In vitro Analysis Provides New Insights into the Pharmacological Actions of Methanol Extract of Seeds of Tamarindus indica L. and its Kupchan Fractions

Sharmeen Asad¹, Farzana Kabir¹, Safaet Alam¹, Fahmida Tasnim Richi², Irin Pervin Anny¹, Mst. Luthfun Nesa¹ and Mohammad A. Rashid³

¹Department of Pharmacy, State University of Bangladesh, Dhaka-1205, Bangladesh
²Department of Pharmacy, Faculty of Pharmacy, University of Dhaka, Dhaka-1000, Bangladesh
³Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Dhaka, Dhaka-1000, Bangladesh

(Received: July 19, 2021; Accepted: October 10, 2021; Published (web): January 29, 2022)

Abstract

Tamarind or Tamarindus indica L. is a multipurpose plant distributed throughout the tropics including Bangladesh. The present study was conducted to establish the preliminary antioxidant, antimicrobial, anti-inflammatory and thrombolytic activities of methanol extract of T. indica seeds along with its Kupchan fractions. To evaluate the antioxidant properties, the total phenolic content of T. indica was determined and expressed in gallic acid equivalent (GAE). Alongside, DPPH free radical scavenging assay was performed to ensure the antioxidant properties of the seeds where the methanolic crude extract revealed the maximum activity having IC₅₀ value of 9.43 μg/ml. In the antimicrobial assay by disk diffusion method, only non-polar fractions of the extract showed mild antimicrobial activity against the test organisms tested while the polar crude methanol extract exhibited the maximum inhibition (58.16%, p < 0.001) of hypotonic solution-induced erythrocyte rupture in anti-inflammatory investigation among all the partitionates. During evaluation of thrombolytic activity in terms of percent of clot lysis, the methanol soluble fraction exhibited the highest percent of thrombolysis (23.5%) as compared to the reference standard, streptokinase (64.25%). The findings of the current study rationalize some of the traditional uses of T. indica and preliminarily ascertain its bioactive potential, which may act as a base for phytochemical and mechanism-based pharmacological studies of the plant in future.

Key words: Tamarindus indica, seed, antioxidant, antimicrobial, thrombolytic, membrane stabilizing activity.

Introduction

From the ancient era, medicinal plants assume a vital role in the security of human wellbeing. It has been accounted that 66% of the world's plant species contain medicinal properties (Krishnaiah et al., 2011). These medicinal plants contain various components of therapeutic properties so they can be utilized as drugs or formulations to treat different human illnesses (WHO, 1993). The possibility of the research work with these medicinal plants will not be fading out easily due to their natural abundance (Alam et al., 2020; Emon et al., 2020).

Tamarindus indica L. (family: Fabaceae, subfamily: Caesalpinioideae, Common name: Tamarind) is a tropical fruit with a plethora of nutritional and medicinal values which is locally known as “Tetul” (Kumar and Bhattacharya, 2008). Though it is indigenous to Africa, it has been naturalized and produced throughout Asia- Bangladesh, Indonesia, and Thailand (El-Siddig, 2006). Traditionally, it is used to heal wounds and treat fever, stomachache, colds and coughs, diarrhea along with as an antidote for snake bites (Buchholz and Melzig, 2016; Kaur

Corresponding author: Mohammad A. Rashid, E-mail: r.pchem@yahoo.com

DOI: https://doi.org/10.3329/bpj.v25i1.57835
and Bhullar, 2016). Previous pharmacological investigations of various parts i.e. fruit, leaves, bark, pulp and flowers have reported its role as antidiabetic, anti-inflammatory, antimicrobial and its potential role in the treatment or prevention of obesity and other chronic diseases (Bhadoria et al., 2011; Reis et al., 2016). Abundant bioactive compounds such as tannins, polyphenols, and phenolic antioxidants (β-sitosterol, eicosanoic acid, n-hexacosane, pinitol, and proanthocyanidins) also have been isolated from this plant (Landgraf Guiguer and Barbalho, 2016). Enriched in nutrients and chemical diversity, otherwise inedible and wasted seeds have been considered for their newfound usage as an inexpensive alternative protein source after detailed processing to remove tannins (Alam et al., 2021; El-Siddig, 2006; Siddhuraja et al., 1995). In continuation, to reduce the waste of tamarind seeds, we herein conducted in vitro assays on methanol extract of T. indica seeds to find out the logical evidence for its medicinal properties and explore its preliminary total phenolic content, DPPH free radical scavenging, antimicrobial, anti-inflammatory (membrane stabilizing) and thrombolytic potential.

