

Antitumor, Analgesic and Anti-inflammatory Activities of *Glochidion multiloculare* (Rottler ex Willd) Voigt

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Abstract

The antitumor, analgesic and anti-inflammatory activities of the methanol extract (MEGM) of *Glochidion multiloculare* and its petroleum ether (PEFGM), carbon tetrachloride (CTFGM) and chloroform (CHFGM) soluble fractions were investigated. The antitumor activity of the extract/fractions was determined against Ehrlich's ascites carcinoma (EAC) in mice at 20 mg/kg body weight intraperitoneally. Increase of survival time by 24 ± 0.12 , 26 ± 0.40 , 21 ± 0.12 and 27 ± 0.42 days was observed by the MEGM, PEFGM, CTFGM and CHFGM treated tumor bearing mice compared to the control group (20 ± 0.12 days). Hematological studies revealed that hemoglobin (Hb) content was decreased in EAC treated mice whereas restoration to normal levels was observed in extract/fractions treated animals. Significant decrease in RBC and increase in WBC counts were observed in extract/fractions treated animals when compared to EAC affected animals. Both extract and fractions at the dose of 100 mg/kg b.w. significantly increased the pain threshold in tail immersion method and reduced the writhing caused by acetic acid induced method. The MEGM, PEFGM, CTFGM and CHFGM showed anti-inflammatory activities at 100 mg/kg b.w. This is the first report of antitumor, analgesic and anti-inflammatory potential of *G. multiloculare*.

Key words: *Glochidion multiloculare*, antitumor, analgesic, anti-inflammatory

Introduction

According to the estimates of the WHO, more than 80% of people in developing countries depend on traditional medicine for their primary health needs. A recent survey shows that more than 60% of cancer patients use vitamins or herbs as therapy (Madhuri and Pandey, 2008; Sivalokanathan *et al.*, 2005). There is evidence in the literature that anti-inflammatory compounds could be also anticancerous. These compounds could modify the redox environment of cancer cells and thus their behavior (Schafer and Buettner, 2001).

Glochidion was regarded as a genus of the family Euphorbiaceae, which consists of monoecious, rarely dioecious trees or shrubs. But molecular phylogenetic studies have shown that *Phyllanthus* is paraphyletic over *Glochidion*. A recent revision of the family Phyllanthaceae has subsumed *Glochidion* into *Phyllanthus* (Hoffmann *et al.*, 2006). *Glochidion multiloculare* (Roxb. ex Willd.) Muell.-Arg., Phyllanthaceae (synonym:

Phyllanthus multilocularis), locally known as Aniatori, Keotomi, Keoura, Paniatori, Pannyaturi is an evergreen shrub or small tree. The plant is found in Bhutan, India, Myanmar, Nepal and Bangladesh. Traditionally many *Phyllanthus* species are used in haemorrhoids, diarrhoea, dysentery, anaemia, jaundice, dyspepsia, insomnia etc. and some of them can induce diuresis (Ghani, 1998). Previous phytochemical investigations revealed the isolation of glochidiol, glochilocudiol, glochidone and dimedone (Talapatra *et al.*, 1973). Many secondary metabolites were isolated from *Glochidion* species, including tannins (Chen *et al.*, 1995), glycosides (Otsuka *et al.*, 2003), lignans (Otsuka *et al.*, 2000), terpenoids (Hui and Li, 1976).

To the best of our knowledge from the literatures, the biological activities of *G. multiloculare* were not explored extensively. The objectives of the present study were to examine the antitumor, analgesic and anti-inflammatory potential of *G. multiloculare*.

Materials and Methods

Plant materials: The stem bark of *G. multiloculare* was collected from Modhupur, Tangail in the month of April, 2009 and identified by Mr. Sarder Nasir Uddin, Scientific Officer, Bangladesh National Herbarium, Dhaka, where a voucher specimen (DACB- 34200) representing this collection has been deposited.

