300	Birinde
4162	Kasrail	5194	Kalagora	5310	Orgoja
4163	Khazur chari	5195	Patjait	5316	Nonamurchi
4239	Binni	5196	Holdemota	5319	Gandhakusturi
4267	Birpala	5197	Kanchachikon	5327	Huglapata
4271	Rayda	5198	Dholeswar mota	5329	Gota
4272	Dhaki rayda	5199	Calendamota	5330	Dorkumur
4768	Kaijhuri	5212	Semmua	5337	Changi
4773	Dudhsail	5213	Kotijira	5345	Rasasail
4777	Kashra	5217	Ashkor	5347	Sackhorkhana
4778	Katarangi	5218	Baskor	--	BR11
4792	Basi	5219	Halisail		
4793	Sada pankach	5220	Horakani		

* BRRI Genebank accession number.

Field experiment. The experiment was conducted at the experiment field of Bangladesh Rice Research Institute (BRRI), Gazipur during T. Aman 2012. A levee was made surrounding plots to maintain standing water up to 5.0 cm inside. Land was prepared 15 days before transplanting/seedling. Ploughing and cross ploughing followed by laddering was done by power tiller. Weeds were cleaned manually. The seedlings of the tested germplasms were raised in plastic tray in the Plant Pathology net house. Thirty-day-old 2-3 seedlings per hill were transplanted with a spacing of 20 cm × 15 cm. Fertilizers were applied @ 405: 150: 202: 135: 10 g decimal⁻¹ of urea, TSP, MOP, gypsum and zinc sulphate. All fertilizers were applied in basal, except urea (Anonymous, 2010). For agronomic, weed management, irrigation and drainage and insect management current standard recommendations were followed (Anonymous, 2007).

Preparation of inoculum. One hundred PDA plates in glass petridishes were prepared following the standard procedure. The fungus (*Rhizoctonia solani*) was grown in the petridishes containing PDA medium and

incubated for seven days at room temperature (25 to 30°C) for growth and development of the pathogen.

Inoculation of pathogen. Inoculations were done at maximum tillering stage (Bhaktavatsalam *et al.*, 1978). Two methods of inoculation were employed for inoculation of germplasms by *Rhizoctonia solani*. After seven days of inoculation lesion length and leaf sheath length were measured and calculated. The methods were as follows:

a. Hill inoculation-Total hill were inoculated with *Rhizoctonia solani* Kuhn culture (7 days) grown on PDA medium. Prior to inoculation, eight hills were tagged randomly in the central area of each plot in the field for inoculation. Inoculation was done by inserting a piece of culture medium (cutting the culture medium into eight pieces) at the middle of each hill in the afternoon, colonized by the ShB pathogen in a tagged rice hill and maintained standing water onward of the crop growth to maintained high moisture below canopy level for disease development (Sharma and Teng, 1990).

b. Detached sheath inoculation-Detached sheath was inoculated in moist test tube (Fig. 1). In detached sheath inoculation method, one tiller from each entry was taken *i.e.* three tillers for three replications. Tillers were cut in such a way that leaf sheath did not separate from stem or remain in contact with stem and uniform in size. Water soaked cotton was placed at the bottom of the test tube and then placed 6-9 mm mycelial block (growing pathogen) inside the sheath. The test tube was then plugged with soaked cotton.

Data recording. The disease severity was recorded from the data collected from 25 hills in each replication of each treatment. Severity was calculated by relative lesion height (RLH) (McKinney, 1923). Data were recorded for each treatment following standard evaluation system (SES) for rice in 0-9 scale (Anonymous, 1996). Data of the lesion height, plant height, 1000 grain weight and grain yield (g hill⁻¹) were also recorded. In detached sheath inoculation method, ShB severity was measured by RLH using the following formula-

$$RLH = \frac{\text{Lesion height (cm)}}{\text{Leaf sheath height (cm)}} \times 100$$

Integrated management of ShB of rice

Field experiment. The experiment was conducted in the farmer's field with BR11 at Fulpur, Mymensingh during T. Aman 2013. Plant to plant spacing was 15 cm and row to row distance was 16 cm. Randomized RCBD was used with four replications. Plot size was



Fig. 1. Detached sheath inoculation method of screening against ShB of rice.

