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Antidiarrheal activity of methanolic leaf extract of *Rumex vesicarius*

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Abstract

This study evaluates the antidiarrheal activity of *Rumex vesicarius* (leaf) by using *in vitro* and *in vivo* assays. Antidiarrheal effect of *R. vesicarius* was evaluated using castor oil-induced diarrhea model in rat. Weight and volume of the intestinal content were assessed using the enteropooling method. Atropine (3 mg/kg, i.p) was used as positive control. *R. Vesicarius* at the doses of 200 and 400 mg/kg p.o. significantly retarded castor oil-induced enteropooling and intestinal transit. The gastrointestinal transit rate was studied and *R. vesicarius* at the doses of 200 and 400 mg/kg significantly inhibited ($p < 0.001$) weight and volume of intestinal content. *R. vesicarius* caused concentration-dependent (0.01–1 mg/mL) relaxation of spontaneous contractions in isolated rabbit jejunum tissue preparation and inhibited K^+ -80 induced contractions (0.01–5 mg/mL), similar to verapamil, suggestive of calcium channel blockade. Results obtained herein indicate that *R. vesicarius* may contain effective compounds which can be used as an antidiarrheal agent.

Introduction

Diarrhea affects 70% of the world population (Ouyang and Chen, 2004), especially in the third world countries where sanitary conditions are alarming. Acute diarrhea being the most common, is usually caused by an infectious agent, even though drugs, poisons or acute inflammatory reactions can contributing factors (Thapar and Sanderson, 2004). Rotavirus is the major causative agent of infectious diarrhea, particularly in young children now-a-days, however, other viral (*Enterovirus*, *norovirus* and *adenovirus*), bacterial (*Salmonella* sp., *Shigella* sp., *Escherichia coli*, *Camphylobacter* and *Vibrio cholerae*) and parasitic (*Cryptosporidium* and *Giardia*) agents are important pathogens (Allen et al., 2004).

Rational use of herbal medicines is accepted universally; here we attempt to investigate the folklore claim of *Rumex vesicarius* L. leaf extracts for antidiarrheal activity. *R. vesicarius* has been in use cooling agent, astringent, anti-venom agent and appetizer for the treatment of allaying pain of toothache, nausea, and

insect bite, seeds were used for dysentery (Dymoke, 1972).

In the Ayurvedic system of medication, it was used as stomachic (Ahirrao et al., 2011), antitumor, analgesic, flatulence, spleen disease, high cough, asthma, laxative, bronchitis, dyspepsia, heart troubles, alcoholism and biliousness (Kirtikar and Basu, 1987). In the Unani system of medication, it was used as a tonic, in leucoderma, scabies and as diuretic (Kirtikar and Basu, 1987). In other folk medicines, it was used to eradicate piles, constipation and hiccup (Hariparasad, 2011). Reptile insect, urinary affection, hepatoprotective, dysmenorrhea, blood purifier, depurative, sedative, alkalinity, chronic catarrh, renal disorders, dyspepsia, bloody dysentery and coronary (Madhavashetty et al., 2008), vomiting (Khan et al., 2013), leucoderma, antiviral, lymphatic glandular system disease, antidiabetic, rectal prolapsus, aphrodisiac anticholesterol, impetigo and carbuncles (Pullaiah and Ali, 1999), antioxidant (Rao, 2003), anthelmintic (Rao et al., 2012), cancer and inflammation (Aggarwal et al., 2006), spasmogenic and



spasmolytic (Khan et al., 2014a), diuretic (Rao et al., 2011), antifungal (Amira et al., 2011), and antipyretic (Khan et al., 2014b). This study reports the antidiarrheal activity of methanolic leaf extracts of *R. vesicarius*.

Materials and Methods

Plant material

R. vesicarius was collected in December, 2012 from the sandy fields of Mondka Shahjamal District, Muzaffar Garh, Pakistan. The plant material was authenticated by expert taxonomist, Prof. A. H. Dasti at the Department of Botany, Bahauddin Zakariya University, Multan, Pakistan (voucher F.P.ST-215). The plant material was made free from foreign adulterants and vegetative debris by hand picking and leaves were detached from the plant, washed and shade dried. Within 8 days, the leaves became crispy. The special electrical herbal grinder was used to form the coarse powder. Uniform dark green powder was obtained with a characteristic smell.

