Evaluation of Bioactivities of *Gouania tiliaefolia* Lam., an Indigenous Traditional Medicinal Plant of Bangladesh

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Abstract

The crude methanol extract of *Gouania tiliaefolia* Lam. was partitioned by the modified Kupchan method and the fractions were evaluated for total phenolic content, antioxidant, cytotoxic, thrombolytic, hypotonic and heat-induced membrane stabilizing activities. The petroleum ether soluble fraction (PESF) and methanol extract (ME) showed the highest phenolic content of 78.30 ±1.60 mg and 70.37 ±0.84 mg, respectively, which were expressed in gallic acid equivalent (GAE). Similar trends were observed in case of anti-oxidant and cytotoxic activities, where the PESF possessed the highest free radical scavenging activity and brine shrimp lethality (IC50 = 2.88 ±0.02 µg/ml, LC50 = 2.59 ±0.14 µg/ml), followed by ME (IC50 = 4.79 ±0.17 µg/ml, LC50 = 3.38 ±0.08 µg/ml) and CSF (IC50 = 37.51 ±0.96 µg/ml, LC50 = 73.55 ± 0.26 µg/ml). In case of assays for thrombolytic and membrane stabilizing activities, all extractives showed insignificant results compared to the respective standards. The crude methanol extract of *G. tiliaefolia* was used to examine the in-vivo analgesic (central and peripheral), antidiarrheal and antidiabetic activities in Swiss albino mice. In case of castor oil induced diarrhea, the ME gave better reduction of diarrhea by 71.43% (at 400 mg/kg-body weight) compared to loperamide (64.29%). Anti-diabetic activity was evaluated by oral glucose tolerance test and the ME showed 71.42% and 75.39% reduction of blood glucose at doses 200 and 400 mg/kg-body weight, respectively when compared with the standard glibenclamide that reduced blood glucose by 66.17%. The central- and peripheral-analgesic activity was evaluated by the tail-flick test and acetic acid induced writhing test, respectively. In both the cases, ME demonstrated dose-dependent analgesic activity compared to the standards.

Key words: *Gouania tiliaefolia* Lam., total phenolic content, antioxidant, cytotoxicity, thrombolytic, antidiarrheal, antidiabetic

Introduction

Despite the revolution of modern medicine in the twentieth century, it has been estimated that about one-third of the world’s population fails to receive affordable medicines. This has led to a surge in the use of traditional (herbal), complementary and alternative medicines in both developed and developing countries (Twarog et al., 2004). In addition, international public health organizations like World Health Organization (WHO) are encouraging the use of traditional medicines due to its wide biological activities, higher safety-profile and lower costs compared to the synthetic drugs (Sharma et al., 2008; Moniruzzaman et al., 2018). Complementary and alternative medicines, as well as herbal medicines contribute a significant role in providing health care along with mainstream allopathic treatment in Bangladesh. Reportedly more than 80% of the Bangladeshi people use non-allopathic (i.e., Ayurveda, Siddha, Unani and Homoeopathy) medicines for their primary healthcare and herbs constitute a major ingredient of

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these alternative systems of medicine (Kadir et al., 2013). Though people carry an erroneous belief that medications of natural (e.g., herbal) origin are inherently safe and possess lesser side effects (Firenzuoli et al., 2004), natural constituents could be carcinogenic, initiate allergic symptoms, and interfere with drug bioavailability and metabolism etc. inside the body (Firenzuoli et al., 2007), which requires proper investigation, pure compound isolation, characterization and biological evaluation.

_Gouania tiliacefolia_ Lam. (family: Rhamnaceae), locally known as ‘Moshkantur’ is a native plant of Bangladesh that has been used extensively for various ailments. For example, juice from the crushed leaves has been used orally and topically to alleviate fever and headache (Khatun et al., 2013). Tribal population in the hill tracts of Bandarban and Rangamati, commonly use the paste of whole plant to cover any wounded area and sores (Motaleb et al., 2015; Sarwar et al., 2018). In the Philippines, it is widely used among locals for curing stomach ulcer (Carag et al., 2017).