Chemicals: Sodium chloride, propylene glycol, trypan blue, methylene blue, methyl violet, sodium sulphate, and bleomycin were purchased from Merck Limited, Mumbai, India. Diclofenac-Na and indomethacin were collected from Square Pharmaceuticals Ltd., Bangladesh. All other chemicals and reagents were used of highest analytical grade.

Preparation of extract: The air dried and powdered plant material (1000 g) was extracted with methanol (3000 mL) for 7 days) at room temperature with occasional shaking and stirring. The extractive was filtered through fresh cotton plug and followed by Whatman no. 1 filter paper. The filtrate was then concentrated by a Buchii rotavapor at low temperature and pressure and to afford methanol (MEGM) extract (41.7398 g). The cold methanol extract (10 g) was subjected to solvent-solvent partitioning using modified Kupchan protocol (VanWagenen *et al.*, 1993). The extract was partitioned successively into petroleum ether (PEFGM), carbon tetrachloride (CTFGM) and chloroform (CHFGM) soluble fractions.

Animal: Swiss albino mice (25-30 g) of either sex were used for assessing the antitumor activity. The animals were maintained under standard laboratory conditions and had free access to food and water *ad libitum*. The animals were allowed to acclimatize to the environment for 7 days prior to experimental session. They were divided into seven groups, each consisting of twelve animals which were fasted overnight prior to the experiments. Experiments with the animals were performed in accordance with guidelines of the Institutional Animal Ethics Committee, Department of Applied Chemistry & Chemical Engineering, University of Rajshahi, Rajshahi, Bangladesh.

Acute toxicity: The acute oral toxicity of the plant extract in Swiss albino mice was studied as per established protocol (Lorke, 1983).

Biological assays

In vivo antitumor activity

Transplantation of tumor: Ehrlich's ascites carcinoma (EAC) cells were obtained from Indian Institute of Chemical Biology (IICB), Kolkata, India. The EAC cells were maintained *in vivo* in Swiss albino mice by intraperitoneal transplantation of 2×10^6 cells per mouse after every 10 days. Ascitic fluid was drawn from EAC tumor bearing mice at the log phase (days 7-8 of cell implantation) of the tumor cells. Each animal received 0.1 ml of tumor cell suspension containing 2×10^6 tumor cells intraperitoneally.

Treatment schedule: 84 Swiss albino mice were divided into seven groups (n = 12) and given food and water *ad libitum*. All the animals in each group except Group-I received EAC cells (2×10^6 cells/mouse i.p.). This was taken as day '0'. Group-I served as normal saline control (5 ml/kg i.p.) and Group-II served as EAC treated control. After 24 h of EAC transplantation, Group-III, Group-IV, Group-V and Group-VI received MEGM, PEFGM, CTFGM and CHFGM of *G. multiloculare* stem bark at 20 mg/kg i.p. for 9 consecutive days. Group-VII received reference drug bleomycin (0.3 mg/kg i.p) for 9 consecutive days (Rana and Khanam, 2002). After 24 h of last dose and 18 h of fasting, 6 animals from each group were sacrificed by cervical dislocation to measure antitumor and hematological parameters and the rest were kept with food and water *ad libitum* to check percentage increase in life span of the tumor bearing host. The antitumor activity of the extract/ fractions of *G. multiloculare* was measured in EAC bearing animals with respect to the following parameters.

Determination of tumor volume and weight: The mice were dissected and the ascitic fluid was collected from the peritoneal cavity. The volume was measured by taking it in a graduated centrifuge tube and weighed immediately.

Tumor cell count: The ascitic fluid was taken in a WBC pipette and diluted 100 times with phosphate buffer saline (PBS). Then a drop of the diluted cell suspension was placed on the Neubauer's counting chamber and the number of cells in the 64 small squares was counted.

Viable/nonviable tumor cell count: The viability and non-viability of the cells were checked by trypan blue assay with the help of microscope. The cells were stained with trypan blue (0.4% in normal saline) dye. The cells

that did not take up the dye were viable and those that took the dye were nonviable. The viable and nonviable cells were counted using the following equation:

$$\text{Cell count} = (\text{Number of cells} \times \text{dilution factor}) / (\text{Area} \times \text{thickness of liquid film})$$

Determination of median survival time and percentage increase in life span: The mortality was monitored by recording percentage increase in life span (%ILS) and median survival time (MST) (Sur and Ganguly, 1994).

