



BCSIR

Available online at www.banglajol.info

Bangladesh J. Sci. Ind. Res. 52(4), 309-320, 2017

BANGLADESH JOURNAL
OF SCIENTIFIC AND
INDUSTRIAL RESEARCH

E-mail: bjisir07@gmail.com

A comparative biosorption study of sulfide (S^{2-}) by using *Lagenaria siceraria* fruit based raw biosorbent

P. Dar^{1*}, A. Dar^{2*}, D. Ahmed³ and A. M. Dar²

¹Quaid-1 Azam University Islamabad, 45320, Pakistan

²Center of Undergraduate Studies, University of the Punjab, Lahore 54590, Pakistan

²Institute of Chemical Engineering and Technology, University of The Punjab, Lahore 54590, Pakistan

³Department of Chemistry, Forman Christian College, Ferozpur Road, 54600 Lahore, Pakistan

Abstract

In batch mode, the adsorption characteristics of S^{2-} ions on *Lagenaria siceraria* mesocarp, pedicle and peel as raw materials have been studied in comparison to remove sulfide ions from tanneries waste water, under various parameters such as adsorbent dosage, contact time, agitation speed and pH. Maximum removal efficiency (70.96%) was observed on *L. Siceraria* peel followed by pedicle (51.58%) and mesocarp (29.57%) for adsorbent dosage of 0.10 g/L peel for 50 ml solution of sulfide (20 mg/L). The adsorption process was rapid, and it reached equilibrium in early 20-35 min for all adsorbents. Freundlich and Langmuir adsorption isotherms were employed for mathematical description of the adsorption equilibrium and to explain mechanism of adsorption. The maximum amounts of sulfide ions adsorbed, as evaluated by Langmuir isotherm, was 1.0436 mg/g of LS pedicle, 0.886 mg/g of peel and 0.843 mg/g of pedicle. The adsorption process conforms well to a pseudo second order kinetic model with best adsorption results at pH 8.

Keywords: Sulfide; Adsorption; *Lagenaria siceraria*; Isotherms and industrial effluents

Introduction

Sulfide (S^{2-}) is the inorganic chemical ionic form of sulfur. In nature, it occurs in deposits of coal, oil and in mineral ores (NRCC, 1977). Another source is the wastewater of various industries which is considered to be heavily adulterated with sulfide ions (Muruganathan *et al.*, 2004; Mckee and Wolf, 1963; USEPA, 1976; Booras, 1974; Colby and Smith, 1967). Through anaerobic decay of organic matter, sulfur found in fertilizers, fungicides and sewage, sulfate reducing bacteria generate sulfides (Mckee and Wolf, 1963; USEPA, 1976; Adelmah and smith, 1972; Acree and Splittstoesser, 1972). The sulfides and hydrogen sulfides of alkaline earth and alkali metals are water soluble (Sienko and Plane, 1974). Dissociation of soluble sulfide salts occurs into sulfide ions that respond with the hydrogen (H^+) ions in water, resulting in hydrogen sulfide (H_2S) and hydrosulfide ion (HS^-) formation. The pH of water plays a key role in the formation of these species. If the pH is low then the concentration of hydrogen sulfide increases (Mckee and Wolf, 1963; USEPA, 1976; Dar *et al.*, 2015).

From the last few decades the environmental toxicity on living entities has developed a main concern. A huge growth

in population and industrial sector is discharging wastewater continuously to the ecosystem which resulting in pollution and in due course toxicity to living being (Hossain *et al.*, 2012). Hydrogen sulfide is very poisonous even at trace levels (Zutshi, 1970). It is colorless and flammable with the rotten egg smell (Huynh *et al.*, 2011). The toxicity of H_2S is like of carbon monoxide (CO) and hydrogen cyanide (HCN) (Lindenmann, 2004) and it affects the organ system of human being. It has been reported that when H_2S associates with lipophilic molecules, it diffuses through the membranes and disseminate into skin, bones, liver, kidney and lungs (Muruganathan *et al.*, 2010; Hendrickson and Hamilton, 2004; Nikkanen and Burns, 2004; Brenneeman and James, 2000; Guidotti, 1994; Dorman *et al.*, 2002). A direct exposure of H_2S in body, even in minute concentration results in pain, tearing, blurred vision, photophobia, conjunctivitis, corneal bullae (Sax, 2014; Meyer, 1977). Hydrogen sulfide is also found to deactivate and degrade the key enzymes in human body (Muruganathan *et al.*, 2004).

For scientists, there is a challenge to adopt an environment friendly and sustainable method for removal of toxic sulfide species from wastewater (Hossain *et al.*, 2012). On large

*Corresponding author e-mail: parsadar@yahoo.com; a_dar_2@hotmail.com

scale H₂S is removed by aeration by employing complex packed tower aerators, simple air diffusers and mechanical aeration devices in wastewater treatment plants (Muruganathan *et al.*, 2004). Chemical oxidation method with commonly practiced oxidants KMnO₄, O₂, Cl₂, H₂O₂ and ClO⁻ is also used for H₂S removal from water (Muruganathan *et al.*, 2004; Edwards *et al.*, 2011; Tomar and Abdullah, 1994; Millero *et al.*, 1989). Manganese (IV) Oxide (MnO₂) for oxidation of sulfides in wastewater has also been reported (Valeika *et al.*, 2006).

Adsorption is a very cost effective, simple and regenerative technique for removal of toxic chemical species (Dar *et al.*, 2015). Activated carbon derived from biological wastes is a favorable method in the removal of sulfide species and other toxic metals. Feng *et al.* (2005) used activated carbon fibers for the adsorption of H₂S. Hanumantharao *et al.* (2012) used active carbon from *Lagenaria siceraria* shell for the adsorption of fluoride ions from aqueous solution. Plant wastes can be used as an adsorbent due to their free availability, inexpensive value, selective and good adsorption capacity. Untreated plant wastes have been used for the adsorption of pollutants (Saeed *et al.*, 2005; Babarinde *et al.*, 2006; King *et al.*, 2006; Hanafiah *et al.*, 2007; Hanafiah *et al.*, 2006b; Hanafiah *et al.*, 2006c; Karunasagar *et al.*, 2005; Quek *et al.*, 1998; Sawalha *et al.*, 2007b; Ho and Wang, 2004; Ho, 2003; Ho *et al.*, 2004; Bhattacharya *et al.*, 2006; Villaescusa *et al.*, 2004). The adsorption capacity of plant wastes can be increased by pretreatment with certain modifying agents such as base solution of sodium hydroxide before being used for the sanitization and the chelating efficacy can be enhanced by this pretreatment (Gaballah *et al.*, 1997; Ngah and Hanafiah, 2008).