Despite its wide use, till date no scientific data has yet been published to evaluate its pharmacological properties or to evaluate its toxicity _in-vivo_. On the other hand, if the medicinal use of this plant is backed by biological study then further research could be carried out in order to isolate and identify the phytochemical(s) responsible for its bioactivity and later be used for at least new drug discovery and development. Therefore, the objective of this current study was to evaluate the traditional medicinal properties of the _G. tiliacefolia_ by conducting both _in-vitro_ and _in-vivo_ biological assays of the methanol extract of the whole plant and its various organic soluble fractions.

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**Materials and Methods**

_Collection of sample and extraction:_ Fresh whole plant of _G. tiliacefolia_ was collected from Rangamati district, south-east of Bangladesh during October 2018. The plant was then identified by the taxonomist of Bangladesh National Herbarium, Dhaka, Bangladesh.

After the collection and identification, the plant samples were cleaned, sun dried and ground into coarse powder. Then 400 gm of this powder was soaked in 1600 ml of methanol in an amber bottle. The container was sealed with a cotton plug and kept for 7 days with occasional shaking to facilitate the extraction of phyto-constituents. The whole mixture was filtered by cotton plug followed by Whatman number 1 filter paper. The filtrate was further evaporated to dryness using a rotary evaporator at reduced temperature and pressure. The concentrated gummy mass was transferred to a clean beaker.

Different partitions of the crude extract were obtained using the modified Kupchan method (Van Wagenen et al., 1993). For this 5 g of the crude extract was re-dissolved in 10% aqueous methanol and extracted consecutively with petroleum ether, chloroform, dichloromethane and water to provide petroleum ether soluble fraction (PESF, gm), dichloromethane soluble fraction (DCMSF, gm) and chloroform soluble fraction (CSF, gm) and aqueous soluble fraction (AQRSF, gm).

**Drugs and chemicals:** Acetic acid, methanol, Tween-80, vincristine sulfate, diclofenac sodium and normal saline were purchased from retail shops. DMSO was collected from Merck specialties, Mumbai. All other reagents used for these tests were of analytical grade.

**Experimental animal:** Swiss Albino mice of both sexes (4-5 weeks old) weighing between 25-35 gm were obtained from the Department of Pharmacy, Jahangir Nagar University. The mice were kept in the animal house of the State University of Bangladesh and fed with standard rodent feed. As these animals are sensitive towards environmental change, they were kept for 4 days in the laboratory environment prior to experimental use. The Federation of European Laboratory Animal Science Associations (FELASA) guidelines and recommendations were followed to reduce the pain and stress of the experimental mice. In case of _in-vivo_ analysis, 12 Swiss Albino mice were randomly divided into four groups (i.e., positive control, negative control and two test groups receiving methanolic extract at doses of 200 and 400 mg/kg of body weight).
**Determination of total phenolic content:** The total phenolic content of the whole plant of *G. tiliaefolia* was determined by following reported protocol (Skerget *et al.*, 2005) using Folin-Ciocalteu reagent (i.e., a mixture of phosphomolybdate and phosphotungstate) as oxidizing agent and gallic acid as standard.

**Antioxidant activity (DPPH assay):** The free radical scavenging activities (antioxidant capacity) of the plant extracts on the stable radical 1,1-diphenyl-2-picrylhydrazyl (DPPH) were estimated by the method of Brand-Williams *et al.*, 1995).

**Cytotoxic activity (Brine shrimp lethality bioassay):** Brineshrimp lethality bioassay was performed using the procedures reported by Meyer and co-workers (Meyer *et al.*, 1982).

**Anti-inflammatory activity (membrane stabilizing bioassay):** According to the method developed by Shinde and co-workers (Shinde *et al.*, 1999) and further modified by Sikder and co-workers (Sikder *et al.*, 2011), the membrane stabilizing activity of the extractives was determined by evaluating their ability to prevent hypotonic solution and heat-induced hemolysis of human erythrocyte.