Hematological parameters: Collected blood samples were used for the estimation of hemoglobin (Hb) content, red blood cell (RBC) and white blood cell count (Armour et al., 1965).

Analgesic activity

Tail flick test: The animals were divided into six groups with six mice in each group. Group I animals received vehicle (1% Tween 80 in water, 10 mL/kg body weight, negative control) animals of Group VI received Diclofenac-Na at 10 mg/kg body weight (positive control), while animals of Group II, Group III, Group IV and Group V were treated with 100 mg/kg body weight (p.o.) of the MEGM, PEFGM, CTFGM and CHF GM of *G. multiloculare*. Then 1-2 cm of the tail of mice was immersed in warm water kept at constant temperature of 60 °C and the reaction time was the time taken by the mice to deflect their tails was noted. The first reading was discarded and the reaction time was recorded as a mean of the next three readings. A latency period of 20 sec was defined as complete analgesia and the measurement was then stopped to avoid injury to mice. The latent period of the tail-flick response was determined before and 0, 30, 60 and 90 min after the oral administration of drugs (Toma et al., 2003).

Acetic acid-induced writhing test: The analgesic activity of the samples was also studied using acetic acid-induced writhing model in mice. Test samples and vehicle were administered orally 30 min before intraperitoneal administration of 0.7% acetic acid but Diclofenac-Na was administered intraperitoneally 15 min before injection of acetic acid. After an interval of 5 min, the mice were observed for specific contraction of body referred to as 'writhing' for the next 10 min (Ghule et al., 2011).

Anti-inflammatory activity

Carrageenan induced paw edema test in mice: Mice were divided into six groups containing six animals in each. The test groups received 100 mg/kg, p.o. of each extract/fractions. The positive control group received indomethacin (10 mg/kg, p.o.) while the negative control group received 3 mL/kg of 1% Tween 80 in water. After 1 h, 0.1 mL of 1% w/v carrageenan suspension in normal saline was injected into the sub plantar tissue of the right hind paw (Winter et al., 1962). The paw volume was measured at 1, 2, 3 and 4 h after carrageenan injection using a micrometer screw gauge. The percentage inhibition of the inflammation was calculated from the formula: % inhibition = $(1 - D_t/D_o) \times 100$, whereas D_o was the average inflammation (hind paw edema) of the control group of mice at a given time and D_t was the average inflammation of the drug treated (i.e. extract/fractions or reference indomethacin) mice at the same time (Gupta et al., 2005).

Statistical analysis: Antitumor data are expressed as mean \pm S.E.M. (n = 12). Statistical significance (p) calculated by Student's t test. $P < 0.001$ and < 0.05 were considered to be statistically significant. Analgesic and anti-inflammatory data are expressed as mean \pm S.E.M. (n = 6). Statistical significance (p) calculated by ANOVA followed by Dunnett's test ($p < 0.01$ and $p < 0.001$) were considered to be statistically significant.

Results and Discussion

The acute toxicity study was conducted to establish the therapeutic index, i.e., the ratio between the pharmacologically effective dose and the lethal dose on the same strain and species. The extract/fractions of *G. multiloculare* were safe up to a dose of 500 mg/kg (p.o.) body weight which agrees with the previous study (Rahman et al., 2011). Behavior of the animals was closely observed for the first 3h then at an interval of every 4 h during the next 48 h. The extract did not cause mortality in mice during 48 h of observation but little behavioral changes, locomotors ataxia, diarrhea and weight loss were observed. Food and water intake had no significant difference among the groups studied.