Crude extract

The powdered plant material (1 kg) was subjected to maceration in 70% methanol in an amber colored glass bottle at room temperature (25°C) for 8 days with occasional shaking (Aziz et al., 2013a). The soaked material was passed through muslin cloth to remove the vegetative material and the fluid obtained was filtered through Whatman No. 1 Filter paper. The filtrate was evaporated on a rotary evaporator (Rotavapor, BUCHI labrotechnik AG, Model 9230, Switzerland) at 37°C under reduced pressure. The approximate yield was 11% and the extract obtained was stored at -4°C in airtight jars in lab refrigerator.

Preliminary phytochemical screening

Major phytochemical classes were screened by the method described by Aziz et al. (2013b).

Chemicals and drugs

All the chemicals, solvents, and drugs used were of analytical grade. Atropine was purchased from Ethical Laboratories (Pvt.) Ltd. Pakistan. Verapamil was purchased from Aventis Pharmaceuticals (Pvt.) Ltd. Pakistan. All other chemicals used were of the available analytical grade.

Animals and housing condition

Five adults locally breed rabbits (1.0-1.5 kg) of either sex, purchased from the animal market Hussain Agahi Multan, Pakistan, with age limit between 6 to 7 months were used for the experiments. Fifteen rats (200-240 g) were purchased from the animal house Islamia University, Bahawalpur, Pakistan. Animals were provided with fresh green fodder and tap water *ad libitum* and maintained in an air-conditioned room (23-25°C) at the

Faculty of Pharmacy, Bahauddin Zakariya University, Multan. All rabbits were kept in fasting condition for at least 12 hours before the commencement of experiments, but had free access to water. The experiments were approved by the Ethical Committee of the Bahauddin Zakariya University, Multan, (EC/12/2012 dated 07 December 2012).

Castor oil-induced diarrhea

Rats were divided into 4 groups of 3 animals each, diarrhea was induced by administering 1 mL of castor oil orally to rats. The Group I treated as control (2 ml/kg, i.p. saline); Group II received atropine (3 mg/kg, i.p.) served as standard and Group III and IV received *R. vesicarius* (200 and 400 mg/kg, i.p.) 1 hour before castor oil administration. The number of both wet and dry diarrheal droppings was counted every hour for a period of 4 hours mean of the stools passed by the treated groups were compared with that of the positive control groups consisted of animals given an intraperitoneal injection of saline and atropine (Aziz et al., 2014).

Castor oil-induced enteropooling

Intraluminal fluid accumulation was determined by the method of Robert et al. (1976). Overnight fasted rats were divided 4 groups of 3 animals each. The Group I received normal saline (2 mL/kg, i.p.) served as a control; Group II received atropine (3 mg/kg, i.p.) and Groups III and IV received *R. vesicarius* 200 and 400 mg/kg orally respectively. 1 hour before the oral administration of castor oil. Two hours later the rats were sacrificed, the small intestine was removed after tying the ends with thread and weighed. The intestinal contents were collected by milking into a graduated tube and their volume was measured. The intestine was reweighed and the difference between full and empty intestines was calculated.

Small intestinal transit

Rats fasted for 18 hours were divided into 5 groups of 3 animals each. Group I received 2 mL normal saline orally; Group II received 2 mL of castor oil orally with saline 2 mL/kg intraperitoneally; Group III received atropine (3 mg/kg, i.p.); Group IV and V received *R. vesicarius* 200 and 400 mg/kg intraperitoneally respectively, 1 hour before administration of castor oil. One mL of the marker (10% charcoal suspension in 5% gum acacia) was administered orally 1 hour after castor oil treatment. The rats were sacrificed after 1 hour and the distance traveled by the charcoal meal from the pylorus was measured and expressed as the percentage of the total length of the intestine from the pylorus to cecum (Izzo et al., 1999).