2.5 m × 4 m. Plot to plot distance was 0.5 m and block to block distance was 1 m. The best options obtained from the results of different experiments (Parveen, 2016) were included into integrated disease management (IDM) packages and were simulated in the field. The treatments used in this study were shown below:

T₁=FDR (removal of floating debris) + 30 July planting + Potash (K) fertilizer (202 g decimal⁻¹) + Top dressing of urea (247 kg ha⁻¹) in four equal splits at 15 days interval + single spray of fungicide [Azoxystrobin 10% (0.17 kg ha⁻¹) + Tebuconazole 90% (500 ml ha⁻¹)]. T₂= 30 July planting + K-dose + top dressing of urea in four equal splits at 15 days interval + single spray of fungicide. T₃= K-dose + top dressing of urea in four equal splits at 15 days interval + single spray of fungicide. T₄= Top dressing of urea in four equal splits at 15 days interval + single spray of fungicide. T₅= Single spray of fungicide. T₆= Control.

Inoculation of pathogen. Same as hill inoculation method.

Data collection. Twenty-five hills were selected at random from each experimental unit. Number of infected tillers and hills were counted. Incidence was recorded by tiller infection and expressed in percentage, while severity by relative lesion height (RLH) and percent disease index (PDI) (McKinney, 1923). Data were recorded for each treatment following standard evaluation system (SES) for rice in 0-9 scale (Anonymous, 1996). Data on total tiller, infected tiller, plant height, panicle per m², filled grain, unfilled grain, 1000 grain weight (TGW) and grain yield were also recorded. PDI was measured by using the following formula-

$$PDI = \frac{\text{Total rating}}{\text{No. of observation} \times \text{Maximum grade}} \times 100$$

Statistical analysis. The data were subjected to statistical analysis and ANOVA (analysis of variance) were constructed following RCBD by SPSS 2.05 programme for both the experiments. The treatment means were compared by LSD test at probability level P=0.05.

RESULTS AND DISCUSSION

Assessment of germplasm against ShB of rice

Table 2 shows that there was a variation among the germplasms on ShB disease development and yield through hill inoculation in the field. Significant differences on RLH among the germplasms were observed. The highest RLH was recorded in BR11 (83%) and the lowest was in Orgoja (8.33%). The maximum (9) severity (SES) score

of ShB was recorded in Dudhsail, Basi, Chaula mari, Holdemota, Calendamota, Semmua, Kotijira, Halisail, Horakani, Kalisura, Ashfuli, Huglapata and BR11, which were highly susceptible (HS) to ShB disease, whereas the minimum severity score (1) was observed in Orgoja. Gopal ghosh was observed as moderately tolerant to ShB disease with 27.33% RLH and severity score 3. Moreover, Kala binni, Khazur chari, Binni, Kalagora, Patjait and Dorkumur found moderately tolerant to ShB with severity score 5. On the other hand, the highest yield was found in Beto (18.23 g hill⁻¹), Rayda (18.15), Ushi har (18.23) and Buripagli (18.15) and the lowest in Kashra, Calendamota, Orgoja and Sackhorkhana (4.85 g hill⁻¹) germplasms (Table 3).

Table 2. Reaction of screened germplasm against ShB due to artificial inoculation of *Rhizoctonia solani* through hill inoculation method in the field.

Acc. no.	Variety	Growth duration	Plant height (cm)	RLH (%)	SES score	Reaction
4111	Gopal ghosh	150	131	27.33	3	MT
4112	Chata bazail	151	140	47.66	7	HS
4113	Ram dash	152	144	54.00	7	HS
4114	Paizra	149	127	63.00	7	HS
4118	Kala binni	151	129	38.00	5	MT
4149	Beto	155	154	53.00	7	HS
4155	Chini kani	147	141	61.66	7	HS
4156	Minki	156	141	61.33	7	HS
4162	Kasrail	154	141	53.66	7	HS
4163	Khazur chari	148	141	41.33	5	MT
4239	Binni	147	137	43.66	5	MT
4267	Birpala	141	136	54.33	7	HS
4271	Rayda	149	136	50.33	7	HS
4272	Dhaki rayda	146	150	60.00	7	HS
4768	Kaijhuri	142	119	56.33	7	HS
4773	Dudhsail	154	149	69.00	9	HS
4777	Kashra	145	147	51.66	7	HS
4778	Katarangi	145	151	64.66	7	HS
4792	Basi	140	115	75.33	9	HS
4793	Sada pankach	138	149	53.66	7	HS
4794	Kalahati	143	149	62.33	7	HS
4795	Khajur chhori	142	150	56.66	7	HS
4849	Rayeda	145	152	56.33	7	HS
5121	Jamni	147	150	64.66	7	HS
5122	Chaula maghi	149	144	63.33	7	HS
5190	Bushi hara (mota)	150	153	57.00	7	HS
5192	Lohamugra	149	150	55.33	7	HS

Table 2. Continued.