Antitumor activity of extract/fractions against EAC tumor bearing mice was assessed by the parameters such as tumor volume, tumor weight, cell count (viable and

non-viable), mean survival time and % increase of life span. The results are shown in table 1. The tumor volume, tumor weight and viable cell count were found to be significantly ($p < 0.001$) increased and non-viable cell count was significantly ($p < 0.001$) low in EAC control animals when compared with normal saline control animals. Administration of MEGM, PEFGM, CTFGM and CHF GM at a dose of 20 mg/kg significantly ($p < 0.05$) decreased the tumor volume, tumor weight and viable cell count. Furthermore, the median survival time was increased to 25 ± 0.32 (% ILS = 24.6), 26 ± 0.10 (% ILS =

30.3), 23 ± 0.19 (% ILS = 20.5) and 27 ± 0.42 (% ILS = 29.01) after administration of MEGM, PEFGM, CTFGM and CHF GM at a dose of 20 mg/kg b.w., respectively.

Hematological parameters (Table 2) of tumor bearing mice after 14 days were found to be significantly altered compared to the normal saline group. The total WBC count was found to be increased with a reduction of Hb content and RBC. The total number of RBC showed a modest change. At the same time interval on extract/fractions at a dose of 20 mg/kg restored all the altered hematological parameters to almost near normal.

Table 1. Effect of *G. multiloculare* on tumor volume and weight, mean survival time (MST), percentage increase life span (% ILS), viable and non-viable tumor cell count in EAC bearing mice.

Parameter	EAC control	MEGM	PEFGM	CTFGM	CHF GM	Bleomycin
Tumor volume (ml)	3.1 ± 0.21	0.89 ± 0.29	$0.76 \pm 0.34^*$	$1.12 \pm 0.21^*$	$0.72 \pm 0.34^*$	$0.51 \pm 0.21^*$
Tumor weight (g)	3.90 ± 0.24	1.82 ± 0.24	$1.25 \pm 0.21^*$	$2.01 \pm 0.11^*$	$1.22 \pm 0.21^*$	$0.61 \pm 0.11^*$
MST (days)	20 ± 0.12	24 ± 0.12	26 ± 0.40	21 ± 0.12	27 ± 0.42	42.6 ± 0.12
% ILS	00.0	22.2	27.2	19.81	29.01%	98.81
Viable cell ($\times 10^7$ cell/ml)	8.1 ± 0.22	$0.96 \pm 0.22^*$	$0.66 \pm 0.05^*$	$1.52 \pm 0.05^*$	$0.50 \pm 0.05^*$	$0.5 \pm 0.05^*$
Non-viable cell ($\times 10^7$ cell/ml)	0.5 ± 0.24	$2.21 \pm 0.24^*$	$2.13 \pm 0.54^*$	$2.07 \pm 0.05^*$	$2.01 \pm 0.54^*$	$3.3 \pm 0.05^*$
Total cell ($\times 10^7$ cell/ml)	8.6 ± 0.15	3.18 ± 0.15	2.79 ± 0.21	3.59 ± 0.05	2.51 ± 0.21	$3.8 \pm 0.05^*$
Viable %	94.18	30.51	23.66	42.34	23.31%	13.15
Non-viable %	5.82	69.49	76.34	57.66	76.84	86.85

Each point represent the mean \pm SEM. (n = 6), * $p < 0.05$ statistically significant when compared with EAC control group.

Table 2. Effect of *G. multiloculare* on hematological parameter in EAC bearing mice.

Treatment	RBC (cell $\times 10^3/\text{mm}^3$)	WBC (cell $\times 10^3/\text{mm}^3$)	Hemoglobin (g %)
Normal saline control	5.69 ± 0.12	3.92 ± 0.32	13.90 ± 3.1
EAC control	$3.91 \pm 0.80^*$	$5.94 \pm 0.52^*$	$4.95 \pm 1.80^*$
Bleomycin (0.3 mg/kg b.w.)	$5.18 \pm 0.12^{**}$	$3.35 \pm 0.83^{**}$	$12.89 \pm 2.93^{**}$
MEGM (20 mg/kg b.w.)	$4.37 \pm 0.16^{**}$	$4.59 \pm 0.12^{**}$	$7.93 \pm 1.62^{**}$
PEFGM (20 mg/kg b.w.)	$4.83 \pm 0.61^{**}$	$4.49 \pm 0.39^{**}$	$8.73 \pm 1.01^{**}$
CTFGM (20 mg/kg b.w.)	$4.01 \pm 0.29^{**}$	$4.97 \pm 0.32^{**}$	$7.23 \pm 1.79^{**}$
CHF GM (20 mg/kg b.w.)	$4.81 \pm 0.61^{**}$	$4.44 \pm 0.39^{**}$	$8.50 \pm 1.00^{**}$