Materials and Methods

Collection, drying, and extraction: T. indica seeds were collected in August of 2015 from Manikgonj district of Bangladesh, and identified by an expert taxonomist in Bangladesh National Herbarium, Dhaka. A voucher specimen was also recorded (DACB 36425) for this collection. The seeds were thoroughly cleaned before being sun dried for two weeks followed by grinding into a coarse powder. Approximately 300 g of dried ground seeds were steeped in 1.5 L of methanol for four weeks followed by filtering through cotton plug and Whatman No. 1 filter paper. The filtrate was concentrated at 45°C maintaining decreased pressure using a rotary evaporator. The petroleum ether (PESF), carbon tetrachloride (CTCSF), and aqueous soluble fractions (AQSF) were obtained from the concentrated methanol extract (ME, approximately 10 g) using the modified Kupchan partitioning protocoll (VanWagenen et al., 1993). Thereafter, the pharmacological functions of the crude extract (ME) and its Kupchan fractions were studied individually.

Drugs and chemicals: All of the reagents employed in this research work, were of analytical grade and purchased from reputable vendors. Merck (Darmstadt, Germany) supplied the methanol, Tween-80, and ascorbic acid. Sigma Chemicals Co. (St. Louis, MO, USA) provided 1,1-diphenyl-2-picryl-hydrazyl radical (DPPH) and Folin-Ciocalteu reagent (FCR) whereas, normal saline solution was purchased from Beximco Infusion Ltd., Bangladesh and acetylsalicylic acid and streptokinase were procured from Incepta Pharmaceuticals Ltd., Bangladesh during this research work.

Antioxidant activity analysis: The antioxidant properties of the extract were assessed using two distinct methods.

a) Total phenolics analysis: Considering Folin-Ciocalteu reagent as an oxidizing agent and gallic acid as a standard, the total phenolic content of T. indica extracts was determined (Harbertson and Spayd, 2006). In this experiment, 2.5 ml of Folin-Ciocalteu reagent diluted 10 times with water, and 2 ml of sodium carbonate (7.5 percent w/v) solution were merged with 0.5 ml of each extract (2 mg/ml) in water. After that, the mixtures were allowed to sit at room temperature for 20 minutes. Finally, with a UV-visible spectrophotometer, the absorbance was recorded at 760 nm. Total phenolics were determined using a calibration curve created by measuring the standard gallic acid values (0 – 100 g/ml) and reported as mg of GAE (gallic acid equivalent)/gm of dried extract.

b) Determination of free radical scavenging activity: The antioxidant activity of the methanol crude extract of T. indica and its Kupchan fractions on the stable radical 1,1-diphenyl-2-picrylhydrazyl (DPPH) was assessed using ascorbic acid as a positive control (Brand-Williams et al., 1995). With a successive serial dilution method, 2.0 mg of each extract was dissolved in methanol, producing solutions with different concentrations of 500, 250, 125, 62.50, 31.25, 15.62, 7.81, 3.91, 1.95, and 0.98
g/ml. The extracts were then combined with 3.0 ml of a DPPH-methanol solution (20 g/ml) and allowed to stand for 20 minutes. Finally, the absorbance was measured at 517 nm and from these values, the corresponding percentage of inhibition was determined utilizing the following equation:

\[
\text{% Inhibition} = \left\{ \left( A_0 - A_1 \right) / A_0 \right\} \times 100
\]

Where, \( A_0 \) = the absorbance of control; \( A_1 \) = the absorbance of sample.

Then % inhibitions were plotted against respective concentrations used and from the graph, the IC\(_{50}\) was calculated.