Acc. no.	Variety	Growth duration	Plant height (cm)	RLH (%)	SES score	Reaction
5193	Chaula mari	145	151	72.66	9	HS
5194	Kalagora	149	141	42.33	5	MT
5195	Patjait	149	152	45.00	5	MT
5196	Holdemota	150	146	68.66	9	HS
5197	Kanchachikon	153	156	64.66	7	HS
5198	Dholeswar mota	154	165	60.33	7	HS
5199	Calendamota	155	161	66.33	9	HS
5212	Semmua	152	142	69.33	9	HS
5213	Kotijira	150	134	70.00	9	HS
5217	Ashkor	149	146	55.33	7	HS
5218	Baskor	150	158	49.33	7	HS
5219	Halisail	148	149	66.00	9	HS
5220	Horakani	148	166	67.33	9	HS
5221	Kalisura	149	144	74.33	9	HS
5222	Akra	148	174	54.00	7	HS
5223	Ushi har	152	144	52.66	7	HS
5250	Ashfuli	161	98	66.66	9	HS
5286	Ranaisalut	165	147	59.00	7	HS
5289	Buripagli	163	165	58.33	7	HS
5298	Harisankar	153	164	51.33	7	HS
5300	Birinde	157	150	64.66	7	HS
5310	Orgoja	160	160	8.33	1	R
5316	Nonamurchi	155	152	55.00	7	HS
5319	Gandhakusturi	152	139	65.00	7	HS
5327	Huglapata	154	147	73.33	9	HS
5329	Gota	151	152	57.66	7	HS
5330	Dorkumur	159	153	41.66	5	MT
5337	Changi	151	151	55.66	7	HS
5345	Rasasail	159	113	62.33	7	HS
5347	Sackhorkhana	153	128	53.66	7	HS
--	BR11	145	115	83.00	9	HS

LSD (P=0.05)

MT=Moderately tolerant, HS=Highly susceptible, R=Resistant.

Table 3. Yield and 1000 grain weight (TGW) of screened germplasms against ShB due to artificial inoculation of *Rhizoctonia solani* through hill inoculation in the field.

Acc. no.	Variety	TGW (g)	Yield (g hill-1)
4111	Gopal ghosh	20.13	6.92
4112	Chata bazail	21.14	8.17
4113	Ram dash	24.63	9.05
4114	Paizra	25.05	9.60
4118	Kala binni	29.11	10.05
4149	Beto	20.38	18.23
4155	Chini kani	9.19	5.30
4156	Minki	29.27	6.32
4162	Kasrail	26.14	14.55
4163	Khazur chari	21.44	7.24
4239	Binni	10.22	8.22
4267	Birpala	20.33	10.92
4271	Rayda	24.37	18.15
4272	Dhaki rayda	12.40	10.36
4768	Kaijhuri	29.16	10.28
4773	Dudhsail	14.03	10.07
4777	Kashra	16.05	4.85

Table 3. Continued.