Each point represent the mean \pm SEM. (n = 6), * $p < 0.001$ statistically significant and ** $p < 0.005$ statistically significant; when compared with EAC control group.

In the antitumor activity test, a regular and quick increase in ascetic tumor volume was observed. A quick increase of fluid with tumor growth would be a way to meet the nutritional requirement of tumor cells (Prasad and Giri, 1994). Treatment with extract/fractions reduced the intraperitoneal tumor burden, thereby fall the tumor weight, cell growth and increased the life span of the tumor bearing mice. Therefore, extract/fractions increased the life span of EAC bearing mice may be by declining the

nutritional fluid volume and delaying the cell division (Sur et al., 1997). Anemia encountered in ascites carcinoma usually due to iron deficiency and finally leads to reduced RBC number (Gupta et al., 2007). Administration of extract/fractions reversed the hemoglobin content, thus supporting its haematopoietic activity without inducing myelotoxicity. Earlier study of *Glochidione* species indicated the presence of tannins, glycosides, lignan and terpenoid. The mechanism of anticancer activity of

phenolics (tannins) could be by troubling the cellular division during mitosis at the telophase stage (Gawron and Kruk, 1992). Therefore, it can be taken into evidence that the plant extract/fractions exhibited antitumor activity through these antitumor phytochemicals.

The tail withdrawal reflex time following administration of the MEGM, PEFGM, CTFGM and CHFGM were found to statistically significant (* $p < 0.01$ and ** $p < 0.001$) and was comparable to the reference drug Diclofenac-Na (Table 3).

Table 4 shows the effects of the extract/fractions on acetic acid-induced writhing in mice. The oral administration of extract/fractions significantly (* $p < 0.01$ and ** $p < 0.001$) inhibited (% inhibition of writhing of MEGM, PEFGM, CTFGM and CHFGM were 70, 66.36, 83.25 and 84.09, respectively) writhing response induced by acetic acid.

Table 3. Effects of *G. multiloculare* on tail withdrawal reflex of mice induced by tail flick method.

Treatment group	Dose (mg/kg b.w.)	Tail flick time			
		0 Min	30 Min	60 Min	90 Min
Vehicle control (1% Tween-80)	-	1.30 ± 0.11	1.35 ± 0.12	1.43 ± 0.05	1.40 ± 0.11
Standard (Diclofenac-Na)	10	1.40 ± 0.02	2.26 ± 0.25**	3.13 ± 0.10**	3.06 ± 0.20**
MEGM	100	1.20 ± 0.10	1.38 ± 0.10**	2.31 ± 0.15**	1.48 ± 0.10*
PEFGM	100	1.6 ± 0.20	1.80 ± 0.26**	2.30 ± 0.11**	1.53 ± 0.08*
CTFGM	100	1.6 ± 0.12	2.05 ± 0.04**	2.34 ± 0.03**	1.17 ± 0.07*
CHFGM	100	1.45 ± 0.01	1.53 ± 0.21**	1.91 ± 0.16**	1.23 ± 0.10*

Values are mean ± SEM, (n = 6); * $p < 0.01$ and ** $p < 0.001$, Dunnett's test as compared to control.

Table 4. Effects of the *G. multiloculare* on acetic acid-induced writhing in mice.

Treatment group	Dose (mg/kg b.w.)	Number of writhing	% Inhibition of writhing
Vehicle control (1% Tween-80)		110.0 ± 10.23	-
Standard (Diclofenac-Na)	10	15.0 ± 3.25**	86.36
MEGM	100	33.0 ± 8.57**	70
PEFGM	100	37.0 ± 5.57**	66.36
CTFGM	100	18.42 ± 5.0**	83.25
CHFGM	100	17.5 ± 4.50**	84.09

Values are mean ± SEM, (n = 6); * $p < 0.01$ and ** $p < 0.001$, Dunnett's test as compared to control.

