MOLECULAR IDENTIFICATION OF LAVENDULA DENTATA L., MENTHA LONGIFOLIA (L.) HUDS. AND MENTHA × PIPERITA L. BY DNA BARCODES

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Abstract

Five DNA barcodes were tested for identification and discrimination of Lavendula dentata L., Mentha longifolia (L.) Huds. and Mentha \times piperita L. New DNA barcodes have been registered for L. dentata from Taif, Saudi Arabia. The separate clading of L. dentata and M. longifolia through the phylogenic analyses proved their endemism to Saudi Arabia. The phylogenetic trees revealed from the ITS2, matK and trnH data demonstrated that all Mentha species formed monophyletic clusters except hybrid M. \times piperita from Taif which formed separate clades distinguishing it from the two parents; M. aquatica L. and M. spicata L. DNA barcoding could be considered as a good approach for distinguishing and identifying the mint plants, though it was not possible to confirm the relationship between hybrids and their putative parents.

Introduction

The family Lamiaceae comprising about 7,173 species under 236 genera possesses medicinal and aromatic herbs such as lavender, basil, mint, rosemary and thyme, that have been widely utilized as teas, spices, traditional medicines or raw material for the food and pharmaceutical industries (Theodoridis *et al.*, 2012).

Lavendula dentata is one of five naturally growing lavender species in Saudi Arabia that has been known as the main center of origin of the genus (Miller, 1985). Lavender species as medicinal plants, are distributed in highlands of Albaha, Asir, and Taif and are exploited for the production of high-quality lavender honeys. Locally known as Habak, Al-Madinah mint or wild mint, Mentha longifolia and peppermint, Mentha × piperita are present in the spontaneous flora of Saudi Arabia but also under cultivation. Traditionally, they have been used as medicinal agents to treat colds, cough, headaches, asthma and digestive disorders. Recent studies proved the antiviral, antimicrobial, antioxidant, anti-inflammatory and anticancer characteristics as therapeutic activities for the extracts derived from Mentha species (Anwar et al., 2017). Hybridization and polyploidy play an important role in the speciation of the members belonging to genus Mentha such as M. × piperita that is considered as a hybrid of the two mints; M. spicata and M. aquatica (Mogosan et al., 2017) making them good targets for molecular studies.

Various studies have been performed to identify and classify species of Lamiaceae collected from Saudi Arabia based on anatomical and cytological studies (Abdel Khalik, 2016) and biochemical analyses (Kasem, 2016), however, very little is known about DNA barcoding information. DNA sequences for the species under study will be compared in a database against retrieved sequences of identified individuals from the GenBank. If the query sequence matches with one in the database, this will help in identification, discrimination or gaining a new barcodes for these species (Hajibabaei *et al.*, 2007). Therefore, the objectives of this research include:

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i) utility of specific DNA regions, two nuclear internal transcribed spacers (ITS and ITS2) and the plastid DNA regions (rbcL, matK and trnH) for developing DNA barcodes and subsequently identification for the three species; $Lavendula\ dentata$, $Mentha\ longifolia$ and $M. \times piperita$ occurred in Taif highlands of Saudi Arabia; ii) discriminating between species under study and those retrieved from the GenBank and iii) exploring the interspecific variation between M. longifolia and M. $\times piperita$.

Materials and Methods

Plant materials

Two wild species, namely *Lavendula dentata* and *Mentha longifolia*, and the hybrid species, *M.* × *piperita* belonging to family Lamiaceae were collected from Taif highlands, Saudi Arabia. Species identification was confirmed following Collenette (1999).

DNA extraction and amplification

DNA of fresh young leaves was extracted using CTAB method as described by Doyle and Doyle (1987). The purified DNA was amplified for ITS, ITS2, *rbc*L, *mat*K and *trn*H barcodes using universal primers.

PCR sequencing

The PCR products of the three Lamiaceae species for the five DNA barcodes were purified and sequenced at Macrogen Inc., South Korea. All sequences of the three species generated in this research were deposited in GenBank (accession numbers are listed in Table 1).