**Antimicrobial activity analysis:** The disc diffusion technique was employed to investigate the antibacterial activity of methanol crude extract of *T. indica* and its Kupchan fractions (Bauer, 1966) against 5 gram-positive bacteria, 8 gram-negative bacteria and 3 fungi utilizing nutrient agar medium. The extracts were carefully put on the agar plates in the previously designated zones. Positive controls of standard ciprofloxacin and fluconazole discs (30 g/discs) were utilized to confirm activity against test organisms. The zones of inhibition developed by the extracts were analyzed and correlated with the standard.

**Membrane stabilizing activity analysis:** Hypotonic-induced hemolysis was studied by employing previously mentioned method with slight modification to evaluate the membrane stabilizing activity of the methanol extract and its fractions (Shinde et al., 1999). For this 5 ml of whole blood from healthy human volunteers was taken into a tube containing 2.2 mg di-potassium salt of EDTA per 1.0 ml of blood for the preparation of human red blood cell suspension. Then, the collected blood was centrifuged at 3000 rpm for 10 minutes. After that, the supernatant was separated and the blood cells were washed with saline in 10 mM sodium phosphate buffer (pH 7.4) for three times using the same volume as the supernatant followed by reconstitution in the same volume of this isotonic buffer solution. After that, 5 ml of hypotonic solution (50 mM NaCl) in 10 mM sodium phosphate buffered saline (pH 7.4) containing either reference drug, acetyl salicylic acid (0.1 mg/ml) or the extract (1 mg/ml), was combined with 0.5 ml of the resultant solution. The control sample was composed of 0.5 ml of RBCs which was combined with hypotonic buffered saline. Then, the mixture was incubated at room temperature for 10 minutes before being centrifuged at 3000 rpm for 10 minutes and the optical density (OD) of the supernatant was measured at 540 nm. The percentage inhibition of hemolysis was computed by using the equation as followed:

\[
\text{% inhibition of hemolysis} = \left\{ \left( \text{OD}_{\text{control}} - \text{OD}_{\text{test sample}} \right) / \text{OD}_{\text{control}} \right\} \times 100
\]

**Thrombolytic activity analysis:** For the thrombolytic test, 6 ml of venous blood was taken from healthy donors and placed in 10 separate pre-weighed sterile vials (1 ml/tube) (Prasad et al., 2006). Then the vials were kept in an incubator at 37°C for 45 minutes. After the clot had been formed, the serum was fully withdrawn without agitating it and each vial was weighed for the determination of the clot weight (clot weight = weight of clot containing tube – weight of tube alone). Each of the extracts (1 mg/100μl water) was then poured into each vial holding the known weighed clot in the following step. In this study, 100 μl of streptokinase and 100μl of distilled water were utilized as the positive and negative control, respectively. After that, all of the vials were incubated at 37°C for 90 minutes to check for clot lysis. The released fluid was collected after incubation and the vials were further weighed to monitor the difference in weight after the clot was disrupted. As demonstrated below, the alteration in weight acquired before and after clot lysis was represented as a percentage of clot lysis.

\[
\text{% of thrombolysis} = \left( \text{weight of clot after treatment} / \text{weight of clot before treatment} \right) \times 100
\]

**Statistical analysis:** For each experiment, three replicates of each sample were utilized, and statistical analysis was executed by using the Student’s t-test. The mean and standard deviation have been used to present all data.
Results and Discussion

Antioxidant activity analysis: The antioxidant properties of the test sample have been summarized in the Table 1.

a) Total phenolic content: In this study, the highest content of phenolic (27.87 mg of GAE/gm) was evident by the methanol extract of *T. indica* seeds (Table 1). The phenolic content of other fractions was observed in a range of 7-17 mg of GAE/gm signifying mild to moderate range of antioxidant potentials.

b) DPPH free radical scavenging assay: In case of DPPH free radical scavenging activity analysis, the methanol extract exhibited the highest scavenging of IC$_{50}$ = 9.43 μg/ml which was followed by the carbon tetrachloride fraction with IC$_{50}$ = 22.25 μg/ml (Table 1). Here, this study was compared with the standard ascorbic acid with the IC$_{50}$ = 3.05 μg/ml. As, phenolic acids and flavonoids are reported to exhibit prominent antioxidant potential (Emon *et al*., 2021; Rudra *et al*., 2020; Shahidi *et al*., 1992), the free radical neutralizing property of the methanol extract might be due to the said phytoconstituents which was further justified by the highest presence of phenolic content in the same fractionate.