The dorsal horn of the spinal cord is gifted with several neurotransmitters and receptors which are the major targets for pain and inflammation (McCurdy and Scully, 2005). The tail immersion test was considered to be a selective method to study compounds acting through opioid receptor. All the extract/fractions increased pain threshold, which indicates that it may act *via* centrally mediated analgesic mechanism (Elisabetsky *et al.*, 1995). On the other hand, acetic acid-induced writhing response is a susceptible procedure to assess peripherally acting analgesics and represents pain sensation by triggering localized inflammatory response. The response is taken to be mediated by peritoneal mast cells (Ribeiro *et al.*, 2000),

acid sensing ion channels (Voilley, 2004) and the prostaglandin pathways (Hossain *et al.*, 2006). The organic acid has also been postulated to act indirectly by inducing the release of endogenous mediators, which stimulates the nociceptive neurons that are responsive to NSAIDs and narcotics (Adzu *et al.*, 2003).

However, Kerstein *et al.* (2009) recommend that, the inhibitory effect to pain response may be due to local increase of the intracellular Ca^{2+} ion which can affect TRPA1 (Transient Receptor Potential Ankyrin 1) activity resulting in analgesic action. So it is expected that the extract/fractions may hold substances that affect the metabolism of Ca^{2+} ions. Tannins are also found to have a

contribution in antinociceptive activity (Ramprasath *et al.*, 2006).

In the carrageenan induced paw edema test in mice, the extract/fraction showed inhibition on paw edema compared to the control group (Table 5).

Carrageenan test has been used as an experimental animal model for acute inflammation and is believed to be biphasic. The early phase (1–2 h) is mainly mediated by

histamine, serotonin and increased synthesis of prostaglandins. The late phase is sustained by prostaglandin release and mediated by bradykinin, leukotrienes, polymorphonuclear cells and prostaglandins (Kaushik *et al.*, 2012). As the extract/fractions significantly inhibited paw edema in the second phase and this finding suggests a possible inhibition of cyclooxygenase synthesis.

Table 5. Effects of *G. multiloculare* on carrageenan induced paw edema in mice.

Treatment group	Dose (mg/kg b.w.)	Paw edema volume (cm)			
		1H	2H	3H	4H
Vehicle control (1% Tween-80)		1.23 ± 0.11	1.4 ± 0.12**	1.53 ± 0.05**	1.46 ± 0.11**
Standard (Indomethacin)	10	1.23 ± 0.02	1.16 ± 0.25**	1.13 ± 0.10**	1.06 ± 0.20**
MEGM	100	1.3 ± 0.07	1.2 ± 0.05**	1.0 ± 0.07**	0.9 ± 0.045**
PEFGM	100	1.25 ± 0.04	1.15 ± 0.07**	1.0 ± 0.04**	0.95 ± 0.5**
CTFGM	100	1.25 ± 0.32	1.2 ± 0.07**	1.0 ± 0.25**	0.9 ± 0.07**
CHFGM	100	1.3 ± 0.45	1.15 ± 0.05**	0.95 ± 0.02**	0.85 ± 0.05**

Values are mean ± SEM, (n = 6); *p<0.01 and **p<0.001, Dunnett's test as compared to control.

Conclusions

On the basis of our results, it may be concluded that the plant extract and fractions possess antitumor, analgesic and anti-inflammatory potential. However, further studies are necessary to examine the underlying mechanisms of these effects and to isolate the active compound(s) responsible for these pharmacological activities.