Acc. no.	Variety	TGW (g)	Yield (g hill-1)
4778	Katarangi	13.33	8.40
4792	Basi	15.55	10.18
4793	Sada pankaiach	16.26	12.56
4794	Kalahati	12.89	11.03
4795	Khajur chhori	15.19	10.59
4849	Rayeda	12.30	5.82
5121	Jamni	20.49	11.91
5122	Chaula maghi	26.87	16.03
5190	Bushi hara (mota)	27.06	5.55
5192	Lohamugra	27.12	10.17
5193	Chaula mari	21.44	7.24
5194	Kalagora	10.22	8.22
5195	Patjait	20.33	10.92
5196	Holdemota	19.37	10.15
5197	Kanchachikon	12.40	10.36
5198	Dholeswar mota	29.16	10.28
5199	Calendamota	16.05	4.85
5212	Semmua	13.33	8.40
5213	Kotijira	15.55	10.18
5217	Ashkor	16.26	12.56
5218	Baskor	12.89	11.03
5219	Halisail	21.14	8.17
5220	Horakani	24.63	9.05
5221	Kalisura	25.05	9.60
5222	Akra	29.11	10.05
5223	Ushi har	20.38	18.23
5250	Ashfuli	9.19	5.30
5286	Ranisalut	20.33	10.92
5289	Buripagli	24.37	18.15
5298	Harisankar	12.40	10.36
5300	Birinde	29.16	10.28
5310	Orgoja	10.05	4.85
5316	Nonamurchi	12.30	5.82
5319	Gandhakusturi	20.49	11.91
5327	Huglapata	11.87	5.40
5329	Gota	27.06	5.55
5330	Dorkumur	27.12	10.17
5337	Changi	12.40	10.36
5345	Rasasail	29.16	10.28
5347	Sackhorkhana	16.05	4.85
--	BR11	23.98	13.98
LSD (P=0.05)		0.83	0.76

Table 4 shows that Orgoja was resistant against ShB disease of rice with the minimum RLH (11.66%) and severity score (1), whereas Gopal gosh was moderately tolerant to ShB with 40.56% RLH and severity score 5 through detached sheath inoculation method in test tube. But, rest of the germplasms with RLH ranging from 48.33 to 89.66% along with BR11 (90.68%) (Fig. 2) were found highly susceptible against ShB. Comparing

the two inoculation method (*i.e.* hill inoculation and detached sheath inoculation) Orgoja was found as resistant and Gopal gosh as moderately tolerant to ShB disease. In detached sheath inoculation method in test tube, most of the germplasms were found highly susceptible to ShB except Orgoja and Gopal gosh. Dorkumur was found moderately tolerant in field condition but it showed high level of susceptibility to ShB in case of detached sheath

Table 4. Reaction of screened germplasms against ShB due to artificial inoculation of *Rhizoctonia solani* through detached sheath inoculation in test tube.

Acc. no.	Variety	RLH (%)	SES score	Reaction
4111	Gopal ghosh	40.56	5	MT
4112	Chata bazail	70.33	9	HS
4113	Ram dash	60.00	7	HS
4114	Paizra	74.33	9	HS
4118	Kala binni	72.33	9	HS
4149	Beto	82.66	9	HS
4155	Chini kani	61.66	7	HS
4156	Minki	67.33	9	HS
4162	Kasrail	58.00	7	HS
4163	Khazur chari	72.66	9	HS
4239	Binni	78.33	9	HS
4267	Birpala	68.00	9	HS
4271	Rayda	59.66	7	HS
4272	Dhaki rayda	72.33	9	HS
4768	Kaijhuri	63.00	7	HS
4773	Dudhsail	69.00	9	HS
4777	Kashra	53.00	7	HS
4778	Katarangi	57.33	7	HS
4792	Basi	75.33	9	HS
4793	Sada pankach	65.66	9	HS
4794	Kalahati	75.00	9	HS
4795	Khajur chhori	67.33	9	HS
4849	Rayeda	69.66	9	HS
5121	Jamni	64.66	7	HS
5122	Chaula maghi	63.33	7	HS
5190	Bushi hara (mota)	56.00	7	HS
5192	Lohamugra	65.33	7	HS
5193	Chaula mari	72.66	9	HS
5194	Kalagora	65.66	9	HS
5195	Patjait	63.33	7	HS
5196	Holdemota	81.33	9	HS
5197	Kanchachikon	73.66	9	HS
5198	Dholeswar mota	83.00	9	HS
5199	Calendamota	66.33	9	HS
5212	Semmua	78.00	9	HS
5213	Kotijira	76.33	9	HS
5217	Ashkor	55.33	7	HS
5218	Baskor	64.00	7	HS
5219	Halisail	66.00	9	HS
5220	Horakani	77.33	9	HS
5221	Kalisura	74.33	9	HS
5222	Akra	57.33	7	HS
5223	Ushi har	66.00	9	HS
5250	Ashfuli	75.00	9	HS
5286	Ranialut	61.66	7	HS
5289	Buripagli	68.00	9	HS
5298	Harisankar	67.66	9	HS
5300	Birinde	84.66	9	HS
5310	Orgoja	11.66	1	R
5316	Nonamurchi	71.66	9	HS

Table 4. Continued.