Lagenaria siceraria belongs to the family *Cucurbitaceae*. In Pakistan, it locally named as Ghia/ Kaddu / Lauki, and its fruit is a common vegetable, which is plump with several seeds embedded in a spongy pulp (Dar *et al.*, 2014). The plant is an annual, softly pubescent and large climbing herb. The fruit is large, cylindrical, globose or flask shaped having constriction directly above the middle, green, densely hairy, plump, indehiscent, growing pale brown or yellowish, on ripening pulp drying out, sendoff a hard, thick hollow (Prajapati *et al.*, 2010).

A large quantity of mesocarp, peel and pedicle is produced daily in household garbage. For this reason, mesocarp, peel and pedicle of *Lagenaria siceraria* fruit have been tried as adsorbents for toxic sulfide ions from water by batch adsorption. The adsorbent was selected as it is easily available and cheap too. The nature of sorption has been effectively accounted and characterized by the kinetic, isothermal and surface characterization by FTIR. The aim of

this work is to find out ecofriendly adsorbent to eradicate sulfide from water.

Materials and methods

Fresh fruit of *L. siceraria* was collected from city Pattoki, District Kasur, Punjab (Pakistan). It was manually parted into peel, pedicle and mesocarp, washed, dried. All these parts in raw form was crushed and sieved through 60 mesh (ASTM) and stored in air tight glass jars separately.

Chemicals and reagents

All the reagents have analytical grade purity. A stock solution of sulfide was prepared by dissolving calculated quantity of sodium sulfide to make a certain concentration in double distilled water. Standard solutions were made by successive dilution of the stock solution. Ferric chloride (0.01 M) and 1, 10-phenanthroline (0.1%) were prepared in hot double-distilled water. Acetate buffer (CH₃COONa: CH₃COOH) was used to maintain pH at 4.

Batch adsorption experiment

Batch adsorption experiments were performed by using 20 mg/L sodium sulfide (Na₂S) solution (50 ml) at ambient temperature to adjust the different parameters. The experimental specifics are provided in the caption of respective figures. Amount of adsorbent was varied from 0.2–2.0 g. The required time to attain the equilibrium for adsorption was assessed in the range of 5–80 minutes. Parameter of shaking speed was optimized from 50–450 rpm at room temperature (293K). The effect of pH was studied in the range 3–11. In all experiments mesh size of LS mesocarp, pedicle and peel was 20 ASTM.

Equipment and apparatus

Sulfide estimation was done by classical iron-1, 10-phenanthroline complex formation method at 510nm, using spectrophotometer (HITACHI U1800). 1, 10-phenanthroline (2 ml), Acetate buffer (1 ml) and FeCl₃ (2 ml) was the reagents used for sulfide ions estimation, Fe³⁺ was reduced to Fe²⁺ to make the colored complex with 1, 10-phenanthroline. Double distilled water was used for further dilution of solutions. FTIR spectra were recorded from 4000 to 500 cm⁻¹ using Thermo Nicolet iS10 Spectrometer. The discs of samples were prepared by parting adsorbents with KBr (Merck; for spectroscopy) in a mortar pestle.

Adsorption isotherms and kinetics

Adsorption isotherms define the equilibrium relationship between adsorbent and adsorbate (Daret *et al.*, 2015). For biosorption processes kinetic studies have a significant role. Kinetics of adsorption not only defines the mechanism of

metal adsorption on adsorbents but it also define the rate of metal adsorption which wheels the contact time of metals at the solid-liquid interface (Ho and Mckay, 2000). The mechanism of adsorption determined by chemical and physical features of adsorbent and adsorbate, temperature, pH of medium, aids and mass transport process and contact time (Achak *et al.*, 2009).

Batch equilibrium experiments were performed with sulfide solutions ranging from 3-24mg/L (50 ml) at pH 8 and all optimized conditions for all the adsorbents. Linear forms of various isotherms were used to analyze respective parameters. Langmuir equation (Eq. 1) can be linearized by the following form:

$$\frac{1}{q_e} = \frac{1}{bq_m c_e} + \frac{1}{q_m} \quad (1)$$

Where “ C_e ” is the equilibrium concentration in liquid phase (mg/L), “ q_m ” is the monolayer adsorption capacity (mg/L) and “ b ” is the Langmuir constant linked to the free adsorption energy (L/mg).

Freundlich equation (Eq. 2) can be linearized by the following form:

$$\ln q_e = \ln k_f + \frac{1}{n} \ln c_e \quad (2)$$

Where “ K_f ” is a constant symbolic of the adsorption capacity of the adsorbent (mg/g) and the constant $1/n$ shows the intensity of the adsorption.

For kinetics studies, pseudo first order (eq. 3) and second order (eq. 4) kinetics models were employed.

$$\ln(q_e - q_t) = \ln q_e - \left\{ \frac{k_1}{2.303} \right\} t \quad (3)$$

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \left\{ \frac{1}{q_e} \right\} t \quad (4)$$

Where “ k_2 ” is the pseudo second order rate constant, “ q_e ” is the bio-sorption capacity at equilibrium and “ q_t ” is the bio-sorption capacity at time “ t ”.

Results and discussion

There is a critical environmental issue of treatment of industrial wastewater before it is discharged out. Effluents from leather industry are highly contaminated with organic and inorganic contaminants and large amounts of sulfide ions. Unlike organic wastes, these poisonous sulfide ions are non-biodegradable and they can be stored in living tissues, producing various disorders and diseases. Adsorption has ascended as an economical and modest way to treat effluents from industries. Utilizing economical and ecofriendly materials to adsorb pollutants is a primary choice of scientists to treat wastewater so, peel, pedicle and mesocarp of *L. siceraria* fruit has been used as an adsorbent to take out

sulfide ions from water. The fruit of *L. siceraria* is of sweet variety. As the biological and chemical constituents of this fruit concerned, it holds 0.6% fibers, 2.5% carbohydrate, 0.2% proteins, 0.5% mineral matter, <0.01% phosphorous and calcium. sodium, iodine and iron are also present as mineral elements. Amino group is also present in form of alanine, aspartic acid, leucines, phenylalanine, tyrosine etc. The edible portion (mesocarp) of fruit contains riboflavin, thiamine and ascorbic acid. The fruit skin (peel) contains 18.1% cellulose, 17.5% crude protein and 8.0% lignin (Prajapati *et al.*, 2010; Sivannarayana *et al.*, 2013; Tyagi *et al.*, 2012).