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References

- Adzu, B., Amos, S., Kapu, S.D. and Gamaniel, K.S. 2003. Anti-inflammatory and anti-nociceptive effects of *Sphaeranthus senegalensis*. *J. Ethnopharmacol.* **84**, 169-173.
- Armour, D.F.E., Blood, F.R. and Belden, D.A. 1965. *The Manual for laboratory works in mammalian physiology* 3rd Edition. The University of Chicago Press, Chicago, pp. 4-6.
- Chen, L.G., Yang, L.L., Yen, K.Y., Hatano, T., Yoshida, T. and Okuda, T. 1995. Tannins of euphorbiaceous plants. XIII: New hydrolyzable tannins having phloroglucinol residue from *Glochidion rubrum* BLUME. *Chem. Pharm. Bull.* **43**, 2088-2090.

- Elisabetsky, E., Arnador, T.A., Albuquerque, R.R., Nunes, D.S. and Carvalho, A.DoC.T. 1995. Analgesic activity of *Psychotria colorata* (Willd. ex R. and S.) Muell. Arg. alkaloids," *J. Ethnopharmacol.* **48**, 77-83.
- Gawron, A. and Kruk, G. 1992. Cytotoxic effect of xanthotoxol (8-hydroxypsoralen) on TCTC cells *in vitro*. *Pol. J. Pharmacol. Pharm.* **44**, 51-57.
- Ghani, A. 1998. *Medicinal Plants of Bangladesh: Chemical Constituents and Uses*. 1st Edition, Asiatic Society of Bangladesh.
- Ghule, R.S., Venkatnarayanan, R., Thakare, S.P., Jain, H. and Ghule, P.R. 2011. Analgesic activity of *Cuscuta campestris* Yuncker a parasitic plant grown on *Nerium indicum* Mill. *J. Adv. Phar. Educ. Res.* **1**, 45-51.
- Gupta, M., Mazumder, U.K., Halder, P.K., Kandar, C.C., Monikandan, L. and Senthil, G.P. 2007. Anticancer activity of *Indigofera aspalathoides* and *Wedelia calendulaceae* in swiss albino mice. *Iranian J. Pharm. Res.* **6**, 141-145.
- Gupta, M., Mazumder, U.K., Sambath, K.R., Gomath, P., Rajeshwar, Y. and Kakoti, B.B. 2005. Anti-inflammatory, analgesic and antipyretic effects of methanol extract from *Bauhinia racemosa* stem bark in animal models. *J. Ethnopharmacol.* **98**, 267-273.
- Hoffmann, P., Kathriarachchi, H. and Wurdack, K.J. 2006. A revised phylogenetic classification of Phyllanthaceae (Malpighiales; Euphorbiaceae sensu lato). *Kew. Bull.* **61**, 37-53.