Sequences alignment and phylogenetic analyses

The sequences of ITS, ITS2, matK, rbcL and trnH of L. dentata, M. longifolia and M. \times piperita were subjected to BLAST (http://blast.ncbi.nlm.nih.gov/Blast.cgi) to confirm them from the other related Lamiaceae species existing in the GenBank database. Sequence alignments were performed by MUSCLE algorithm (Edgar, 2004; Tamura et al., 2013). The equality of evolutionary rate parameters between sequences of the three species under study and the retrieved species from GenBank were calculated by Tajima's relative rate test (Tajima, 1993). Nucleotide substitution rates and Transition/Transversion bias (R) were estimated using Maximum Likelihood method. The phylogenic trees were constructed by the Maximum likelihood bootstrap (MLB) analysis. A total of 1,000 bootstrap replicates were performed. The software of MEGA6 was used for all operations (Tamura et al., 2013).

Table 1. Accession numbers in GenBank of sequences of *Lavendula dentata*, *Mentha longifolia* and *M*. × *piperita* generated in this study.

Taxa	ITS	ITS2	matK	rbcL	trnH
L. dentata	LC373552.1	LC373553.1	-	LC373554.1	LC373555.1
M. longifolia	-	LC378378.1	-	LC378379.1	-
$M. \times piperita$	-	LC374287.1	LC374288.1	LC374289.1	LC374290.1

Results and Discussion

Identification of Lavendula dentata

Sequences of *L. dentata* for ITS, ITS2, *rbc*L and *trn*H barcoding loci were submitted to BLAST at the GenBank database, however, any sequence of *L. dentata* was detected in the database, thus the present study succeeded in registering new DNA barcodes for *L. dentata* from

Taif. Sequences of species belonging to the genus Lavendula showing high similarities to those of L. dentata were retrieved for the statistical analyses. ITS showed the highest sequence length (775 bp) followed by rbcL (537 bp), ITS2 (358 bp) and trnH (346 bp), whereas, the variable sites percentage after alignment was higher in trnH (24%) than those of ITS, ITS2 and rbcL. The GC ratios scored in loci ITS and ITS2 (60.1 and 65.9) was found greater than those of rbcL and trnH (Table 2). In comparison with the retrieved Lavendula species, the rates of transitions to transversions showed notable substitution changes in the sequences of L. dentata (Table 2). Transitions generally occurred more than transversions. Transition/transversion bias (R) was found relatively high and ranged from 1.19 to 2.82 demonstrating a molecular evolution within Lavendula genome. This putative evolution in L. dentata was confirmed through tests of Tajima relative evolutionary rate that displayed an accelerated rates of evolution (P-values <0.05) for all loci under study. The results revealed that ITS, ITS2, rbcL and trnH have sufficient efficiency in sequence quality as well as in species identification across the genome of the genus Lavendula. For further identification of L. dentata, sequences of the four loci were used to reconstruct four phylogenetic trees (Fig. 1). Except the tree revealed from ITS2, the separate clustering of L. dentata in the phylogenetic trees of ITS, rbcL and trnH proved its endemism to Saudi Arabia. The development of different DNA barcodes is better than single locus for more accurate results (Khan et al., 2013). The identification of species within a community through DNA barcodes contributes to the construction of the barcode library for terrestrial plants (Burgess et al., 2011).

Table 2. Statistics derived from the sequencing, alignment and BLAST processes for all loci employed in the present investigation.

Parameters -		Loci						
		ITS2	rbcL	matK	trnH			
% Variable sites after alignment for Lavendula dentata	0.01	0.06	0.01	-	0.24			
% Variable sites after alignment for Mentha longifolia	0.11	0.12	0.04	-	-			
% Variable sites after alignment for $M. \times piperita$		0.12	0.04	0.29	0.31			
Sequence length of <i>L. dentata</i>	775	358	537	-	346			
Sequence length of <i>M. longifolia</i>	362	347	528	-	-			
Sequence length of $M. \times piperita$	-	349	540	810	403			
GC ratio in L. dentata	60.1	65.9	43.6	-	28.6			
GC ratio in M. longifolia		66.6	44.1	-	-			
GC ratio in M . \times $piperita$		67.9	43.7	34.8	31.2			
Number of the retrieved Lavendula species from the GenBank		2	4	-	2			
Number of the retrieved <i>Mentha</i> species from the GenBank	4	9	9	11	14			