**Thrombolytic activity:** In order to find the thrombolytic ability (i.e., determining percentage clot lysis) of extractives, the method used by Prasad and co-workers (Prasad *et al.*, 2006) was employed in this study. Here, 10 ml blood sample were collected and distributed equally into pre-weighed sterile vial (1 ml/vial) and incubated at 37°C for 45 minutes. After clot formation, the serum was completely removed, and 100 μl aqueous solutions of different partitions along with the crude extracts were added separately. As a negative control, 100μl of distilled water was added to the control vial, while streptokinase was used as the standard. All the vials were incubated at 37°C for 90 minutes and observed for clotlysis using the formula:

\[
\text{Percentage clot lysis} = \frac{\text{Weight of the lysed clot}}{\text{Weight of clot before lysis}} \times 100
\]

**Antimicrobial activity:** The extract and its distinctive portions were assessed by single disk diffusion technique (Murray *et al.*, 1995; Zavala *et al.*, 1997) to decide their antimicrobial potencies at 400 μg/disc. In this examination, ciprofloxacin and fluconazole (30 μg/disc) circles were utilized as the reference for the antibacterial and antifungal screening.

**Central and peripheral analgesic activity:** The tail flicking test (Pizziketti *et al.*, 1985) and acetic acid induced writhing test (Kaushik *et al.*, 2012) were followed to determine the central and peripheral analgesic activity, respectively. In both cases, the mice were divided into four groups (3 mice in each group), positive control, negative control, methanol extract receiving groups (200 and 400 mg/kg-body weight). In the tail flicking method, two doses were fed orally to the mice and the tips of their tails were immersed in hot water at 15 second interval and the time taken to respond was recorded at 30, 60 and 90 minutes interval. Morphine (2 mg/kg-body weight) was injected subcutaneously to the positive control group and 1% Tween-80 in saline mixture (0.1 ml/10 mg) was fed to the negative control group. The pain inhibition process (PIP) was calculated using the following formula:

\[
\text{Pain inhibition percentage (PIP)} = \left[\frac{T_1 - T_0}{T_0}\right] \times 100
\]

where, \(T_1 = \text{post - drug latency}\); \(T_0 = \text{pre-drug latency}\)

For the writhing method, the positive control group received diclofenac sodium (50 mg/kg-body weight) orally while the negative control group received distilled water (10 ml/kg). Two doses (200 and 400 mg/kg-body weight) of crude extract were fed orally and waited for 40 minutes before injecting 1% (v/v) of acetic acid at a dose of 10 ml/kg-body weight, intra-peritonially. The acetic acid induced writhing movement in the mouse and the time taken for licking response was recorded. Percent inhibition
of writhing movement was proportional to the analgesic activity and was calculated using the formula as shown below:

\[
\text{Percentage inhibition} = \frac{\text{Response time in negative control} - \text{Response time in test group}}{\text{Response time in negative control}}
\]

**Hypoglycemic activity assay:** Tail tipping method was used to determine the hypoglycemic effect of the test samples in mice (Dürschlag et al., 1996). In the study, blood was withdrawn from the tip of the tail and the sugar level was measured by using glucometer. A total of twelve mice divided into four groups were used in this study. The mice of negative and positive control groups received 1% Tween-80 in saline (0.1 ml/10 mg) and glibenclamide (5 mg/kg-body weight), respectively. The methanol extract (at 200 and 400 mg/kg-body weight of dose) partition was given orally to mice of treatment groups. After 60 minutes, 10% glucose solution (2 g/kg-body weight) was given orally to all the mice. The blood sugar level was again recorded after 1st, 2nd and 3rd hour of administration of glucose solution.

**Anti-diarrheal activity assay:** Mice of both sexes were fasted for 18 hrs. The selected mice for castor oil-induced diarrheal test were divided into six groups (where, n = 3). Group-I (control group) was given normal saline (2 ml/kg-body weight) orally and group-II (standard group) received loperamide (2 mg/kg-body weight), groups-III and IV received methanol extract of *G. tiliaefolia* at doses of 200 and 400 mg/kg-body weight. After 60 minutes, each mouse of four groups received castor oil (1 ml) orally. Later, they were placed in cages lined with absorbent papers and observed for 4 hrs for the presence of characteristic diarrheal droppings.