Isolated rabbit jejunum preparations

The rabbit was starved overnight and was sacrificed subsequently to a blow on the head. The abdomen was opened and jejunum was dissected out and cut into

segments of about 2 cm in length following removal of adhering mesenteries. The segments were mounted between two stainless steel hooks in a 10 mL tissue bath, containing the normal Tyrode solution (pH 7.4), maintained at 37°C and aerated with carbogen (5% CO₂ + 95% O₂). A preload of 1 g was applied and the tissue was allowed to equilibrate for a period of 30 min during which the tissue was washed with fresh fluid at an interval of every 10 min prior to exposure to any test material. The spontaneous contractions were recorded isototically through a Power Lab Data Acquisition System (AD Instruments, Sydney, Australia) (Khan et al., 2014a).

Statistical analysis

The data expressed are the mean ± standard error of the mean (SEM) and the median effective concentrations (EC₅₀ values) with 95% confidence intervals (CI). The statistical parameter applied in the castor oil-induced diarrhea test is chi-square test. $p < 0.05$ was noted as significantly different. Concentration-response curves were analyzed by nonlinear regression using Graph Pad program version 6 for Windows (Graph Pad, USA).

Results

Phytochemical analysis of *R. vesicarius* methanolic extract shown the presence of alkaloids, glycosides, saponins, tannins, anthraquinones, coumarins and phenols.

Castor oil-induced diarrhea, 30 min after administration of castor oil, the diarrhea was clinically apparent in all the animals of the control group, for the next 4 hours. This was markedly reduced by the intraperitoneal injection of atropine, 3 mg/kg (74.3%). A similar marked reduction in the number of defecations after 4 hours was achieved with *R. vesicarius* the doses of 200 or 400 mg/kg i.p. *R. vesicarius* 200 and 400 mg/kg significantly inhibited the defecation (39.2% and 66.3%). The dose of extract delayed the onset of diarrhea and only 30% of animals showed diarrhea at first hour ($p < 0.001$) (Table I). While, in castor oil induced interpooling both the doses 200 and 400 mg/kg significantly inhibited the weight of intestinal content, 26.3 and 49.1 respectively (Table II). Furthermore, on castor oil-induced small intestinal transit both the doses of *R. vesicarius* shown 29.9 and

Table I

Castor oil-induced diarrhea in rats

Group	Treatment	Number of defecation (within 4 hours)	%Inhibition of defecation
I	Castor oil (1 mL p.o) + saline (2 mL/kg i.p)	27.2 ± 2.1	---
II	Castor oil (1 mL p.o) + atropine (3 mg/kg i.p)	6.9 ± 0.2 ^b	74.3
III	Castor oil (1 mL p.o) + <i>R. vesicarius</i> (200 mg/kg i.p)	16.5 ± 0.9 ^a	39.2
IV	Castor oil (1 mL p.o) + <i>R. vesicarius</i> (400 mg/kg i.p)	9.1 ± 0.7 ^b	66.3

Effect of *R. vesicarius* on castor oil-induced diarrhea in rats: *R. vesicarius* was administered i.p 1 hour before castor oil administration. Values are expressed as mean ± SEM from the experiments; ^a $p < 0.01$, ^b $p < 0.001$ when compared with castor oil + saline-treated group

Table II

Effect of *R. vesicarius* on castor oil-induced enteropooling in rats

Group	Treatment	Weight of intestinal content (g)	%Inhibition of weight intestinal content
I	Castor oil (1 mL p.o) + saline (2 mL/kg i.p)	2.4 ± 0.1	---
II	Castor oil (1 mL p.o) + atropine (3 mg/kg i.p)	1.8 ± 0.1 ^b	24.4
III	Castor oil (1 mL p.o) + <i>R. vesicarius</i> (200 mg/kg i.p)	1.7 ± 0.1 ^a	26.1
IV	Castor oil (1 mL p.o) + <i>R. vesicarius</i> (400 mg/kg i.p)	1.2 ± 0.1 ^b	49.3

Effect of *R. vesicarius* on castor oil-induced enteropooling in rats: *R. vesicarius* was administered i.p 1 hour before castor oil administration. Values are expressed as mean ± SEM from the experiments; ^a $p < 0.01$, ^b $p < 0.001$ when compared with castor oil + saline-treated group