Acc. no.	Variety	RLH (%)	SES score	Reaction
5319	Gandhakusturi	64.66	7	HS
5327	Huglapata	76.66	9	HS
5329	Gota	89.66	9	HS
5330	Dorkumur	48.33	7	HS
5337	Changi	72.00	9	HS
5345	Rasasail	62.33	7	HS
5347	Sackhorkhana	57.33	9	HS
--	BR11	90.66	9	HS
LSD (P=0.05)		17.52		

MT=Moderately tolerant, HS=Highly susceptible, R=Resistant.

inoculation method (Fig. 2). In general, dwarf, short duration and photo insensitive varieties were more susceptible to ShB. Prasad and Eizenga (2008) tested 73 *Oryza* genotypes for identifying resistant sources. They found only seven accessions moderately resistant to ShB. On the other hand, Moni (2012) found no resistant variety against ShB.

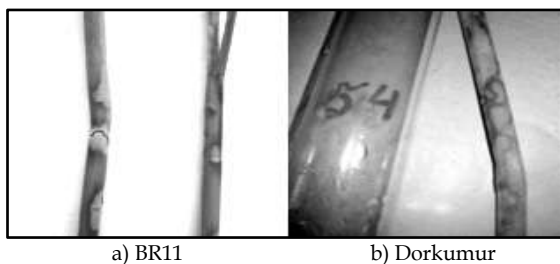


Fig. 2. ShB symptoms of BR11 and Dorkumur due to artificial inoculation of *Rhizoctonia solani* through detached sheath inoculation method in test tube.

Integrated management of ShB of rice

Table 5 shows that the integrated management packages of ShB of rice resulted profound effect. Relative lesion height (RLH) was the maximum (68%) in T₆ (Control). The minimum RLH was 8% in T₁ (FDR + 30 July planting + Potash (K) fertilizer (202 g decimal⁻¹) + top dressing of urea (247 kg ha⁻¹) in four equal splits at 15 days interval + single spray of fungicide) and T₃ (K-dose + top dressing of urea in four equal splits at 15 days interval + single spray of fungicide). RLH was significantly different in different treatment combinations. T₂ (30 July planting + K-dose + top dressing of urea in four equal splits at 15 days interval + single spray of fungicide) and

T₃ (K-dose + top dressing of urea in four equal splits at 15 days interval + single spray of fungicide) significantly differed in RLH. T₄ (Top dressing of urea in four equal splits at 15 days interval + single spray of fungicide) and T₅ (Single spray of fungicide) was different in RLH. Difference between T₃ and T₄ in RLH was also significant. There was significant difference in PDI (Percent disease index) among the treatment combinations. The maximum PDI was 69% in T₆ and the minimum 5% in T₁. T₂ and T₃ also differed significantly. Similarly, PDI of T₄ differed significantly from that of T₅. Moreover, tiller infection was 5% in T₁ which was significantly different from T₂ with 17%. T₃ and T₄ were also different in tiller infection. There was 25% tiller infection in T₄ and 39% in T₅. The maximum tiller infection was 86% in T₆. Besides, hill infection was 79% in T₆ (Control) as compared to 47% in T₅ (Single spray of fungicide). The difference was significant. In T₁ only 3% of the hills became infected, but it was 15% in T₂, 19% in T₃ and 35% in T₄ and all the treatments differed significantly.

Table 5. Effect of integrated disease management (IDM) on ShB of BR11 rice variety during T. Aman 2013.

Treatment	RLH (%)	PDI (%)	Tiller infection (%)	Hill infection (%)
T ₁	8f	5f	5f	3f
T ₂	17e	16e	17e	15e
T ₃	23d	25d	21d	19d
T ₄	36c	39c	25c	35c
T ₅	49b	51b	39b	47b
T ₆	68a	69a	86a	79a

Means followed by the same letter in a column did not differ significantly at the 5% level by LSD.