FTIR analysis of adsorbent

Comparison of the Fourier transform infrared (FTIR) spectrum for the adsorbents; mesocarp (Fig.1), pedicle (Fig. 2) and peel (Fig. 3) before and after absorption has been recorded. Core functional groups that can be active in adsorption are –OH, –CH, C=O, C–H, C–O, S=O Gangwal *et al.*, 2010; Pavia *et al.*, 2001; Heneczowski *et al.*, 2001; Kumar *et al.*, 2013; Marimuthu and Gurumoorthi, 2013). It shows that surface of LS mesocarp, pedicle and peel can arrange for various direct and indirect chemical and physical interfaces with sulfide ions in water. A direct interface takes place because of attraction between functional groups bearing positive charge and negative sulfide ions. Various mineral elements in water and water itself can play a role of mediated interactions for an indirect attraction between negatively charged functional groups and sulfide ions by process of chemisorption, such as –O–Na⁺–S²⁻ and –O–HOH–S²⁻. On the other hand, evidence and other supporting information about such interactions are limited in literature. A comparative study by FTIR analysis has been done to study the presence and identification of functional groups present in the adsorbents, before and after adsorption of sulfide ions. After adsorption minute shift in peak positions and emergence of various new peaks were observed in spectra (Table-I). These changes were due to the binding of sulfide ions with functional groups of the adsorbents. Thus, shifting of bands confirms the adsorption of sulfide ions on the surface of *L. siceraria* mesocarp, pedicle and peel.

Effect of adsorbent dose

The influence of mesocarp, pedicle and peel, on adsorption of 50 ml solution of sulfide (20 mg/L) was studied with different dosage in the range, 0.10–1.50 g/L. Results disclosed that the adsorption efficiency is very much dependent on added quantity of adsorbents. Maximum removal was 29.57%, 51.58% and 70.96% for doses of 0.10 g/L of mesocarp, pedicle and peel respectively (Fig. 4) for 50 ml solution of sulfide (20 mg/L). The inclination in graph is because of the intermediation between adsorbent or it may be the failure of

Table I. Comparative FTIR analysis of *Lagenaria siceraria* mesocarp, pedicle and peel, before and after adsorption

FTIR Analysis					
LS mesocarp (B)*	LS mesocarp (A)*	LS pedicle (B)	LS pedicle (A)	LS peel (B)	LS peel (A)
3327.34 -OH (H-bonded)	3299.55 -OH (H-bonded)	3334.15 -OH (H-bonded)	3327.65 -OH (H-bonded)	-	3735.03 -OH (Free)
2918.83 -OH (carboxylic acid)	2923.99 -OH (carboxylic acid)	-	2918.60 -OH (carboxylic acid)	-	3648.59 -OH (Free)
-	2853.13 -CH (str. Alkane)	1634.92 C=O (amide)	1634.28 C=O (amide)	3291.36 -OH (H-bonded)	3291.55 -OH (H-bonded)
-	1729.96 C=O (aldehyde)	-	1417.74 C=C (aromatic)	2923.76 -OH (carboxylic acid)	2922.17 -OH (carboxylic acid)
1617.27 C=O (amide)	1645.92 C=O (amide)	-	-	-	2359.40 CN
-	1516.15 C=O (with phenyl)	1031.49 C-O Or S=O (sulfoxide)	1031.68 C-O Or S=O (sulfoxide)	-	1716.52 C=O (aldehyde)
1418.22 C-H	1463.88 C-H	-	-	1629.11 C=O (amide)	1635.79 C=O (amide)
-	-	-	-	1515.60 (with phenyl)	1540.48 (with phenyl)
1242.83 C-O (acids)	1238.27 C-O (acids)	-	-	-	1456.71 C-H
-	1165.65 C-O	-	-	-	1373.80 C-H
1032.08 C-O Or S=O (sulfoxide)	1032.24 C-O Or S=O (sulfoxide)	-	-	1231.81 C-O (acids)	1238.27 C-O (acids)
-	-	-	-	1027.12 C-O Or S=O (sulfoxide)	1031.55 C-O Or S=O (sulfoxide)

* B (Before Adsorption), A* (After Adsorption)

metal ions in solution with respect to open binding sites (Dar *et al.*, 2015; Anwar *et al.*, 2010). A quick adsorption was observed in start when sufficient sulfide ions were available for numerous bareadsorption sites. On the other hand, there was no momentous change in adsorption after 0.90 g/L addition of adsorbent. Consequently, the ideal adsorbent dosage was selected as 0.10 g/L each of mesocarp, pedicle and peel for sulfide and pedicle showed maximum removal efficiency.

Effect of contact time

Contact time was assessed as an important aspect affecting adsorption efficiency. By using 0.10 g of adsorbent in 50 ml of 20 mg/L sulfide solution, time of contact of adsorption has been changed from 5-65 min for experiments. In case of mesocarp as adsorbent, sulfide removal efficiency was 57.74% during early 20 min and after that, no significance rise or decline has been observed. For pedicle as adsorbent,

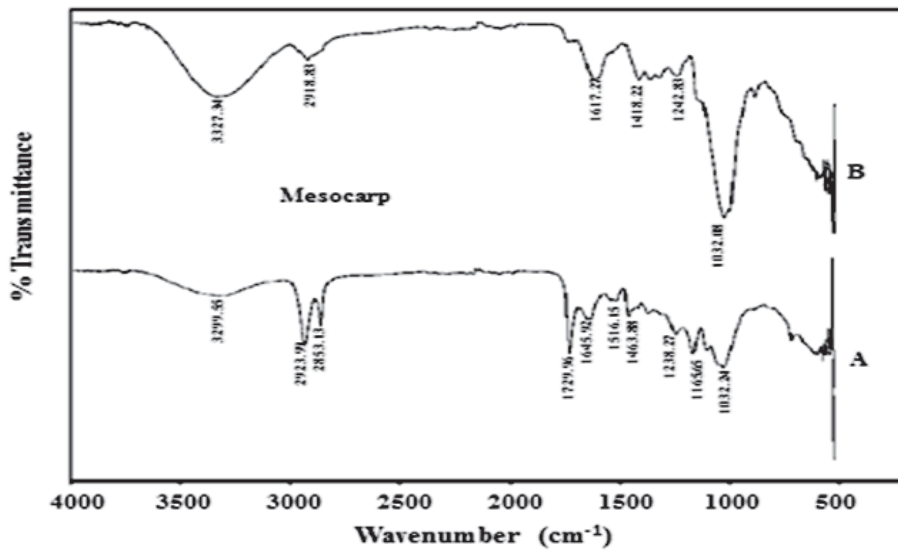


Fig. 1. FTIR spectrum of mesocarp (B=Before Adsorption, A=After Adsorption)