Identification of Mentha longifolia

Sequences of ITS, ITS2 and *rbc*L were used to identify *M. longifolia*. ITS2 recorded the lowest sequence length, whereas, the variable sites (%) and GC ratio of it were greater than those of ITS and *rbc*L (Table 2). Transitions were found to be more than transversions leading to substitution changes in the sequences of *M. longifolia* (Table 3). An evolution within *M. longifolia* genome was noticed through the high transition/transversion bias (R) that ranged from 3.51 in *rbc*L to 1.81 in ITS2. Except data of *rbc*L, Tajima relative evolutionary rate displayed an accelerated rates of evolution (*P*-values <0.05) in *M. longifolia* (Table 4). Sequences of *M. longifolia* for ITS, ITS2 and *rbc*L that submitted to BLAST at the GenBank retrieved 4, 9 and 9 *Mentha* species, respectively (Table 2). *M. longifolia* and the retrieved *Mentha* species reconstructed three phylogenetic trees (Fig. 2) which revealed that *M. longifolia* was represented in separate clade demonstrating variability between it and other *Mentha* species, and proved its

endemism to Saudi Arabia. Similar result was obtained by Khan et al. (2013) in Senecio asirensis using nrDNA ITS.

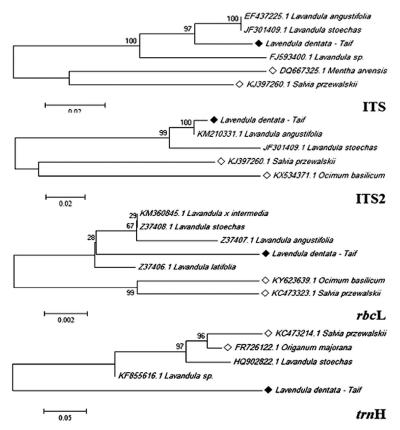


Fig. 1. Phylogenetic trees of Lavendula dentata and the retrieved species based on four loci. (\Diamond) refers to the outgroup.

Identification of M. \times piperita

Sequences of ITS2, rbcL, matK and trnH were used to identify M. × piperita. As found in M. longifolia, ITS2 showed the lowest sequence length (349 bp) and the highest GC ratio (67.9). Whereas, the percentage of variable sites (31%) was detected in trnH locus (Table 2). An obvious evolution was also observed within M. × piperita genome through the high transition/transversion bias (R) that ranged from 0.79 in matK to 7.01 in rbcL (Table 3). The previous result was supported by Tajima relative evolutionary rate that displayed an accelerated rates of evolution (P-values <0.05) in M. × piperita except that of rbcL (Table 4). The null hypothesis of equal evolution rates between M. × piperita from Taif and its ancestors; M. spicata and M. aquatica from one hand, and the retrieved M. × piperita from the other hand, was rejected because the P-values were lower than 0.05 in ITS2, matK and trnH revealing the accelerated evolutionary rate of M. × piperita from Taif and subsequently reflecting the variance among them. M. × piperita and the retrieved Mentha species from the GenBank library were analyzed to form four phylogenetic trees (Fig. 2). The phylogenetic trees from the ITS2, matK and trnH data demonstrated that all the

Table 3. Mean nucleotide substitution rates and transition/transversion bias (R) in loci for L. dentata, M. longifolia and M. × piperita.

Taxa	Locus		Tran	Transition					Transversion	ersion				(R)
		A→G	G→A	T→C	$C{\rightarrow}T$	A→T	T→A	A→C	C→A	T→G	G→T	C→G	G→C	
L. dentata	ITS	7.75	4.59	30.31	17.18	3.73	3.63	6.58	3.63	6.13	3.73	6.13	6.58	1.40
	ITS2	14.68	5.81	29.66	15.59	3.30	2.14	6.28	2.14	29.66	3.30	5.40	6.28	1.79
	rbcL	7.19	8.78	24.48	33.65	3.80	3.51	2.76	3.51	2.87	3.80	2.87	2.76	2.82
	trnH	6.07	23.09	7.57	19.07	6.44	8.32	2.56	8.32	3.27	6.44	3.27	2.56	1.19
M. longifolia	ITS	11.20	11.64	21.66	10.70	3.74	5.65	7.57	5.65	5.44	3.74	5.44	7.57	1.15
	ITS2	17.14	8.07	30.52	12.37	2.40	2.44	5.92	2.44	5.19	2.40	5.19	5.92	1.81
	rbcL	15.21	17.97	2.51	25.75	3.14	2.97	2.34	2.97	2.51	3.14	2.51	2.34	3.51
M. x piperita	matK	8.27	15.50	7.48	15.62	9.34	8.32	4.47	8.32	4.44	9.34	4.44	4.47	0.79
	trnH	8.44	23.85	7.16	28.37	6.45	5.92	1.63	5.92	2.10	6.45	2.10	1.63	1.48
	ITS2	17.25	8.10	28.84	11.59	2.56	2.62	6.36	2.62	5.57	2.56	5.57	6.36	1.63
	rbcL	17.09	20.19	21.49	28.93	1.77	1.67	1.31	1.67	1.41	1.77	1.41	1.31	7.01

