REGENERATION OF INORGANIC NUTRIENT IN EUTROPHIC ENVIRONMENT BY A BACTERIUM, XANTHOMONAS SP.

Md. Abdul Karim* and Rehena Nasrin Happy

Laboratory of Microbiology, Department of Botany, University of Dhaka, Dhaka-1000, Bangladesh

Most water bodies in Bangladesh have been polluted due to industrial growth, urbanization. Increased amount of pollutants in water make it to become discolored, smelly and change the quality which may be dangerous to life. In extreme cases, large number of fishes may die. Sewage is a major household waste and it is one of the main sources of water pollution. It contains many organic constituents and microorganisms. They are potential hazards for the aquatic organisms and human beings. If they are allowed to mix with river and seawater without proper treatment, this may pose a threat for the living being of water and other consumers. Release of untreated wastewater into the near-shore environment for decades has the potential to degrade the environment (Biamon and Hajen 1983), infect indigenous species (Matches *et al.* 1974). From a microbiological perspective this has perhaps the most extensive impact on community structure, eutrophication and oxygen demand.

The river Buriganga is economically the very important to Dhaka city but it is threatened by pollution and possession. The river is found to be Dhaka's main outlet of sewage waste. There have been reports that the river is being polluted by discharge of industrial effluents, municipal wastewaters, household wastes, clinical and pathological wastes, oils and human excreta (Kamal *et al.* 1999). The largest share of pollution load into the river Buriganga appears to be from about 200 tannery industries in the Hazaribagh and Rayerbazar area. Studies show that up to 15,000 m³ of liquid wastes, 19,000 kg of solid wastes and 17,600 kg of Biological Oxygen Demand (BOD) load goes into the Buriganga each day from the industries (Faisal *et al.* 2001).

Since the bacteria are well known agents of mineralization and transformation of organic and inorganic matters in aquatic ecosystem. Therefore, the objectives of the present study were: (i) isolation of indigenous bacterial isolates possessing high metabolic activities, and (ii) stimulation of inorganic nutrient regeneration by adding those bacteria.

^{*}Author for correspondence: <makarim10@yahoo.com>.

For the isolation of effective bacterium that might be active at high temperature, surface water samples (5 cm depth) were collected from the river Buriganga using 2 L plastic bottles during summer when temperature was about 32°C and were carried to the laboratory within an hour of sampling. The temperature, Dissolved Oxygen (DO), pH, Total Dissolved Solids (TDS) of the collected water samples were measured using standard kits.

From 110 fast growing bacterial isolates, 36 were selected considering growth pattern and colony morphology and finally BW-2 was selected for its biotechnological application.

Characteristics		Results	Characteristics	Results	Provisional Identification
Gram reaction		-	KOH	+	
Cell shape		Short rod,	Utilization of citrate	_	
-		occur singly			Xanthomonas
Spore		Non spore former	Utilization of propionate	-	sp.
Catalase		+	Indole formation	-	
Oxidase		+	Dihydroxy acetone	-	
			production		
VP		-	Fermen D-glucose (no	-	
			tation gas)		
MR		-	L-arabinose	-	
Levan		-	D-xylose	-	
Phenayal alanine		-	D-mannitol	-	
deaminase					
Protease		+	Degradation of tyrosine	+	
Deep glucose agar		Aerobic	Degradation of urea	+	
Motility		-	Nitrate reduction	-	
Hydrolysis	Starch	+	Egg yolk lecithinase	_	
	Casein	+	Egg yolk lipase	-	
	Gelatin	+			

Table 1 Morphological, physiological and biochemical characteristics of bacterial isolate BW-2

"+" indicate positive and "-" indicate negative

Bacterial inocula were prepared following method described by Karim *et al.* 2013. Five ml of bacterial cells re-suspension was used as inoculum and it was added into the treatment bottles. Bacterial activities were evaluated by the changes of inorganic nutrient regeneration into the water during incubation with or without adding bacteria. Just after adding bacteria into the experimental glass bottles, two glass bottles (in duplicate) were taken for chemical analysis without any disturbance of the system and considered as 0-day. The experiments were continued up to 15 days with 5 days interval. For chemical analysis, water samples were passed through the filter paper (Whatman No. 42, England) to eliminate suspended solid particles. The concentrations of inorganic form of nitrogen (DIN: ammonium, nitrate and nitrite) and phosphorus (DIP) in control and experimental glass bottles were measured following methods

described by Karim *et al.* 2013. Important physiological and biochemical characteristics were studied for the identification of the selected isolate (Sneath *et al.* 1986, SAB 1957 and Claus 1995).