Table III				
Effect of <i>R. vesicarius</i> on castor oil-induced small intestinal transit in rats				
Group	Treatment	Total length of intestine (cm)	Distance travelled by marker (cm)	%Intestinal transit inhibition
I	Saline (2 mL/kg i.p)	82.8 ± 1.6	45.5 ± 1.7	44.9
II	Castor oil (1 mL p.o) + saline (2 mL/kg i.p)	80.2 ± 2.9	72.8 ± 1.2	9.2
III	Castor oil (1 mL p.o) + atropine (3 mg/kg i.p)	83.5 ± 2.8	35.8 ± 1.3 ^b	57.0
IV	Castor oil (1 mL p.o) + <i>R. vesicarius</i> (200 mg/kg i.p)	80.3 ± 1.8	56.2 ± 1.7 ^a	29.9
V	Castor oil (1 mL p.o) + <i>R. Vesicarius</i> (400 mg/kg i.p)	82.0 ± 1.2	45.1 ± 1.3 ^b	45.0

Effect of *R. vesicarius* on castor oil-induced small intestinal transit in rats: *R. vesicarius* was administered i.p 1 hour before castor oil administration. Values are expressed as mean ± SEM from the experiments; ^ap<0.01, ^bp<0.001 when compared with castor oil + saline-treated group

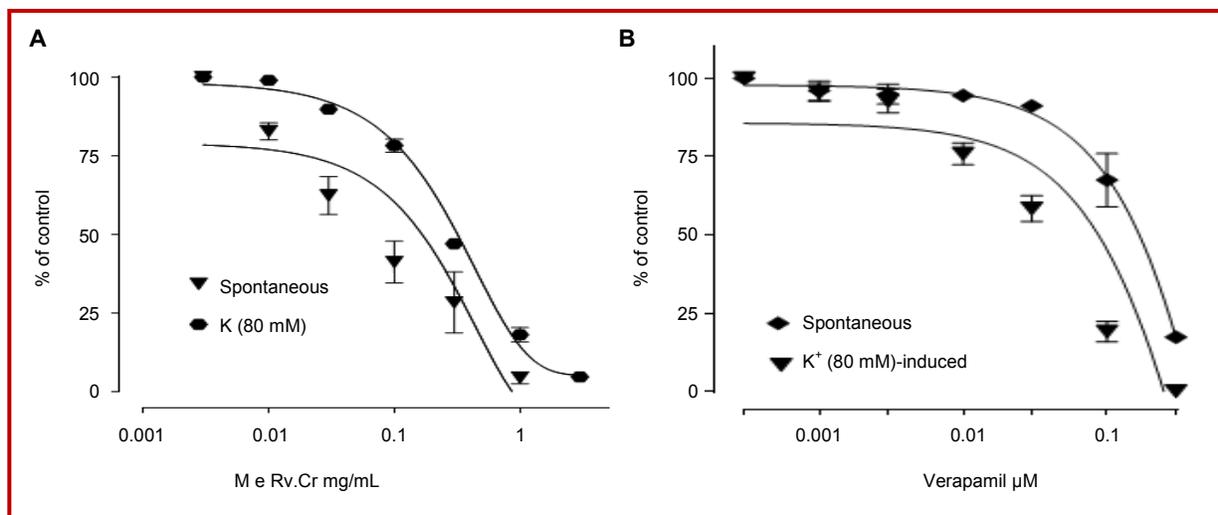


Figure 1: Concentration response curves showing the inhibitory effect of *R. vesicarius* (A) and verapamil (B) on spontaneous and K⁺ (80 mM)-induced contractions in isolated rabbit jejunum preparations. The values shown are mean ± SEM of 5 observations

45% inhibition (Table III). *R. vesicarius* caused concentration-dependent (0.01–1 mg/mL) relaxation of spontaneous contractions in isolated rabbit jejunum tissue preparation and inhibited K⁺-80 induced contractions (0.01–5 mg/mL), similar to verapamil, suggestive of calcium channel blockade (Figure 1). The calcium channel blockade effect was confirmed when pretreatment of the jejunum preparation with *R. vesicarius* produced a concentration-dependent (0.03–1 mg/mL) rightward shift in Ca⁺⁺ concentration response curves, as caused by verapamil (Figure 2).