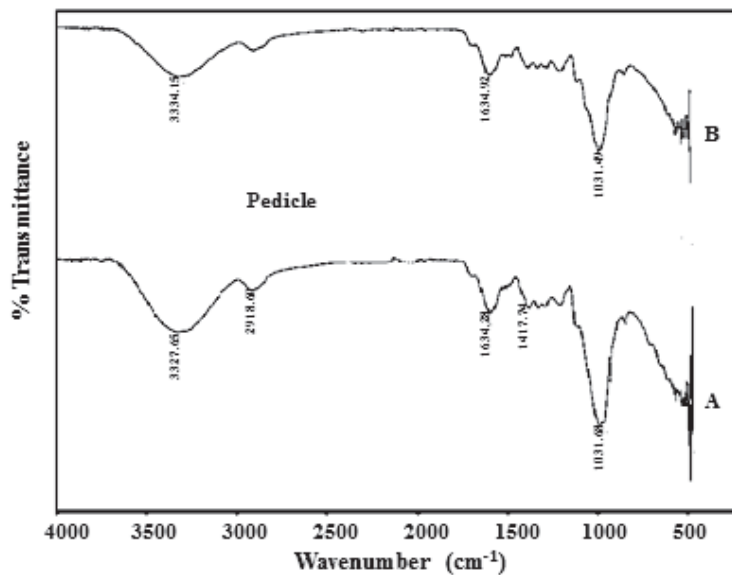


Fig. 2. FTIR spectrum pedicle (B=Before Adsorption, A=After Adsorption)

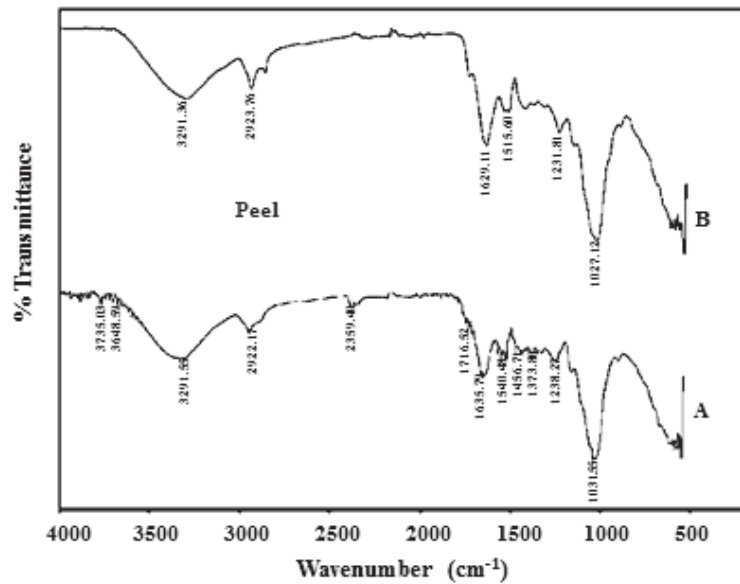


Fig. 3. FTIR spectrum peel (B=Before Adsorption, A=After Adsorption)

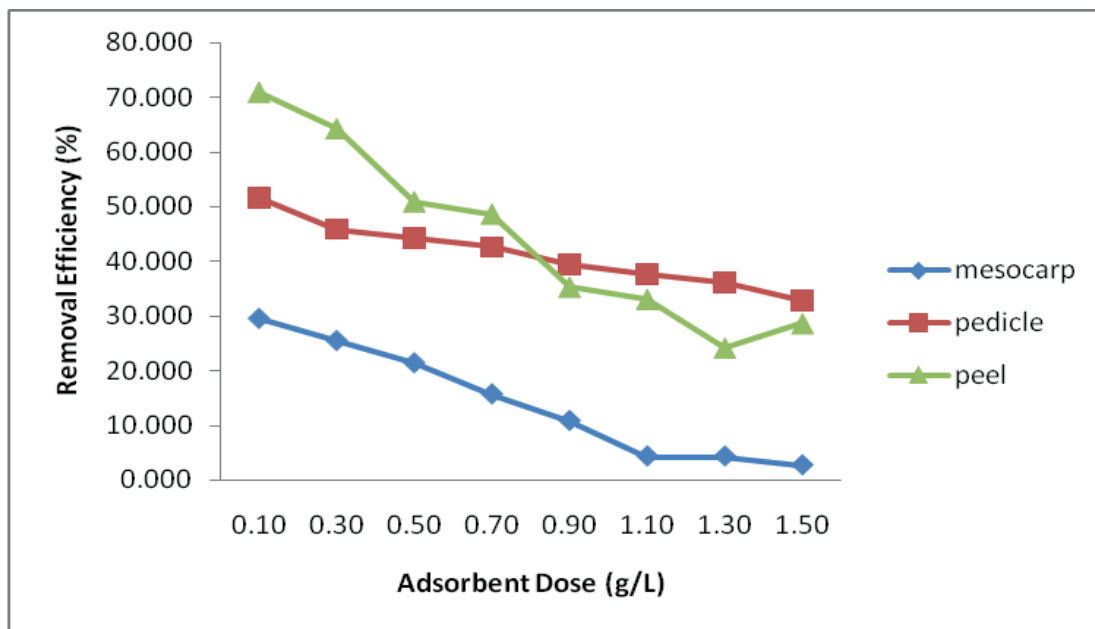


Fig. 4. Effect of adsorbent dose on removal of sulfide ions, sulfide ions 20 mg/L, volume 50 ml; mesocarp, pedicle, peel 0.10-1.50 g; pH = 8; time 40 min (for mesocarp), 20 min (for pedicle), 35 min (for peel); agitation speed 250 rpm (for mesocarp and pedicle) 50 rpm (for peel); temperature = 293K.

sulfide removal efficiency was 58.23% during early 40 min and in case of peel as adsorbent, sulfide removal efficiency was 40.13% during early 35 min. It can be recommended that

under given conditions, maximum adsorption has taken place and equilibrium has been attained in 20, 40, 35 min for mesocarp, pedicle and peel as adsorbent respectively (Fig.5).