Table 1. Total phenolic content, free radical scavenging, membrane stabilizing (% inhibition of hypotonic solution induced hemolysis) and thrombolytic activities of *T. indica* whole seeds.

<table>
<thead>
<tr>
<th>Sample/standard</th>
<th>Total phenolic content (mg of GAE/gm of extract)</th>
<th>DPPH Free radical scavenging activity (IC$_{50}$μg/ml)</th>
<th>Membrane stabilizing activity (% inhibition of hypotonic solution induced)</th>
<th>Thrombolytic (% clot lysis)</th>
</tr>
</thead>
<tbody>
<tr>
<td>METI</td>
<td>27.87±0.12</td>
<td>9.43±0.22**</td>
<td>0.89±0.01</td>
<td>58.16±0.33**</td>
</tr>
<tr>
<td>PESF</td>
<td>15.03±0.10</td>
<td>39.15±0.34**</td>
<td>1.65±0.04</td>
<td>22.38±1.09**</td>
</tr>
<tr>
<td>CTCSF</td>
<td>17.31±0.22</td>
<td>22.25±0.28**</td>
<td>1.92±0.01</td>
<td>10.01±0.30**</td>
</tr>
<tr>
<td>AQSF</td>
<td>7.38±0.19</td>
<td>77.31±0.29**</td>
<td>1.03±0.03</td>
<td>51.63±0.77**</td>
</tr>
<tr>
<td>AA</td>
<td>3.05±0.15**</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>ASA</td>
<td>--</td>
<td></td>
<td>0.60±0.02</td>
<td>71.89±0.29**</td>
</tr>
<tr>
<td>SK</td>
<td>--</td>
<td></td>
<td>2.13±0.08</td>
<td>--</td>
</tr>
<tr>
<td>Blank</td>
<td>--</td>
<td></td>
<td>2.13±0.08</td>
<td>--</td>
</tr>
</tbody>
</table>

Here, METI: Methanolic crude extract of *T. indica* seeds, PESF: Pet ether soluble fraction, CTCSF: Carbon tetrachloride fraction, AQSF: Aqueous soluble fraction, AA = Ascorbic acid, ASA: Acetyl salicylic acid, SK: Streptokinase. All values are presented as mean± standard deviation. *p*< 0.005, **p*< 0.001 is considered as statistically significant as compared to negative control.

Antimicrobial activity analysis: In the probing of antimicrobial activity by disc diffusion assay, only pet ether soluble fraction and carbon tetrachloride fraction of seeds of *T. indica* exhibited a smaller extent of antimicrobial activity (zone of inhibition = 7-11 mm) against the test organisms at concentrations of 400 μg/ml (Table 2). It can be summarized that the non-polar fractions (pet ether and carbon tetrachloride) were observed with more activity against the test strains as compared to the polar fractions (methanol and water).

Membrane stabilizing activity analysis: In table 1, the percentage of erythrocyte membrane rupture inhibition of different fractionates of the seed of *T. indica* has been presented. The highest RBC membrane protecting capacity was exhibited for the methanol extract of seeds of *T. indica* (58.16%) when compared with the standard acetyl salicylic acid (71.89%). The second highest membrane stabilizing activity was noticed by the aqueous soluble fraction (51.63%) whereas mild erythrocyte membrane protecting capacity was indicated by the pet ether soluble fraction (22.38%) and carbon tetrachloride...
fraction (10.01%). Stabilization of erythrocyte membrane has long been extrapolated to stipulate the effects of different compounds on the stabilization of lysosomal membrane that is known to be related to the release of inflammatory cytokines (Omale and Okafor, 2008; Shinde et al., 1999). Based on this results, it can be proposed that the more polar fractions were noticed with more ability to resist the cell lysis as compared to the non-polar pet ether and carbon tetrachloride fraction which is suggestive of the presence of constituents with therapeutic potentials for inflammatory conditions.

Table 2. Antimicrobial activity of different fractions of seeds of T. indica.