**Results and Discussion**

The crude extract of *G. tiliaefolia* and its Kupchan partitionates were analyzed for *in-vitro* bioactivities, while the methanol extract at 200 and 400 mg/kg-body weight were used for the *in-vivo* bioassays.

The highest phenolic content was obtained in PESF (78.30±1.60 mg of GAE/g), followed by the methanol extract (70.37±0.84 mg) and chloroform soluble fractions (48.52±1.16 mg). This high phenolic content could be an indication for the presence of different phyto-constituents such as flavonoids and tannins, which are well known for their antioxidant activity (Akinmoladun et al., 2007; Pietta et al., 2000; Hagerman et al., 1999).

Similar trends were also observed in case of evaluating the free radical scavenging potential, cytotoxic efficacy and thrombolytic activity. In all the cases, the PESF gave better activity compared to the ME and CSF. The IC\textsubscript{50} values for free radical scavenging activity test were found as 24.79±0.17, 28.88±0.02 and 37.51±0.96 µg/ml, in case of PESF, ME and CSF, respectively (Table 1). The significant high levels of free radical scavenging and cytotoxic activities of *G. tiliaefolia* test samples could be attributed to the presence of these phenolics and flavonoid contents.

In this case, the IC\textsubscript{50} of PESF and ME were comparable to the standard BHT (25.18±1.89 µg/ml). In case of cytotoxic activity, the PESF showed the highest cytotoxic activity with LC\textsubscript{50} value of 2.59 ± 0.14 µg/ml followed by methanol extract with LC\textsubscript{50} value of 3.38±0.08 µg/ml as compared to the standard vincristine sulfate (LC\textsubscript{50} = 0.45±0.00 µg/ml) (Table 1). The highest capacity to lyse blood clot was also shown by the PESF (46.45±0.23%) and ME (30.30±0.08%) as compared to the standard streptokinase (67.36±0.19%) (Table 1).

During the evaluation of membrane stabilizing activity using hypotonic solution induced hemolysis, all the fractions of *G. tiliaefolia* showed satisfactory inhibition; but, the percent inhibitory effects for all
fractions were lower than the standard acetyl salicylic acid (85.37±3.00%). Highest percentage of inhibition of hemolysis was exhibited by the PESF (69.18±0.54%) followed by the ME (58.05±1.32%) (Table 1).

During the inhibition of heat induced hemolysis, PESF, DCMSF and ME revealed the inhibitory effect close to the standard acetyl salicylic acid (42.11±0.10%) (Table 1). The observations in thrombolytic assay may also have significance since it may lead to the discovery of new cardiovascular drugs (Hussain et al., 2014). The highest thrombolytic and membrane stabilizing activity (hypotonic, heat induced) were shown by the PE and ME of *G.tiliaefolia*. Even though, in both the cases the results were lower but promising when compared to the standards.

However, further study in order to isolate the compound(s) responsible could result in the discovery of better anti-coagulating agents.

The methanol extract of *G. tiliaefolia* showed a dose dependent inhibition of castor oil-induced diarrhea in Swiss albino mice. The methanol extract when given at 200 and 400 mg/kg-body weight reduced diarrhea by 39.29% (p < 0.01) and 71.43% (p < 0.001), respectively (Table 2).