- Hossain, M.M, Ali, M.S., Saha, A. and Alimuzzaman, M. 2006. Antinociceptive activity of whole plant extracts of *Paederia foetida*. *Dhaka Univ. J. Pharm. Sci.* **5**, 67- 69.
- Hui, W-H. and Li, M-M. 1976. Lupene triterpenoids from *Glochidion eriocarpum*. *Phytochemistry* **15**, 561-562.
- Kaushik, D., Kumar, A., Kaushik, P. and Rana, A.C. 2012. *Adv. Pharmacol. Sci.* doi:10.1155/2012/245431, Article ID 245431 (2012).
- Kerstein, P.C., Camino, D. del, Moran, M.M. and Stucky, C.L. 2009. Pharmacological blockade of TRPA1 inhibits mechanical firing in nociceptors. *Mol. Pain* **5**, 19.
- Lorke, D. 1983. A new approach to practical acute toxicity testing. *Arch. Toxicol.* **54**, 275-187.
- Madhuri, S. and Pandey, G. 2008. Some dietary agricultural plants with anticancer properties. *Plant Arch.* **8**, 13-16.
- McCurdy, C.R. and Scully S.S. 2005. Analgesic substances derived from natural products (natureceuticals). *Life Sci.* **78**, 476-484.
- Otsuka, H., Hirata, E., Shinzato, T. and Takeda, Y. 2000. Isolation of lignan glucosides and neolignan sulfate from the leaves of *Glochidion zeylanicum* (Gaertn.) A. Juss. *Chem. Pharm. Bull.* **48**, 1084-1086.
- Otsuka, H., Kijima, H., Hirata, E., Shinzato, T., Takushi, A., Bando, M. and Takeda, Y. 2003. Glochidionionosides A-D: megastigmane glucosides from leaves of *Glochidion zeylanicum* (Gaertn.) A. Juss. *Chem. Pharm. Bull.* **51**, 286-290.
- Prasad, S.B. and Giri, A. 1994. Antitumor effect of cisplatin against murine ascites Dalton's lymphoma. *Indian J. Exp. Biol.* **32**, 155-162.
- Rahman, M.H., Alam, M.B., Chowdhury, N.S., Jha, M.K., Hasan, M., Khan, M.M., Rahman, M.S. and Haque, M.E. 2011. Antioxidant, analgesic and toxic potentiality of *Stephania japonica* (Thunb.) Miens. Leaf. *Int. J. Pharmacol.* **7**, 257-262.
- Ramprasath, V.R., Shanthi, P. and Sachdanandam, P. 2006. Immunomodulatory and anti-inflammatory effects of *Semecarpus anacardium* LINN. Nut milk extract in experimental inflammatory conditions. *Biol. Pharm. Bull.* **29**, 693-700.
- Rana, A.Y.K.M.M. and Khanam, J.A. 2002. *Aristolochia indica* whole plant extract as an antineoplastic Agent. *J. Med. Sci.* **2**, 202-205.
- Ribeiro, R.A., Vale, M.L., Thomazzi, S.M., Paschoalato, A.B., Poole, S., Ferreira, S.H. and Cunha, F.Q. 2000. Involvement of resident macrophages and mast cells in the writhing nociceptive response induced by zymosan and acetic acid in mice. *Eur. J. Pharmacol.* **387**, 111-118.
- Schafer, F.Q. and Buettner, G.R. 2001. Redox state of the cell as viewed through the glutathione disulfide/glutathione couple. *Free Rad. Biol. Med.* **30**, 1191-1212.
- Sivalokanathan, S., Ilayaraja, M. and Balasubramaniam, M.P. 2005. Efficacy of *Terminalia arjuna* (Roxb.) on N-nitrosodiethylamine induced hepatocellular carcinoma in rats. *Indian J. Exp. Biol.* **43**, 264-267.
- Sur, P. and Ganguly, D. K. 1994. Tea plant root extract (TRE) as an antineoplastic agent. *Planta Med.* **60**, 106-109.
- Sur, P., Bag, S.P. and Khanam, J.A. 1997. Choroacetohydroxamic acid as antitumor agent against *Ehrlich ascites* carcinoma in mice. *Neoplasma* **44**, 197-201.
- Talapatra, S.K., Bhattacharya, S., Maiti, B.C. and Talapatra, B. 1973. Structure of glochilodiol. New triterpenoid from *Glochidion multiloculare*. Natural occurrence of dimedone. *Chem. Ind. (London, United Kingdom)* **21**, 1033-1034.
- Toma, W., Graciosa, J.S., Hiruma-Lima, C.A., Andrade, F.D.P., Vilegas, W. and Souza-Brita, A.R.M. 2003. Evaluation of the analgesic and antiedematogenic activities of *Quassia amara* bark extract. *J. Ethnopharmacol.* **85**, 19-23.
- VanWagenen, B.C., Larsen, R., Cardellina, J.H.I.I., Randazzo, D., Lidert, Z.C., and Swithenbank, C. 1993. Ulosantoin, a potent insecticide from the sponge *Ulosareutzleri*. *J. Org. Chem.* **58**, 335-337.
- Voilley, N. 2004. Acid-sensing ion channels (ASICs): new targets for the analgesic effects of non-steroid anti-inflammatory drugs (NSAIDs). *Curr. Drug Targets Inflamm. Allergy* **3**, 71-79.
- Winter, C.A., Risley, E.A. and Nuss, G.W. 1962. Carrageenin-induced edema in hind paw of the rat as an assay for antiinflammatory drugs. *Pro. Soc. Exp. Biol. Med.* **111**, 544-547.