Table 6 shows that the effect of integrated management of ShB on yield and yield components. The maximum number of panicles per m² was recorded in T₁ (260) and the minimum in T₆ (Control) (227). There was no difference between T₅ (231) and T₆. However, the number of panicles per m² was 251 in T₂, 245 in T₃, 238 in T₄ and 231 in T₅ and all the treatments differed significantly. Number of filled grains per panicle was also significantly different in different treatments. It was 150 and 145 in T₁ and T₂. The minimum number of filled grains per panicle was recorded in T₆ (120) which differed significantly for that in T₅ (125). Significant difference was also observed between T₃ (139) and T₄ (131). Number of unfilled grains was the lowest in T₁ and the maximum in T₆. Significant difference was also observed between T₃ and T₄ as well as T₅ and T₆. Similarly, difference between T₄ and T₅ was also significant in number of unfilled grains per panicle. But there was no effect of integrated management of ShB on grain size. Weight of 1000 grain was 20 g in all treatments. Significant difference was observed between the treatments in grain yield of rice due to integrated management of ShB disease. The maximum yield was recorded in T₁ (6.3 t ha⁻¹) and the minimum in T₆ (3.6 t ha⁻¹). Yield was 6.0 t ha⁻¹ in T₂ as compared to 5.5 t ha⁻¹ in T₃ and the difference was significant. Similarly, T₄ produced 5.2 t ha⁻¹ which was significantly lower than that of T₅ (4.5).

Finally, the present study revealed that the best IDM package was T₁ which included removal of floating debris, transplanting on 30 July, potash (K) fertilizer (202 g decimal⁻¹), urea top dressing (247 kg ha⁻¹) in four equal splits at 15 days interval and single spray of Azoxystrobin (10%) + Tebuconazole (90%) combination. Because, the maximum RLH, PDI, tiller infection and hill infection were found in control plot (T₆), whereas it was lower in the IDM packages and minimum in T₁ plot. Grain yield was also significantly higher

in the IDM plots due to minimum incidence of ShB. Because, ShB was very low and grain yield was maximum in the plots where IDM was applied against ShB of rice due to its trace infection. Therefore, it can be concluded that the IDM package (T₁) though highly effective to control ShB of rice, but the result needs validation across the ecosystem. However, *Rhizoctonia solani* is an universal soil borne facultative and epidemic pathogen. The pathogen is difficult to control unless control measure is taken on time. Many scientists narrated that a single method of control is not effective in most cases to control ShB but IDM is recommended by the researchers (Mew *et al.*, 2004). Host resistance is a sustainable and economic method but there is no such resistant cultivar (Groth *et al.*, 1993). Antagonist such as *Trichoderma* may be a good option to include in IDM package (Dey *et al.*, 2004). ShB infection at flowering stage reduce grain yield due to higher amount of unfilled grains (Cu *et al.*, 1996) as because of damage of leaf sheath by the disease, affect water and nutrients supply to the growing spikelets (Lee and Rush, 1983).

Table 6. Effect of IDM on yield and yield components of BR11 during T. Aman 2013.

Treatment	Panicle per m ²	Filled grain panicle ⁻¹	Sterile pikelet panicle ⁻¹	TGW (g)	Yield (t ha ⁻¹)
T ₁	260a	150a	40f	20	6.25a
T ₂	251b	145b	47e	20	6.00b
T ₃	245c	139c	53d	20	5.52c
T ₄	238d	131c	61c	20	5.15d
T ₅	231e	125d	67b	20	4.49e
T ₆	227e	120e	61a	20	3.60f
Significance	*	*			*
CV (%)	5.15	8.65	18.40	0.0	19.16
LSD 0.05	4.00	3.50	4.90	NS	0.22

Means followed by the same letter did not differ at the 5% level by LSD. NS=Not significant. TGW=1000 grain weight

CONCLUSIONS

ShB of rice is considered as one of the major constraints of rice production in Bangladesh. Almost all HYVs and hybrid varieties are susceptible to the disease. Method for controlling the disease is an urgent need. Among the 57 germplasms, the local cultivar Orgoja (acc. no. 5310) showed resistance to ShB in both hill inoculation in field and detached sheath inoculation in test tube, which could be used in resistance breeding for varietal improvement programme of rice. On the other hand, the best integrated disease management (IDM) package was T₁ which included removal of floating debris, transplanting on 30 July, potash (K) fertilizer (202 g decimal⁻¹), top dressing of urea (247 kg ha⁻¹) in four equal splits at 15 days interval and single spray of Azoxystrobin (10%) + Tebuconazole (90%) combination. Because, ShB was very low and grain yield was high in the plots where T₁ package was applied. Therefore, it can be concluded that the IDM package (T₁) though highly effective to control ShB of rice, but the result needs validation in the farmer's field in different seasons with different rice varieties across the different AEZs of Bangladesh.