Mentha species formed monophyletic clusters except the hybrid M. \times *piperita* from Taif which formed separate clades. The differences between M. \times *piperita* under study and the other retrieved *Mentha* species could be explained due to an evolutionary process.

Little divergence in rbcL tree (Fig. 2) and the acceptance of the null hypothesis of equal evolutionary rates among Mentha species through rbcL data (Table 4) could be due to the symmetry in rbcL sequence of Mentha species. Kshirsagar et~al. (2015) reported the same limitation of rbcL gene in closely related species of the two genera Ardisia Sw. and Swertia~L. These results were in accordance with those of Theodoridis et~al. (2012) who showed that matK and trnH were more useful in discriminating Lamiaceae species than rbcL. It was noticed that ITS2, rbcL, matK and trnH distinguished $M. \times piperita$ from the two parents, M. aquatica and M. spicata through the phylogentic trees. These genetic differences might be due to most commercial

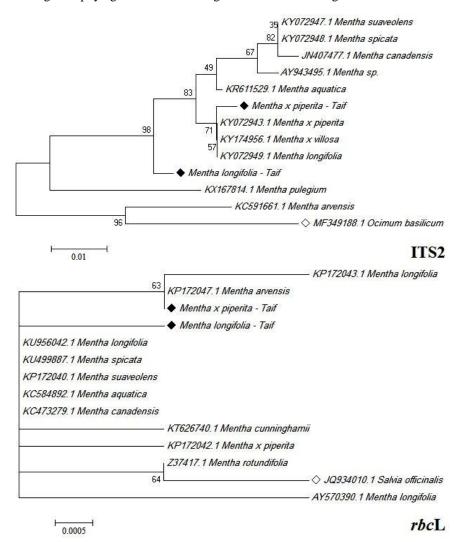


Fig. 2. Phylogenetic tree diverges between *Mentha longifolia* and M. \times *piperita* based on ITS2 and rbcL sequences. (\Diamond) refers to the outgroup.

hybrids, i.e. *M.* × *piperita* is sterile or subfertile, therefore, crossing with parental or nonparental species is expected. They may also form complex hybrid populations through vegetative propagation and polyploidy. These possibilities lead to great genetic diversity and subsequently to several taxonomic problems (De Mattia *et al.*, 2011).

Table 4. Tajima relative rate tests of loci for L. dentata, M. longifolia and M. \times piperita.