### Table 1. Total phenolic content, antioxidant and cytotoxic activity of test samples of *G.tiliaefolia*.

<table>
<thead>
<tr>
<th>Samples/ Standards</th>
<th>Total phenolic content (mg of GAE/g of dried extract)</th>
<th>Anti-oxidant activity IC50 (µg/ml)</th>
<th>Cytotoxic activity LC50 (µg/ml)</th>
<th>Thrombolytic activity % of clot lysis</th>
<th>Membrane stabilizing activity % Inhibition of hypotonic solution induced hemolysis</th>
<th>% Inhibition of heat induced hemolysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>ME</td>
<td>70.37±0.84</td>
<td>28.88±0.02</td>
<td>3.38±0.08</td>
<td>30.30±0.08</td>
<td>58.05±1.32</td>
<td>32.85±0.56</td>
</tr>
<tr>
<td>PESF</td>
<td>78.30±1.60</td>
<td>24.79±0.17</td>
<td>2.59±0.14</td>
<td>46.45±0.23</td>
<td>69.18±0.54</td>
<td>39.70±0.22</td>
</tr>
<tr>
<td>DCMSF</td>
<td>10.12±0.55</td>
<td>164.40±27.17</td>
<td>255.54±0.45</td>
<td>10.93±0.01</td>
<td>54.21±0.78</td>
<td>36.60±0.92</td>
</tr>
<tr>
<td>CSF</td>
<td>48.52±1.16</td>
<td>37.51±0.96</td>
<td>73.55±0.26</td>
<td>24.72±0.01</td>
<td>54.72±0.45</td>
<td>21.85±0.07</td>
</tr>
<tr>
<td>AQSIF</td>
<td>38.58±0.77</td>
<td>39.01±0.84</td>
<td>139.74±0.77</td>
<td>11.63±0.02</td>
<td>54.20±2.55</td>
<td>26.79±1.37</td>
</tr>
<tr>
<td>VS (std.)</td>
<td>-</td>
<td>0.45±0.00</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>BHT (std.)</td>
<td>-</td>
<td>25.18±1.89</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>ASA (std.)</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>85.37±3.00</td>
<td>42.11±0.10</td>
</tr>
<tr>
<td>SK (std.)</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>67.36±0.19</td>
<td>-</td>
</tr>
</tbody>
</table>

ME = Methanol extract; PESF= Pet ether soluble fraction; DCMSF = Dichloromethane soluble fraction; CSF = Chloroform soluble fraction; AQSIF = Aqueous soluble fraction; VS = Vincristine sulfate; BHT = Butylated hydroxy toluene; GAE = Gallic acid equivalent, ASA= Acetyl salicylic acid; SK = Streptokinase

### Table 2. Antidiarrheal activity of crude methanol extract of *G. tiliaefolia*.

<table>
<thead>
<tr>
<th>Test groups</th>
<th>No. of mice</th>
<th>Number of diarrheal feces Mean±SEM</th>
<th>% Reduction of diarrheal feces</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (1% Tween-80)</td>
<td>3</td>
<td>9.33±0.24</td>
<td>-</td>
</tr>
<tr>
<td>Loperamide (standard)</td>
<td>3</td>
<td>3.33±0.24</td>
<td>64.29</td>
</tr>
<tr>
<td>(2 mg/kg-body weight)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>ME (200 mg/kg-body weight)</td>
<td>3</td>
<td>5.67±0.47</td>
<td>39.29</td>
</tr>
<tr>
<td>ME (400 mg/kg-body weight)</td>
<td>3</td>
<td>2.67±0.24</td>
<td>71.43</td>
</tr>
</tbody>
</table>

Here, n = 3; Values are expressed as mean ± SEM; p < 0.05 significant when compared to negative control.
The methanol extract of *G. tiliaefolia* was found to lower the blood sugar level significantly (p < 0.001) during evaluating its hypoglycemic activity by tail tipping method. Its efficacy was evident even after 3rd hour of administration of the glucose solution.

The blood sugar level was reduced by 66.17% (p < 0.001) and 71.42% (p < 0.001) at 200 and 400 mg/kg-body weight of doses, respectively (Table 3).

In case of central analgesic activity assay by tail-flick method, 105.95% (p < 0.01) and 165.07% (p < 0.001) elongation of the reaction time was observed after 30 and 60 minute of administration of ME (200 mg/kg-body weight), respectively. The methanol extract at 400 mg/kg-body weight showed 129.73% (p < 0.01) and 190.83% (p < 0.001) of pain inhibition at the same time period (Table 4).