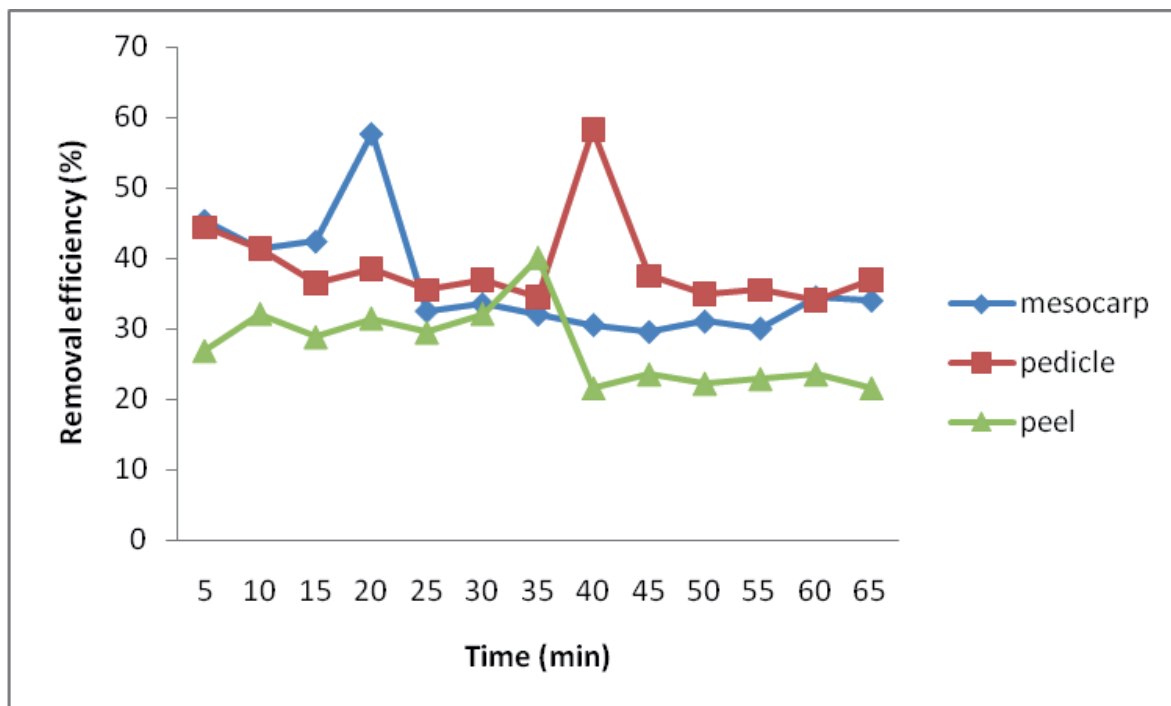


Fig. 5. Time of contact versus adsorption efficiency of sulfide ions on mesocarp, pedicle and peel; sulfide ions 20 mg/L; volume 50 ml; mesocarp, pedicle and peel 0.10 g; pH = 8, time 5-65 min; temperature 293K.

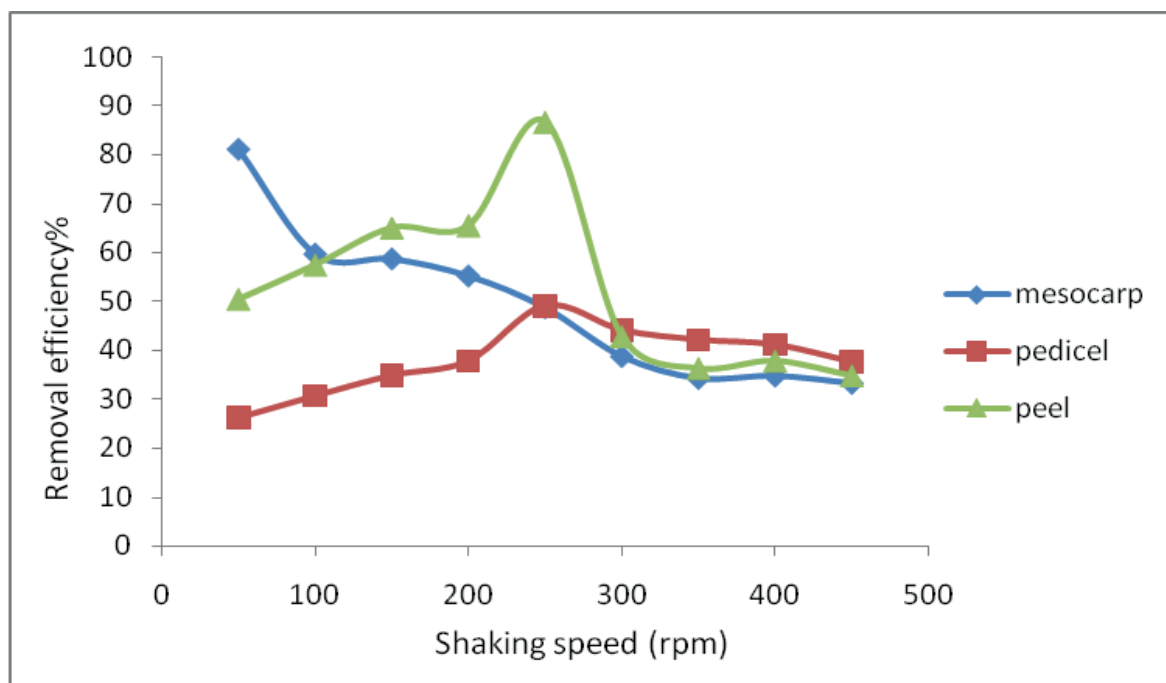


Fig. 6. Agitation speed versus adsorption efficiency of sulfide ions on mesocarp, pedicle and peel; sulfide ions 20 mg/L; volume 50 ml; mesocarp, Pedicle and peel 0.10 g; pH = 8; time 40 min (for mesocarp), 20 min (for pedicle), 35 min (for peel); agitation speed 50-450 rpm for mesocarp, pedicle and peel; temperature = 293K.

Effect of agitation speed

Agitating speed of 50 ml aqueous solutions of sulfide (20 mg/L) with mesocarp, pedicle and peel particles (0.10 g) changed from 50-450 rpm. In case of mesocarp, adsorption efficiency was maximum 86.68% at 250 rpm then it started to reduce gradually. For pedicle adsorption efficiency was 49.16% at 250 rpm and peel showed adsorption efficiency 81.60% at 50 rpm. When agitation speed was less, adsorbent (peel) disperse in the solution and made maximum contact with adsorbate which gave the high adsorption. At high agitation speed, adsorbent-adsorbate, adsorbent-adsorbent and adsorbate-adsorbate collision occurred which result in suppressed adsorbent sites and did not allow sufficient time to the adsorbate to adsorb onto the surface of adsorbents (Dar *et al.*, 2015; Anwar *et al.*, 2010; Anwar *et al.*, 2009) so, adsorption efficiency of peel reduced but for mesocarp and pedicle adsorption efficiency increase with maximum contact with adsorbate (Fig.6).

Effect of pH

For the adsorption of metal ions, an important factor that plays its role is the pH of solution. The pH effect on adsorption was studied in range 3-11. As shown in Fig. 7,

maximum adsorption of sulfide ions was observed at pH 8 for all of three adsorbents but peel has been shown maximum adsorption efficiency 76.32 % (mg/g). Thus we can say at higher pH, adsorption sites trigger. Away from optimum pH, adsorption of sulfide ions decreases, as the sulfide ions jumps to less negative (HS^-) or neutral hydrogen sulfide (H_2S) and adsorbents start to protonate and develop more positive charge (Dar *et al.*, 2015). This outcomes in reduce attraction between adsorbents and adsorbate. Adsorption of sulfide ions on *Lagenaria siceraria* mesocarp pedicle and peel is electrostatic by nature which depends on pH.