Loci	Outgroup	Testin	g group	RI	RD	RA	RB	χ2	P value
		(A)	(B)						
ITS	L. angustifolia	L. dentata-Taif	L. stoechas	512	0	36	0	36.0	< 0.05
ITS2	L. angustifolia	L. dentata-Taif	L. stoechas	280	0	2	17	11.8	< 0.05
rbcL	L. angustifolia	L. dentata-Taif	L. stoechas	529	0	5	0	5.00	< 0.05
trnH	L. angustifolia	L. dentata-Taif	L. stoechas	126	6	60	16	25.5	< 0.05
ITS	M. suaveolens	M. longifolia-Taif	M. spicata	70	0	33	0	33.0	< 0.05
	M. spicata	M. longifolia-Taif	M. suaveolens	70	0	33	1	30.12	< 0.05
ITS2	$M. \times piperita$	M. longifolia-Taif	M. longifolia	297	0	9	0	9.00	< 0.05
	M. longifolia	M. longifolia-Taif	$M. \times piperita$	297	0	9	0	9.00	< 0.05
rbcL	$M. \times piperita$	M. longifolia-Taif	M. longifolia	524	0	1	0	1.00	>0.05
	M. longifolia	M. longifolia-Taif	$M. \times piperita$	524	0	1	1	0.00	>0.05
ITS2	M. spicata	$M. \times piperita$ -Taif	M. aquatica	272	0	7	0	7.00	< 0.05
	M. aquatica	$M. \times piperita$ -Taif	M. spicata	272	0	7	0	7.00	< 0.05
	M. spicata	$M. \times piperita$ -Taif	$M. \times piperita$	296	0	4	0	4.00	< 0.05
rbcL	M. spicata	$M. \times piperita$ -Taif	M. aquatica	525	0	1	0	1.00	>0.05
	M. aquatica	$M. \times piperita$ -Taif	M. spicata	525	0	1	0	1.00	>0.05
	M. spicata	$M. \times piperita$ -Taif	$M. \times piperita$	524	0	1	1	0.00	>0.05
matK	M. spicata	$M. \times piperita$ -Taif	M. aquatica	411	1	263	0	263	< 0.05
	M. aquatica	$M. \times piperita$ -Taif	M. spicata	411	1	263	0	263	< 0.05
	M. spicata	$M. \times piperita$ -Taif	$M. \times piperita$	400	1	250	0	250	< 0.05
trnH	M. spicata	$M. \times piperita$ -Taif	M. aquatica	177	0	114	2	108.1	< 0.05
	M. aquatica	$M. \times piperita$ -Taif	M. spicata	177	0	114	2	108.1	< 0.05
	M. spicata	$M. \times piperita$ -Taif	M. × piperita	178	1	114	1	111.0	< 0.05
ITS2	M. spicata	M. longifolia-Taif	$M. \times piperita$ -Taif	320	0	3	6	1.0	>0.05
	M. spicata	$M. \times piperita$ -Taif	M. longifolia-Taif	320	0	3	6	1.0	>0.05
rbcL	M. spicata	M. longifolia-Taif	$M. \times piperita$ -Taif	524	0	1	1	0.00	>0.05
	M. spicata	$M. \times piperita$ -Taif	M. longifolia-Taif	524	0	1	1	0.00	>0.05

The Tajima relative rate test was used to examine the equality of evolutionary rate for *L. dentata*, *M. longifolia* and $M. \times piperita$ and other relative species with different outgorups.

Discrimination between M. longifolia and $M \times piperita$

Sequences of ITS2 and rbcL were used to discriminate between M. longifolia and $M \times piperita$. Statistics in Table 1 revealed slight differences between them. Results of mean nucleotide substitution rates, transition/transversion bias (R) and Tajima relative evolutionary rate were similar in these two taxa (Tables 3 & 4). ITS2 and rbcL trees were used to assess genetic divergences between M. longifolia and $M \times piperita$. A suitable divergence was detected in the

RI is the identical sites in all three sequences

RD is the divergent sites in all three sequences

RA is the number of unique differences in the sequence A

RB is the number of unique differences in the sequence B

 $[\]chi$ 2 test statistic more than 3.841 (P<0.05) indicates accelerated evolution

P value greater than 0.05 is often used to accept the null hypothesis of equal rates between lineages

two phylogentic trees displaying the efficacy of the two barcodes in distinguishing between them through the Maximum Likelihood method (Fig. 2). Thakur *et al.* (2016) stated that the convenient barcode exhibits large interspecific but little intraspecific divergence and its sequence length must be short enough to be available in a single amplification. This comparability of interspecific sequence variation is a significant aspect for barcoding identification of species in local floras. Establishing a local barcode data will be useful in several ecological applications, such as the reconstruction of community phylogenies, palaeoecological studies of ecosystems and analyzing the diets of human and other animals (Valentini *et al.*, 2009). Depending on these data, the DNA barcoding could be considered as a good approach for distinguishing and identifying the mint plants, however, it was not possible to confirm the relationship between hybrids and their putative parents.

Finally, it could be concluded that the identification and discrimination of L. dentata, M. longifolia and $M \times piperita$ were necessary and valuable for their great economic importance. ITS, matK and trnH were found to be more effective barcodes than ITS2 and rbcL for the authentication of these species and hybrid. DNA barcoding provided new insight that will contribute to the taxonomy of Lamiaceae taxa around the world and the conservation of the genetic resources of these valuable taxa occurring in Saudi Arabia.