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REFERENCES

- AIS (Agricultural Information Service). 2019. Krishi Diary 2019, Agricultural Information Service, Khamarbari, Farmgate, Dhaka 1215, Bangladesh. www.ais.gov.bd.
- Ali, M A, M M Rahman, M A Latif, M Hossain, N R Sharma, S Akter, T A Mia and M A Nahar. 2003. Survey of rice sheath disease caused by different *Rhizoctonia* sp. in Bangladesh. In: Paper presented in the stakeholder workshop on Rice sheath blight disease complex, 3 December 2003. BRRI, Gazipur.
- Anonymous. 1996. Standard Evaluation System for Rice. The International Rice Research Institute, Los Banos, Laguna, Philippines. p. 64.
- Anonymous. 2007. Modern Rice Cultivation (Adhunik Dhaner Chas-Bangla version). Publication no. 5, 13th Edition. Bangladesh Rice Research Institute, Gazipur 1701.
- Anonymous. 2010. Modern Rice Cultivation (Adhunik Dhaner Chas-Bangla version). Publication no. 5, 15th Edition. Bangladesh Rice Research Institute, Gazipur 1701.
- Ashrafuzzaman, M H, M Jalaluddin, M I Khalil and I Hossain. 2005. Integrated management of sheath blight of Aman rice. Bangladesh J. Plant. Pathol. 21 (1 and 2): 53-58.
- BBS. 2018. Statistical Pocket Book Bangladesh 2017. Bangladesh Bureau of Statistics, Statistics and Informatics Division, Ministry of Planning, Govt. of the People's Republic of Bangladesh.
- Bhaktavatsalam, G, K Satyanarayana, A P K Reddy and V T John. 1978. Evaluation for sheath blight resistance in rice. Int. Rice Res. Newsl. 3: 9-10.
- CRRI. 1977. Annual report. Central Rice Research Institute, Cuttack, India.
- Cu, R M, T W Mew, K G Cassman and P S Teng. 1996. Effect of sheath blight on yield in tropical, intensive rice production system (in Chinese). Plant Disease. 80: 1103-1108.
- Damicone, J R, M V Patel and W F Moore. 1993. Density of sclerotia of *Rhizoctonia solani* and incidence of sheath blight in rice fields in Mississippi. Pl. Dis. 77(3): 257-260.
- Dasgupta, M K. 1992. Rice sheath blight: the challenge continues. Plant diseases of International Importance. 1: 130-157.
- Dey, N. 2014. Harnessing the potential of indigenous rice lines: an issue of food sovereignty. In: J A Francis (ed.). CTA 2014. Department of Biotechnology, Visva-Bharati, Santiniketan, India.
- Dey, R, K K Pal, D M Bhatt and S M Chauhan. 2004. Growth promotion and yield enhancement of peanut (*Arachis hypogaea* L.) by application of plant growth-promoting rhizobacteria. Microbiological Research. 159: 371-394.
- Ganie, S A, J Karmakar, R Roy Chowdhury, T K Mondal and N Dey. 2014. Assessment of genetic diversity in salt-tolerant rice and its wild relatives for ten SSR loci and one allele mining primer of *saltol* gene located on 1st chromosome. Plant Systematics and Evolution (DOI 10.1007/s00606-014-0999-7, Published online).