Discussion

R. vesicarius exhibited a dose-dependent protective effect against diarrhea. It is also noted that *R. vesicarius* significantly inhibited castor oil-induced intestinal fluid accumulation and the volume of intestinal content, dose

dependently more than atropine. *R. vesicarius* significantly reduced the castor oil induced intestinal transit. In this study, atropine produced a significant reduction in the number of stools and increased intestinal transit time possible due to its anti-cholinergic effect (Crocì et al., 1997). However, it did not inhibit castor oil induced enteropooling and gain in weight of intestinal content, suggesting thereby that mediators other than acetylcholine are involved in castor oil induced enteropooling. An increase in intestinal transit time with atropine could also result due to the reduction in gastric emptying (Pierce et al., 1997). Castor oil is also reported to induce diarrhea by increasing the volume of intestinal content by prevention of the reabsorption of water. The liberation of ricinoleic acid results in irritation and inflammation of the intestinal mucosa, leading to release of prostaglandins, which results in stimulation of secretion (Iwao and Terada, 1962), thereby prevents the reabsorption of NaCl and water

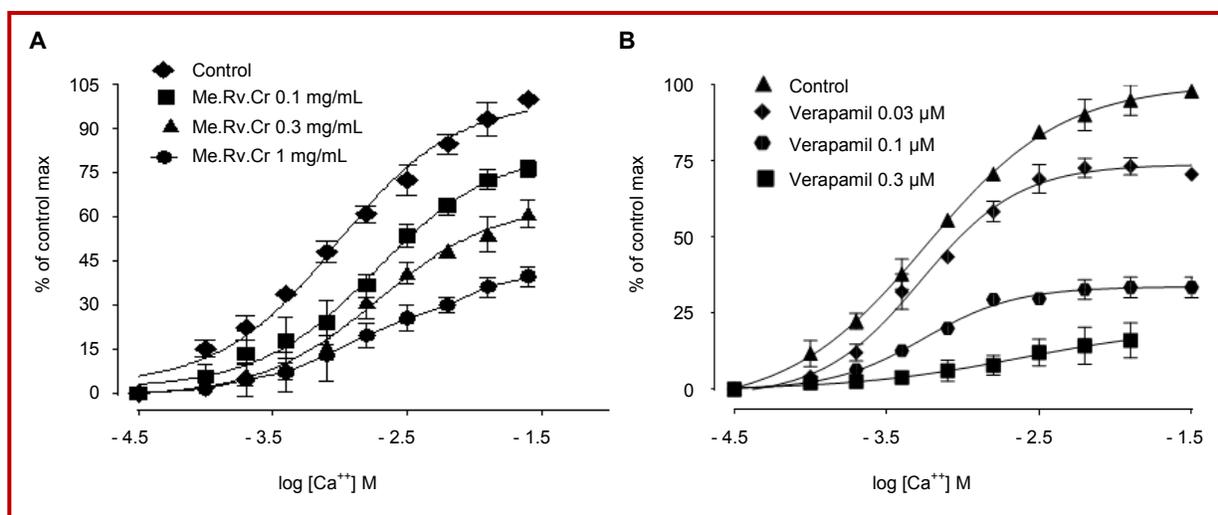


Figure 2: Effect of the *R. vesicarius* on concentration-response curve of Ca²⁺ on isolated rabbit jejunum preparations (A), and effect of the verapamil (standard) on concentration-response curve of Ca²⁺ on isolated rabbit jejunum preparations (B); Values are expressed as mean \pm S.E.M.; n=5

(Mascolo et al., 1994). Probably *R. vesicarius* increased the reabsorption of NaCl and water by decreasing intestinal motility as observed by the decrease in intestinal transit by charcoal meal.