Adsorption isotherms and kinetics study

Table II contains parameters and correlation coefficients for Freundlich and Langmuir isotherms. Freundlich isotherm for sulfide ions adsorption on LS mesocarp, pedicle and peel is shown in Fig.8a-8c. The plotted isotherms holds good for the adsorption of sulfide ions by adsorbents as indicated by their R_2 values that are approaching to 1. Parameter 'n' indicated the adsorption intensity measure for sulfide ion on adsorbents. Ultimate adsorption capacity ' K_f ' of sulfide ions, as calculated from

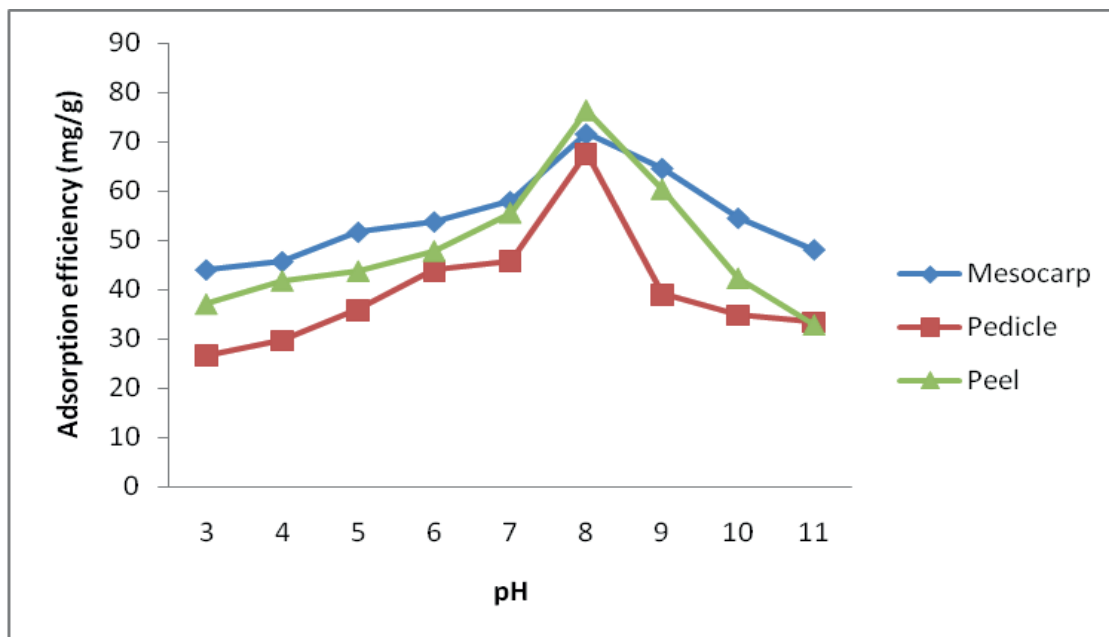


Fig. 7. Effect of pH on adsorption efficiency of sulfide ions on mesocarp, pedicle and peel; sulfide ions 20 mg/L; volume 50 ml; mesocarp, Pedicle and peel 0.10 g; pH = 8; time 40 min (for mesocarp), 20 min (for pedicle), 35 min (for peel); agitation speed 50-450 rpm for mesocarp, pedicle and peel; temperature = 293K.

Table II. Comparison of parameters of different isotherm models

Adsorbent	Isotherm	Freundlich	Langmuir
	R^2	0.9389	0.9066
Mesocarp	Parameters	$n = 1.1359$	$q_m = 0.843\text{mg/g}$
		$K_F = 0.0862\text{ mg/g}$	$b = 0.1365\text{ L/g}$
	R^2	0.8744	0.8869
Pedicle	Parameters	$n = 0.9772$	$q_m = 1.0436\text{ mg/g}$
		$K_F = 0.0589\text{mg/g}$	$b = 0.0766\text{ L/g}$
	R^2	0.9459	0.9571
Peel	Parameters	$n = 1.4865$	$q_m = 0.886\text{ mg/g}$
		$K_F = 0.120\text{mg/g}$	$b = 0.147\text{ L/g}$

Freundlich isotherm is 0.0862, 0.0589 and 0.120 mg/g for mesocarp, pedicle and peel respectively, indicating peel as a good adsorbent by studying this model.

Langmuir isotherm for sulfide ions adsorption on LS mesocarp, pedicle and peel is shown in Fig.9a-9c and the related correlation coefficients and parameters of the isotherm are given in Table 2. Maximum adsorption capacity ' q_m ' is 0.843, 1.0436 and 0.886 mg/g for mesocarp, pedicle and peel respectively showing maximum Langmuir monolayer coverage of sulfide ions by 0.10 g LS pedicle. Correlation coefficient ' R_2 ' values approaching to one, evidently submit that Langmuir isotherm holds good to describe adsorption of sulfide ions on used adsorbents. Energy of sorption ' b ' for mesocarp, pedicle and peel is 0.1365, 0.0766 and 0.147 L/g respectively.

By studying the reaction kinetics, adsorption mechanism of sulfide ions was assessed. Pseudo first and second order reaction kinetics models were used for this purpose. It was

Table III. Comparison of Pseudo first and second order reaction kinetics model parameters

Adsorbent	Pseudo first Order	Pseudo Second Order
	$R^2 = 0.0252$	$R^2 = 0.9682$
Mesocarp	$q_m = 0.0018\text{mg/g}$	$q_m = 0.3020\text{mg/g}$
	$R^2 = 0.7269$	$R^2 = 0.9508$
Pedicle	$q_m = 0.0032\text{mg/g}$	$q_m = 0.3536\text{ mg/g}$
	$R^2 = 0.2482$	$R^2 = 0.9558$
Peel	$q_m = 0.0072\text{mg/g}$	$q_m = 0.2078\text{ mg/g}$

found that for the adsorption of sulfide ions, adsorbents followed the pseudo second order kinetic model. Maximum adsorption capacity (q_m) of 0.3536 mg/g was obtained for pedicle (as shown in Table 3) with pseudo second order kinetic model followed by mesocarp and peel.