The morphological, physiological and biochemical characteristics of the bacterial isolate *Xanthomonas* sp. are presented in Table 1. The isolate BW-2 exhibited remarkably high growth rates between 30°C and 40°C but didn't grow well at relatively low (10°C) and high temperature (55°C) and was considered to be mesophilic (Fig. 1). In regards to proteolytic activities, BW-2 possessed high activities by clear zone into the medium and increased with incubation period (Fig. 2).



Fig. 1 Growth of BW-2 bacterial isolate at different temperatures.



Fig. 2 Proteolytic activity of the bacterial isolate (BW-2) at different hours of incubation at 37°C.

The initial concentration of DIP in control was 0.06 mg/l, while in the system of adding *Xanthomonas* sp. it increased to 0.29 mg/l (Fig. 4). Net releases of DIP were 0.13 mg/l in control and 0.23 mg/l after adding *Xanthomonas* sp., while, net releases of DIN were 2.52 mg/l in control and 5.18 mg/l after adding *Xanthomonas* sp. (Fig. 3).



Fig. 3 Effect of Xanthomonas sp. on the change of dissolved inorganic phosphorus (A) and nitrogen (B) after different days of incubation period. Without Xanthomonas sp. (o) and with Xanthomonas sp. (o).

After adding *Xanthomonas* sp., net amounts of DIN and DIP regeneration in water were markedly enhanced and about two folds higher than the control counterpart. So, result of the experiment conducted with *Xanthomonas* sp. and incubated at *in situ* temperature showed that *Xanthomonas* sp. could be effective at high temperature. Further study at the next stage regarding the removal of enhanced inorganic nutrients from the water is recommended.

LITERATURE CITED

- BIAMON, E.J. and HAJEN, T.C. 1983. Survival and distribution of *Aeromanas hydrophila* in nearshore coastal waters of Puerto Rico receiving rum distillery effluent. *Water Res.* **17**: 319-326.
- CLAUS, G.W. 1995. Understanding microbes. (4th Ed.). W.H. Freeman and Company, New York. pp. 547.
- FAISAL, I.M., SHAMIN, R. and JUNAID, J. 2001. Industrial pollution, *In*: Nishat, A., Ullah, M. and Haque, A.K.E. (eds). Bangladesh Environment Outlook-2001. Centre for Sustainable Development (CFSD), Dhaka.
- KAMAL, M.M., MALMGREN-HANSEN, A. and BADRUZZAMAN, A.B.M. 1999. Assessment of pollution of the river Buriganga, Bangladesh, using a water quality model. *Water. Sci. Technol.* **40**(2): 129-36.
- KARIM, M.A., NAZNEEN, A. and HOQUE, S. 2013. Heterotrophic activities in eutrophic lake sediment by indigenous bacterium *Bacillus firmus. Bangladesh J. Zool.* **41**(1): 73-78.
- MATCHES, J.R., LISTON, J. and CURRAN, D. 1974. *Clostridium perfringens* in the environment. *Appl. Microbiol.* **28**: 655-660.
- SAB (Society of American Bacteriologists). 1957. Manual of microbiological methods. McGraw-Hill Book Co. Inc. NY. pp. 1-315.
- SNEATH, P.H.A., MAIR, N.S., SHARPE, M.E. and HOLT, J.G. (Eds.). 1986. Bergey's manual of systematic bacteriology. Vol. 2. The Williams and Wilkins Co., Baltimore, USA. pp. 965-1594.

(Manuscript received on 2 October, 2013; revised on 19 March, 2014)