The secretory diarrhea is associated with an activation of Cl⁻ channels, causing Cl⁻ efflux from the cell, the efflux of Cl⁻ results in massive secretion of water into the intestinal lumen and profuse watery diarrhea (Galvez, 1993). *R. vesicarius* may inhibit the secretion of water into the lumen by altering this mechanism. Antidiarrheal property of medicinal plants is found to be due to tannins, alkaloids, saponins, flavonoids, sterols and/or triterpenes and reducing sugars (Ammon et al., 1985). The phytochemistry of *R. vesicarius* revealed the presence of alkaloids, triterpenoids, tannins, flavonoids, phenols, gums, carbohydrates and mucilage (Hariparasad, 2011). These phytoconstituents may mediate the antidiarrheal property of the *R. vesicarius*. Although the antidiarrheal properties of the reported active terpenoids are well established. Sesquiterpenes, diterpenes, flavonoids, terpenes, and terpenoid derivatives are known for inhibiting release of autocoids and prostaglandins, thereby inhibit the motility and secretion induced by castor oil (Vimala et al., 1997; Longanga et al., 2000). *R. Vesicarius* is reported effective in many antimicrobial studies against the bacteria and fungus causing infectious diarrhea (Mostafa and ElBakry, 2011; Sahli and Abdulkhair, 2011).

Conclusion

The presence of antidiarrheal activity *in vivo* and *in vitro* assays in *R. vesicarius* mediated possibly through a calcium antagonistic mechanism, which might explain the traditional use of the plant in abdominal cramps, diarrhea and dysentery.

References

- Aggarwal BB, Ichikawa H, Garodia P, Weerasinghe P, Sethi G, Bhatt ID, Pandey MK, Shishodia S, Nair MG. From traditional Ayurvedic medicine to modern medicine: Identification of therapeutic targets for suppression of inflammation and cancer. *Expert Opin Ther Targets*. 2006; 10: 88-118.
- Ahirrao YA, Patil DA. Ethnomedicinal claims against stomach complaints in Buldhana District (Maharashtra, India). *Life Sci Leaflet*. 2011; 1: 16-25.
- Allen SJ, Okoko B, Martinez E, Gregorio G, Dans LF. Probiotics for treating infectious diarrhea. *Cochrane Database Syst Rev*. 2004; 2: CD003048.
- Amira M, Abu T, Kadriya E, Fatimah OA. Assessment of antifungal activity of *Rumex vesicarius* L. and *Ziziphus spinachristi* (L.) Willd. Extracts against two phytopathogenic fungi. *Afr J Microbiol Res*. 2012; 5: 1001-11.
- Ammon HV, Soergel KH. Diarrhea in Bockus Gastroenterology. 4th ed. Philadelphia, Saunders, 1985, pp 125-41.
- Aziz A, Khan IA, Munawar SH, Munzoor Z, Agha S. Evaluation of antitussive activity of *Lycopus europaeus* on cough reflex induced by different cough induced models in mice. *Int J Pharma Sci*. 2014; 3: 412-16.
- Aziz A, Khan IA, Munawar SH, Sadrul, S. Antipyretic study of methanolic bark extract of *Plumeria ubra*, Linn. in various pyrexia induced models. *Int J Res Dev Pharm Life Sci*. 2013a; 2: 680-85.
- Aziz A, Khan IA, Ahmad BA. Evaluation of the antidiarrheal activity of aqueous extract of *Lycopus europaeus* in mice. *Pharma Sci Monitor*. 2013b; 4: 442-49.
- Croci T, Landi M, Emonds-Alt X, Le-Fur G., Maffrand JP, Manara L. Role of tachykinins in castor oil diarrhoea in rats. *Bri J Pharmacol*. 1997; 121: 375-80.
- Dymoke W. A history of the principal drugs of the vegetable origin. 2nd ed. Pharmacographia Indica. Karachi, Hamdard