Conclusion

The present study can be used to overcome water pollution using eco-friendly and cost effective agro-waste constituents like *Lagenaria siceraria* mesocarp, pedicle and peel for the removal of toxic sulfide ions from industrial effluents and wastewaters. All of the three adsorbents show good adsorption results but maximum adsorption capacity of *L. siceraria* pedicle which is termed as Langmuir complete monolayer coverage ' q_m ' shows that 0.10 g of LS pedicle can adsorb 1.0436 mg/g of sulfide ions which is comparable with 0.886 mg/g of peel and 0.843 mg/g of pedicle. Comprehensive analysis of experimental data was conceded by using adsorption isotherms, to describe adsorption of sulfide ions *L. siceraria* mesocarp, pedicle and peel. Adsorbents followed the pseudo second order kinetic model for adsorption of sulfide ions. A physical form of nature was found in this process of adsorption. It is concluded that *L. siceraria* pedicle can be used a green filter for removal of sulfide ions from effluents of tanneries and other industrial wastes containing sulfide as a dangerous species.

Acknowledgements

The authors are thankful to Mr. Farhan Mehmood and Usama Waqas for their technical assistance regarding this research work.

References

- Achak M, Hafidi A, Ouazzani N, Sayadic S and Mandi L (2009), Low cost biosorbent “banana peel” for the removal of phenolic compounds from olive mill wastewater: Kinetic and equilibrium studies, *J. Hazard. Mater.* **166**: 117-125.
- Acree TE and Splittstoesser DF (1972), Prevention of H₂S in wine fermentation, *N.Y. Food Life Sci. Q.* **5**: 19.
- Adelmah IR and Smith LL (1972), Toxicity of hydrogen sulphide to goldfish (*Carassius auratus*) as influenced by temperature, oxygen, and bioassay techniques, *J. Fish. Res. Board Can.* **29**: 1309.
- Anwar J, Shafique U, Waheed-uz-Zaman, Salman M, Dar A, Anwar S (2010), Removal of Pb (II) and Cd (II) from water by adsorption on peels of banana, *Biores. Technol* **101**: 1752-1755.
- Anwar J, Shafique U, Salman M and Waheed-uz-Zaman, (2009), Removal of chromium (III) by using coal as adsorbent, *J. Hazard. Mater.* **171**: 797-801.
- Babarinde NAA, Babalola JO and Sanni RA (2006), Biosorption of lead ions from aqueous solution by maize leaf, *Int. J. Phys. Sci.* **1**: 23-26.
- Bhattacharya AK, Mandal SN and Das SK (2006), Adsorption of Zn (II) from aqueous solution by using different adsorbents, *Chem. Eng. J.* **123**: 43-51.
- Booras SG (1974), Hydrogen sulphide health effects and recommended air quality standard, NTIS PB-233843, Illinois Institute for Environmental Quality.
- Brenneman KA, James RA, Gross EA and Dorman DC (2000) Olfactory neuron loss in adult male CD rats following subchronic inhalation exposure to hydrogen sulfide, *Toxicol. Pathol.* **28**, 326-333.
- Colby PJ and Smith LL (1967), Survival of walleye eggs and fry on paper fibre sludge deposits in Rainy River, *Am. Fish. Soc.* **96**: 278.
- Dar A, Shafique U, Anwar J and Munawar MA (2015), Removal of sulfide ions from water using rice husk, *J Sulfur Chem.* **36**: 187-195.
- Dar P, Ahmed D, Waqas U, Saeed R and Chaudhery R (2014), Comparative analysis of antimicrobial potential of peel and mesocarp of *Lagenaria siceraria* fruit extracts in various solvents against clinically important pathogens, *Phol.* **3**: 100-105.
- Dorman DC, Moulin FJM, McManus BE, Mahle KC, James RA, Struve MF (2002), Cytochrome oxidase inhibition induced by acute hydrogen sulfide inhalation: Correlation with tissue sulfide concentrations in the rat brain, liver, lung, and nasal epithelium, *Toxicol. Sci.* **65**: 18-25.
- Edwards S, Alharthi R, Ghaly AE (2011), Removal of hydrogen sulphide from water, *Am. J. Environ. Sci.* **7**: 295-305.
- Feng W, Kwon S, Borguet E, Vidic R (2005), Adsorption of hydrogen sulfide onto activated carbon fibers: Effect of pore structure and surface chemistry, *Environ. Sci. Technol.* **39**: 9744-9749.
- Gaballah I, Goy D, Allain E, Kilbertus G and Thauront J (1997), Recovery of copper through decontamination of synthetic solutions using modified barks, *Met. Trans.* **B28**: 13-23.
- Gangwal A, Parmar SK and Sheth NR (2010), Triterpenoid, flavonoids and sterols from *Lagenaria siceraria* fruits, *Der Pharmacia Lettre.* **2**: 307-317.
- Guidotti TL (1994), Occupational exposure to hydrogen-sulfide in the sour gas-industry-some unresolved issues, *Int. Arch. Occup. Environ. Health.* **66**: 153-160.
- Hanafiah MAK, Ngah, Zakaria H and Ibrahim SC (2007), Batch study of liquid-phase adsorption of lead ions using Lalang (*Imperata cylindrica*) leaf powder, *J. Biol. Sci.* **7**: 222-230.
- Hanafiah MAK, Ngah WSW, Ibrahim SC, Zakaria H, Ilias WAHW (2006b), Kinetics and thermodynamic study of lead adsorption onto rubber (*Hevea brasiliensis*) leaf powder, *J. Appl. Sci.* **6**: 2762-2767.
- Hanafiah MAK, Shafiei S, Harun MK, Yahya MZA (2006c), Kinetic and thermodynamic study of Cd²⁺ adsorption onto rubber tree (*Hevea brasiliensis*) leaf powder, *Mater. Sci. Forum.* **517**: 217-221.
- Hanumantharao Y, Kishore M and Ravindhranath K (2012), Characterization and adsorption studies of “*Lagenaria siceraria*” shell carbon for the removal of fluoride, *Int. J. Chem Tech. Res.* **4**: 1686-1700.
- Hendrickson RG, Chang A, Hamilton RJ (2004), Co-worker fatalities from hydrogen sulfide, *Am. J. Ind. Med.* **45**: 346-350.