- Groth, D and F Lee. 2002. Rice disease. *In*: W E Smith and R H Dilday (eds.). Rice: Origin, history, technology, and production. John Wiley and Sons, Hoboken, NJ. pp. 413-436.
- Groth, D E, M C Rush, G G Giesler and C A Hollier. 1993. Foliar fungicides for use in the management of rice diseases. *La. Agric. Exp. Stn. Bull.* No. 840.
- Hashiba, T and T Kobayashi. 1996. Rice diseases incited by *Rhizoctonia* species. *In*: B Sneh, S Jabaji-Hare, S Neate and G Dijkstra (eds.). *Klewer Rhizoctonia species: Taxonomy, Molecular Biology, Ecology, Pathology and Disease Control*. Academic Publisher, The Netherlands. pp. 331-340.
- Hossain, M and M A T Mia. 2001. Management of sheath blight disease of rice under farmer's field condition. *Bangladesh J. Plant. Pathol.* 17 (1 and 2): 13-16.
- Jalaluddin, M, M A Kashem, M K Hasna and M I Khalil. 2000. Screening of some somaclonal progenies of rice for resistance to sheath blight and bacterial leaf blight. *Crop Sci.* 11 (1 and 2): 39-42.
- Karmakar, J, R Roychowdhury, R K Kar, D Deb, N Dey and H S Srivastava. 2012. Profiling of selected indigenous rice (*Oryza sativa* L.) landraces of Rarh Bengal in relation to osmotic stress tolerance. *Physiology and Molecular Biology of Plants.* 18(2): 125-132.
- Lee, F N and M C Rush. 1983. Rice sheath blight: A major rice disease. *Plant Disease.* 67 (7): 829-32.
- Li, Di-q, Q-y Tang, Y-b Zhang, J-q Qin, H Li, L-j Chen, S-h Yang, Y-b Zou and S-b Peng. 2012. Effect of nitrogen regimes on grain yield, nitrogen utilization, radiation use efficiency, and sheath blight disease intensity in super hybrid rice. *Journal of Integrative Agriculture.* 11: 134-143.
- Mahbub, A M, M Hossain and A Janaich. 2001. Hybrid rice adoption in Bangladesh. A socioeconomic assessment of farmer's experience: Research Monograph. Series no. 18. Social Science Division, International Rice Research Institute (IRRI), Los Banos, Laguna, Philippines. p. 38.
- McKinney, H H. 1923. A new system of grading plant diseases. *Journal of Agriculture Research* 26: 195-218.
- Mew, T W, H Leung, S Savary, C M V Cruz and J E Leach. 2004. Looking ahead in rice disease research and management. *Critical Reviews in Plant Science.* 23: 103-127.
- Miyakie, I. 1910. Studies über die Pilze der Reispflanzen in Japan. *J. Coll. Agric.* 2: 237-276.
- Moni, Z R. 2012. Biological characterization and management of *Rhizoctonia solani* causal organism of rice sheath blight disease. PhD thesis, Department of Botany, University of Rajshahi, Rajshahi, Bangladesh.
- Parveen, S. 2016. Management of sheath blight disease of rice. PhD thesis, Department of Genetics and Plant Breeding, BAU, Mymensingh, Bangladesh. p. 58.
- Prasad, B and G C Eizenga. 2008. Rice sheath blight disease resistance identified in *Oryza* spp. accessions. *Plant Dis.* 92: 1503-1509.
- Rao, K M. 1995. Sheath blight disease of rice. Daya Publishing House, Delhi-110035.
- Roychowdhury, R, J Karmakar, M K Adak, N Dey and A Mitra. 2013. Physio-biochemical and microsatellite based profiling of lowland rice (*Oryza sativa* L.) landraces for osmotic stress tolerance. *American Journal of Plant Sciences.* 4(12C): 52-63.
- Salam, M U, S M A Hossain, J K Biswas and A J Mridha. 2014. Managing the unmanageable: rice variety technology for future challenging food security in Bangladesh. Extended abstract in the Agronomic visions in challenging future, the proceedings of the 13th conference of the Bangladesh Society of Agronomy, 20 September 2014, Bangladesh Rice Research Institute (BRRI), Gazipur, Bangladesh.
- Shahjahan, A K M, N R Sharma, H U Ahmed and S A Miah. 1986. Yield loss in modern rice varieties of Bangladesh due to sheath blight. *Bangladesh J. of Agricultural Research.* 11(2): 82-90.
- Sharma, N R and P S Teng. 1990. Effects of inoculum source on sheath blight development. *Int. Rice Res. Newsl.* 15: 18-19.
- Wu, W, J Huang, K Cui, L Nie, Q Wang, F Yang, F Shah, F Yao and S Peng. 2012. Sheath blight reduces stem breaking resistance and increases lodging susceptibility of rice plants. *Field Crops Research.* 128: 101-10