<table>
<thead>
<tr>
<th>Test Microorganisms</th>
<th>Diameter of Zone of Inhibition (mm)</th>
<th>METI</th>
<th>PESF</th>
<th>CTCSF</th>
<th>DCMSF</th>
<th>AQSF</th>
<th>Ciprofloxacin</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Gram Positive Bacteria</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bacillus sereus</td>
<td></td>
<td>-</td>
<td>8</td>
<td>8</td>
<td>-</td>
<td>-</td>
<td>40</td>
</tr>
<tr>
<td>B. megaterium</td>
<td></td>
<td>-</td>
<td>9</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>42</td>
</tr>
<tr>
<td>B. subtilis</td>
<td></td>
<td>-</td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>41</td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td></td>
<td>-</td>
<td>10</td>
<td>8</td>
<td>-</td>
<td>-</td>
<td>45</td>
</tr>
<tr>
<td>Sarcina lutea</td>
<td></td>
<td>-</td>
<td>10</td>
<td>9</td>
<td>-</td>
<td>-</td>
<td>46</td>
</tr>
<tr>
<td><strong>Gram Negative Bacteria</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Escherichia coli</td>
<td></td>
<td>-</td>
<td>7</td>
<td>8</td>
<td>-</td>
<td>-</td>
<td>45</td>
</tr>
<tr>
<td>Pseudomonas aureus</td>
<td></td>
<td>-</td>
<td>10</td>
<td>8</td>
<td>-</td>
<td>-</td>
<td>41</td>
</tr>
<tr>
<td>Salmonella paratyphi</td>
<td></td>
<td>-</td>
<td>-</td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>35</td>
</tr>
<tr>
<td>S. typhi</td>
<td></td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>45</td>
</tr>
<tr>
<td>Shigella boydii</td>
<td></td>
<td>-</td>
<td>8</td>
<td>8</td>
<td>-</td>
<td>-</td>
<td>42</td>
</tr>
<tr>
<td>Sh. dysenteriae</td>
<td></td>
<td>-</td>
<td>7</td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>41</td>
</tr>
<tr>
<td>Vibrio mimicus</td>
<td></td>
<td>-</td>
<td>10</td>
<td>9</td>
<td>-</td>
<td>-</td>
<td>38</td>
</tr>
<tr>
<td>V. parahemolyticus</td>
<td></td>
<td>-</td>
<td>11</td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>44</td>
</tr>
<tr>
<td><strong>Fungi</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aspergillus niger</td>
<td></td>
<td>-</td>
<td>7</td>
<td>10</td>
<td>-</td>
<td>-</td>
<td>47</td>
</tr>
<tr>
<td>Candida albicans</td>
<td></td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>42</td>
</tr>
<tr>
<td>Sacharomyces cerevisae</td>
<td></td>
<td>-</td>
<td>7</td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>46</td>
</tr>
</tbody>
</table>

**Thrombolytic activity analysis:** The percentages of clot lysis of different fractions of the seed of T. indica have been illustrated in Table 1. The highest thrombolytic activity (23.5%) was evident by the methanol extract of seeds of T. indica whereas the standard streptokinase showed 64.25% thrombolysis. On the other hand, moderate clot lysis ability was also noticed by carbon tetrachloride fraction (22.51%) and aqueous soluble fraction (17.26%). However, the very weak thrombolytic activity was observed by the pet ether soluble fraction (2.49%). The current result is suggestive of the probable availability of thrombolytic agents in the polar fractions.

**Conclusion**

The methanol extract of seeds of T. indica and its Kupchan fractions were investigated for antioxidant, antimicrobial, anti-inflammatory and thrombolytic activities. After performing the evaluations, it may be assumed that these extractives can be considered as a potential source of antioxidant, anti-inflammatory, and thrombolytic therapeutic agents. Additional comprehensive investigations are recommended to isolate the bioactive components from this plant part.
responsible for the exerted pharmacological actions and to know their in-depth underlying mechanisms.

Declaration

All authors read the manuscript and approve it for the publication and no part of this manuscript has been published before in any journal.

Conflict of interest

The authors state that they have no conflict of interest.

Acknowledgements

The authors are thankful to the authorities of Department of Pharmacy, State University of Bangladesh for providing necessary facilities to carry out this research work.

References