- Henczowski M, Kopacz M, Nowak D and Kuźniar A (2001), Infrared spectrum analysis of some flavonoids, *Acta Pol Pharm.* **58**: 415-20.
- HoYS, Chiu WT, Hsu CS and Huang CT (2004), Sorption of lead ions from aqueous solution using tree fern as a sorbent, *Hydrometallurgy* **73**: 55-61.
- Ho YS (2003), Removal of copper ions from aqueous solution by tree fern, *Water Res.* **37**: 2323-2330.
- Ho YS, McKay G (2000), The kinetics of divalent metal ions onto sphagnum moss peat, *Water Res.* **34**: 735-742.
- HoYS and Wang CC (2004), Pseudo-isotherms for the sorption of cadmium ion onto tree fern, *Process Biochem.* **39**: 759-763.
- Hossain MA, Ngo HH, Guo WS and Nguyen TV (2012), Removal of copper from water by adsorption onto banana peel as bioadsorbent, *Int. J. of Geomate.* **2**: 227-234.
- Huynh Q Thieu VQQ, Dinh TP and Akiyoshi S (2011), Removal of hydrogen sulfide (H₂S) from biogas by adsorption method, 8th Biomass-Asia workshop, Biomass as sustainable energy and environmental technology, with Asia biomass office conference, Thuy Khue-Tay Ho-Hanoi, Vietnam.
- Johnson PD, Watson WA, Brown J and Jefcoat IA (2002), Peanut hull pellets as a single use sorbent for the capture of Cu (II) from wastewater, *Waste Manage.* **22**: 471-480.
- Kadota H and Ishida Y (1972), Production of volatile sulphur compounds in microorganisms, *Annu. Rev. Microbiol* **26**: 127.
- Karunasagar D, Krishna MVB, Rao SV and Arunachalam J (2005), Removal of pre concentration of inorganic and methyl mercury from aqueous media using a sorbent prepared from the plant *Coriandrum sativum*, *J. Hazard. Mater.* **B118**: 133-139.
- King P, Srivinas P, Kumar YP and Prasad VSRK (2006), Sorption of copper (II) ion from aqueous solution by *Tectona grandis* (teak leaves powder), *J. Hazard. Mater.* **B136**: 560-566.
- Kumar BP, Sindhuri M, Jyothshna K, Kumar SV, Manogna A and Madhavi P (2013), Isolation and characterization of natural mucilage from *Lagenaria siceraria*, *Inter. Res. J. Pharm.* **4**: 117-121.
- Lindenmann J, Matzi V, Neuboek N, Ratzenhofer-Komenda B, Maier A and Juettner S (2010), Diving and hyperbaric medicine, *J. South Pacific Underw. Med. Soc.* **40**: 213-217.
- Marimuthu M and Gurumoorthi P (2013), Phytochemical screening and FTIR studies on wild and common south Indian Legumes, *Asian J Pharm Clin Res.* **6**: 141-144.
- McKee JE and Wolf HW (1963), Water quality criteria. 2nd Ed., Resources Agency of California State Water Resources Control Board, California.
- Meyer B (1977), Sulfur: Energy and Environment, *Zeitschrift fur Chemie.* **18(7)**: 279.
- Millero FJ, LeFerriere A, Fernandez M, Hubinger S and Hershey JP (1989), Oxidation of hydrogen sulfide with hydrogen peroxide in natural waters, *Environ. Sci. Technol.* **23**: 209-213.
- Muruganathan M, Raju GB and Prabhakar S (2004), Removal of sulfide, sulfate and sulfite ions by electro coagulation, *J. Hazard. Mater.* **109**: 37-44.
- National Research Council of Canada (NRCC) (1977), Sulphur and its inorganic derivatives in the Canadian environment, NRCC No. 15015, Associate Committee on Scientific Criteria for Environmental Quality, Ottawa.
- Ngah WSW and Hanafiah MAKM (2008), Removal of heavy metal ions from wastewater by chemically modified plant wastes as adsorbents: A review, *Biores. Technol.* **99**: 3935-3948.
- Nikkanen HE and Burns MM (2004), Severe hydrogen sulfide exposure in a working adolescent, *Pediatrics.* **113**: 927-929.
- Pavia DL, Lampman GM and Kriz GS (2001), Introduction to Spectroscopy, 3rd Ed., Thomson Learning Inc., United States.
- Prajapati RP, Kalariya M, Parmar SK and Sheth NR (2010), Phytochemical and pharmacological review of *Lagenaria siceraria*, *J. Ayurveda & Integr. Med.* **1**: 266-272.
- Quek SY, Wase DAJ and Forster CF (1998), The use of sago waste for the sorption of lead and copper, *Water SA.* **24**, 251-256.

- Rahman NNNA, Shahadat M, Won CA and Omar FM (2014), FTIR study and bioadsorption kinetics of bioadsorbent for the analysis of metal pollutants, *RSC Adv.* **4**: 58156-58163.
- Sax NI (2014), *Sax's Dangerous Properties of Industrial Materials*, 12th Ed., Van Nostrand Reinhold Company, New York.
- Saeed A, Akhter MW and Iqbal M (2005), Removal and recovery of heavy metals from aqueous solution using papaya wood as a new biosorbent, *Sep. Purif. Technol.* **45**: 25-31.
- Sawalha MF, Peralta-Videa JR, Romero-Gonza'lez J, Duarte-Gardea M and Gardea-Torresdey JL (2007a), Thermodynamic and isotherm studies of the biosorption of Cu (II), Pb (II) and Zn (II) by leaves of saltbush (*Atriplex canescens*), *J. Chem. Thermodyn.* **39**: 488-492.
- Sawalha MF, Peralta-Videa JR, Romero-Gonza'lez J and Gardea-Torresdey JL (2007b), Biosorption of Cd (II), Cr (III), and Cr (VI) by saltbush (*Atriplex canescens*) biomass: thermodynamic and isotherm studies, *J. Colloid Interface Sci.* **300**: 100-104.
- Sienko MJ and Plane RA (1974), *Chemical principles and properties*, 2nd Ed., McGraw-Hill Publishing Co., New York.
- Sivannarayana T, Hussain SS and Jithendra KP (2013), Pharmacological and pharmaceutical applications of *Lagenaria siceraria* (bottle gourd), *Inter.J. Inven. Pharmc. Sci.* **1**: 288-292.
- Tomar M and Abdullah TH (1994), Evaluation of chemicals to control the generation of malodorous hydrogen sulfide in waste water, *Water Res.* **28**: 2545-2552.
- Tyagi N, Sharma GN and Hooda V (2012), Phytochemical and pharmacological profile of *Lagenaria siceraria*: An overview, *Inter. Res. J. Pharm.* **3**: 1-4.
- USEPA (U.S. Environmental Protection Agency) (1976), *Quality criteria for water*, Washington DC, p 410.
- Valeika V, Beleska K and Valeikiene V (2006), Oxidation of sulphides in tannery wastewater by use of manganese (IV) oxide, *Polish J. of Environ. Stud.* **15**: 623-629.
- Villaescusa I, Fiol N, Mart'nez M, Miralles N, Pocj J and Serarols J (2004), Removal of copper and nickel ions from aqueous solutions by grape stalks wastes, *Water Res.* **38**: 992-1002.
- Zutshi P (1970), Optimum physical and chemical conditions for the Methylene Blue method of estimating gaseous sulphide in the environment, *Talanta.* **17**: 1014-1018.

Received: 07 February 2017; Revised: 30 April 2017;

Accepted: 29 May 2017.