- Publications, 1972, p 2114.
- Kalvez J, Zavzuelo A, Crespo ME, Lorente MD, Ocete MA, Jimenez J. Anti-diarrhoeic activity of *Euphorbia hirta* extract and isolation of an active flavanoid constituent. *Planta Medica*. 1993; 59: 333-36.
- Hariparasad PS. Phytochemical screening and pharmacognostic evaluation of *Rumex vesicarius* L. *Int J Pharmtech Res*. 2011; 3: 1078-82.
- Izzo AA, Mascolo N, Capasso R, Germano MP, DePasquel R, Capasso F. Inhibitory effect of cannabinoid agonists on gastric emptying in the rat. *Arch Pharmacol*. 1999; 360: 221-23.
- Iwao I, Terada Y. On the mechanism of diarrhea due to castor oil. *Japanese J Pharmacol*. 1962; 12: 137-45.
- Khan IA, Aziz A, Munawar SH, Munzoor Z. Antiemetic activity of methanolic leaf extract of *Rumex vesicarius* Linn. *Int J Pharm Res Allied Sci*. 2013; 2: 33-37.
- Khan IA, Aziz A, Saqib F, Munawar SH, Manzoor Z, Raza MS. Pharmacological evaluation of *Rumex vesicarius* Linn leaf extract and fractions in rabbit gastrointestinal ailment. *Afri J Pharm Pharmacol*. 2014a; 8: 333-41.
- Khan IA, Aziz A, Munawar SM, Manzoor Z, Sarwar HS, Afzal A, Raza MA. Study on antipyretic activity of *Rumex vesicarius* leaves extract in albino rabbits. *Vet World*. 2014b; 3: 41-45.
- Kirtikar KR, Basu BD. Indian medicinal plants. Vol. 2. Dehradun, India, International Book Distributors, 1987.
- Longanga QA, Verduyze A, Foriers A. Contribution to the ethnobotanical, phytochemical and pharmacological studies of traditionally used medicinal plants in the treatment of dysentery and diarrhea in Lomela area, Democratic Republic of Congo (DRC). *J Ethnopharmacol*. 2000; 71: 411-23.
- Madhavashetty K, Shivaji K, Tulasirao K. Flowering plants of Chittor District, Andhra Pradesh, India. 2nd edi. Students Offset Printer, 2008, p 298.
- Mascolo N, Izzo AA, Avtore G, Barbato F, Capasso F. Nitric oxide and castor oil induced diarrhea. *J Pharmacol Exper Ther*. 1994; 268: 291-95.
- Mostafa AM, ElBakry EA. Evaluation of antibacterial and antioxidant activities of different plant parts of *Rumex vesicarius* L. (polygonaceae). *Int J Pharm Pharmaceut Sci*. 2011; 3: 109-18.
- Ouyang H, Chen JZ. Therapeutic roles of acupuncture in functional gastrointestinal disorders. *Aliment Pharmacol Therapeut*. 2004; 20: 831-41.
- Pierce NF, Carpenter CJ, Elliot HZ, Greenough WB. Effects of prostaglandins, theophylline and cholera exotoxin upon transmucosal water and electrolyte movement in canine jejunum. *Gastroenterology* 1977; 60: 22-32.
- Pullaiah T, Ali MD. Flora of Andhra Pradesh (India). Vol. 2. Scientific Publishers, 1999, p 817.
- Rao R. Bioactive phytochemicals in Indian foods and their potential in health promotion and disease prevention. *Asia Pac J Clin Nut*. 2003; 12: 9-22.
- Rao KN, Sunitha Ch, Sandhya S, Rajeshwar T. Anthelmintic activity of different extracts on aerial parts of *Rumex vesicarius* Linn. *Int J Pharm Sci Rev Res*. 2012; 12: 64-66.
- Rao KN, Sunitha Ch, David B, Sandhya S, Shwetha D, Murali K. Diuretic activity on different extracts and formulation on aerial parts of *Rumex vesicarius* Linn. *J Chem Pharm Res*. 2011; 3: 400-08.
- Robert A, Nezamis JE, Lancaster C, Hanchar AJ, Klepper MS. Enteropooling assay: A test for diarrhea produced by prostaglandins. *Prostaglandins* 1976; 11: 809-28.
- Sahli AA, Abdulkhair WM. Inhibition of beta-lactamase enzyme of *Pseudomonas aeruginosa* by clavulanic acid of *Rumex vesicarius* L. *Afr J Agric Res*. 2011; 6: 2908-15.
- Thapar N, Sanderson IR. Diarrhea in children: An interface between developing and developed countries. *Lancet* 2004; 363: 641-53.
- Vimala R, Nagarajan S, Alam M, Susan T, Joy S. Anti-inflammatory and antipyretic activity of *Michelia champaca* Linn., (white variety), *Ixora brachiata* Roxb. and *Rhynchosia cana* (Willd.) D.C. flower extract. *Indian J Exp Biol*. 1997; 35: 1310-14.

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