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References

- Abdel Khalik, K.N. 2016. A Systematic revision of the genus *Plectranthus* L. (Lamiaceae) in Saudi Arabia based on morphological, palynological and micromorphological characters of trichomes. Am. J. Plant Sci. 7: 1429–1444.
- Anwar, F., Alkharfy, K.M., Rehman, N., Adam, E.H.K. and Gilani, A. 2017. Chemo-geographical variations in the composition of volatiles and the biological attributes of *Mentha longifolia* L. essential oils from Saudi Arabia. Int. J. Pharmacol. **13**: 408–424.
- Burgess, K.S., Fazekas, A.J., Kesanakurti, P.R., Graham, S.W., Husband, B.C., Newmaster, S.G., Percy, D.M., Hajibabaei, M. and Barrett, S.C.H. 2011. Discriminating plant species in a local temperate flora using the *rbcL+matK* DNA barcode. Methods Ecol. Evol. **2**: 333–340.
- Collenette, S. 1999. Wild Flowers of Saudi Arabia. National Commission for Wildlife Conservation and Development (NCWCD), Riyadh.
- De Mattia, F., Bruni, I., Galimberti, A., Cattaneo, F., Casiraghi, M. and Labra, M. 2011. A comparative study of different DNA barcoding markers for the identification of some members of Lamiaceae. Food Res. Int. 44: 693–702.
- Doyle, J.J. and Doyle, J.L. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem. Bull. 19: 11–15.
- Edgar, R.C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Res. **32**(5): 1792–1997.
- Hajibabaei, M., Singer, G.A.C., Hebert, P.D.N. and Hickey, D.A. 2007. DNA barcoding: how it complements taxonomy, molecular phylogenetics and population genetics. Trends Genet. **23**: 167–172.
- Kasem, W.T. 2016. Pollen grains and seed morphology as related to biochemical patterns in five species of genus *Ocimum* L. (Lamiaceae Juss.) of Saudi Arabia. J. Phytol. 8: 42–48.
- Khan, S., Al-Qurainy, F., Nadeem, M. and Tarroum, M. 2013. Selection of chloroplast DNA markers for the development of DNA barcode and reconstruction of phylogeny of *Senecio asirensis* Boulos and J.R.I. wood. Pak. J. Bot. **45**(2): 703–710.

- Kshirsagar, P., Umdale, S., Chavan, J. and Gaikwad, N. 2015. Molecular authentication of medicinal plant, *Swertia chirayita* and its adulterant species. Proc. Natl. Acad. Sci. India Sect. B, Biol. **87**(1): 1–7.
- Miller, A.G. 1985. The genus *Lavendula* in Arabia and Tropical NE Africa. Notes Roy. Bot. Gard. Edinburgh **42**(3): 503–528.
- Mogosan, C., Vostinaru, O., Oprean, R., Heghes, C., Filip, L., Balica, G. and Moldovan, R. 2017. A comparative analysis of the chemical composition, anti-inflammatory, and antinociceptive effects of the essential oils from three species of *Mentha* cultivated in Romania. Molecules **22**: 263–274.
- Tajima, F. 1993. Simple methods for testing molecular clock hypothesis. Genetics 135: 599-607.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. 2013. MEGA6: Molecular evolutionary genetics analysis version 6.0. Mol. Biol. Evol. 30: 2725–2729.
- Thakur, V., Tiwari, S., Tripathi, N., Tiwari, G. and Sapre, S. 2016. DNA barcoding and phylogenetic analyses of *Mentha* species using *rbc*L sequences. Ann. Phytomed. **5**(1): 59–62.
- Theodoridis, S., Stefanaki, A., Tezcan, M., Aki, C., Kokkini, S. and Vlachonasios, K.E. 2012. DNA barcoding in native plants of the Labiatae (Lamiaceae) family from Chios Island (Greece) and the adjacent Cesme-Karaburun Peninsula (Turkey). Mol. Ecol. Resour. 12: 620–633.
- Valentini, A., Miquel, C., Nawaz, M., Bellemain, E., Coissac, E., Pompanon, F., Gielly, L., Cruaud, C., Nascetti, G., Wincker, P., Swenson, J. and Taberlet, P. 2009. New perspectives in diet analyses based on DNA barcoding and parallel pyrosequencing: the *trnL* approach. Mol. Ecol. Resour. 9: 51–60.

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