On the other hand, in case of acetic acid induced writhing assay, the crude extract of *G. tiliaefolia* inhibited the number of writhing by 54.17% (p < 0.01) and 64.58% (p < 0.001) at two different doses, which were lower but significant when compared to the 77.08% inhibition by the standard diclofenac sodium (Table 5).

Table 3. Antidiabetic activity of methanol extract of *G. tiliaefolia*.

<table>
<thead>
<tr>
<th>Test groups</th>
<th>Average blood glucose level (mmol/l)</th>
<th>% Reduction of glucose level</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Before treatment After treatment (minute)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>30 60 120</td>
<td>180 120 min 180 min</td>
</tr>
<tr>
<td>Control (1% Tween-80)</td>
<td>4.80±0.78</td>
<td>14.1±4.23 17.17±0.62</td>
</tr>
<tr>
<td>Glibenclamide (5 mg/kg bw)</td>
<td>5.67±0.81</td>
<td>16.63±2.41 5.23±1.03</td>
</tr>
<tr>
<td>ME (200 mg/kg bw)</td>
<td>4.17±0.23</td>
<td>13.8±3.1 6.17±0.64</td>
</tr>
<tr>
<td>ME (400 mg/kg bw)</td>
<td>3.47±0.07</td>
<td>15.3±1.31 9.23±4.69</td>
</tr>
</tbody>
</table>

Here, n = 3; Values are expressed as mean ± SEM; p < 0.05 significant when compared to negative control.

Table 4. Central analgesic activity of crude methanol extract of *G. tiliaefolia*.

<table>
<thead>
<tr>
<th>Test groups</th>
<th>Mean of tail immersion time (minute) ± SEM</th>
<th>% Elongation of response</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 30 60 90</td>
<td>0 min 30 min 60 min</td>
</tr>
<tr>
<td>Control (1% Tween-80)</td>
<td>1.69±0.08</td>
<td>1.85±0.19 2.29±0.14</td>
</tr>
<tr>
<td>Morphine (2 mg/kg)</td>
<td>1.75±0.10</td>
<td>5.68±0.07 9.05±0.15</td>
</tr>
<tr>
<td>ME (200 mg/kg)</td>
<td>1.99±0.06</td>
<td>3.81±0.21 6.07±0.23</td>
</tr>
<tr>
<td>ME (400 mg/kg)</td>
<td>1.98±0.04</td>
<td>4.25±0.36 6.66±0.30</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SEM (n = 3); p < 0.05 significant when compared to negative control.
Table 5. Peripheral analgesic activity of crude methanol extract of *G. tiliaefolia*.

<table>
<thead>
<tr>
<th>Test groups</th>
<th>Writhing count</th>
<th>Number of writhing</th>
<th>% Writhing</th>
<th>% Inhibition of writhing</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>M1</td>
<td>M2</td>
<td>M3</td>
<td>Mean ± SEM</td>
</tr>
<tr>
<td>Control</td>
<td>17</td>
<td>17</td>
<td>14</td>
<td>16.00±1.00</td>
</tr>
<tr>
<td>(1% Tween-80)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Diclofenac sodium (50 mg/kg bw)</td>
<td>4</td>
<td>5</td>
<td>2</td>
<td>3.67±0.88</td>
</tr>
<tr>
<td>ME (200 mg/kg bw)</td>
<td>9</td>
<td>7</td>
<td>6</td>
<td>7.33±0.88</td>
</tr>
<tr>
<td>ME (400 mg/kg bw)</td>
<td>6</td>
<td>5</td>
<td>6</td>
<td>5.67±0.33</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SEM (n = 3); p < 0.05 significant when compared to negative control.

**Conclusion**

This preliminary *in vitro* and *in vivo* bioassays with the crude extract and different organic partitions of *G. tiliaefolia* provides evidence that the plant possess potential biologically active chemical constituents responsible for its various medicinal properties. However, further study needs to be carried out in order to isolate and purify the compounds responsible for such pharmacological activities. Determination of the structure and analyzing the structure activity relationship could lead to discovery of potential drug candidates.

**References**


