

Original article:

The Protective Effect of Azelnidipine for the Prevention of Heart Fibrosis Occurrence on Balb/c Mice with Iron Overload

Hadi Sarosa¹, Udin Bahrudin², Ag Soemantri³, Siti Fatimah-Muis⁴, Nur Arfian⁵, Ichiro Hisatome⁶

Abstract:

Background: Iron overload can cause DNA oxidation which increase TGF β 1, type 1 fibrillarprotein and myocardium fibrosis. Myocardium fibrosis is the main cause of death on the state of iron overload. The iron influx towards the cell during iron overload is still unknown, some research suggested LTCC acts as iron influx. This research aims to investigate the role of azelnidipine as type L calcium channel blocker, lowering TGF β 1, collagen and myocardium fibrosis. **Method:** The research subjects consisted of 25 male Balb-C mice(8 weeks, 30-40mg) divided into 5 groups. Group 1 (NaCl+S) 0,3 cc Na Cl 0,9% (I.P) and drug solvent (Aquabidest, CMC and Nipagin) orally. Group 2 Fe+S) 0.3 cc 1,5 mg Fe+sucrose (Venofer®) (I.P) and drug solvent (Aquabidest, CMC and Nipagin) orally. Group 3 (Fe+Dfx) 1,5 mg Fe+sucrose (Venofer®) (I.P) and deferasirox 20 mg/kg body weight/day orally, group 4 (Fe+Azl) 1,5 mg Fe+sucrose (Venofer®) (I.P) and azelnidipine 14 mg/day orally and group 5 (Fe+Dfx-Azl) 1,5 mg Fe+sucrose (Venofer®) (I.P) and mixture of deferasirox 20 mg/kg body weight/day and azelnidipine 14 mg/day orally. Fe-sucrose was diluted with NaCl 0.9 %. Intraperitoneal injection were administered intermittently for 60 days of treatment. **Result:** The highest Expression of TGF β , collagen I and fibrosis area fractions are in group Fe+S. The result of Post Hoc test between 2 treatment groups indicated that there were no difference in TGF β expression between groups NaCl+S with Fe+Dfx (P>0.05) , Fe+Dzl (P>0.05). There are no significant in collagen expression between groups NaCl+S with Fe+Dfx (P > 0.05) ,Fe+Dzl (P>0.05). **Conclusion:** Azelnidipine, LTCC have roles on the influx of iron into the myocardium, lowering TGF β , collagen I expressions and myocardium fibrosis.

Keywords: Azelnidipine; iron overload; expression of collagen; myocardium fibrosis

Bangladesh Journal of Medical Science Vol. 19 No. 02 April'20. Page : 223-228
DOI: <https://doi.org/10.3329/bjms.v19i2.44999>

Introduction

Iron plasma overload is still a problem for people with thalassemia. Iron plasma overload occurred due to haemolysis, repeated transfusion and also due to

the iron metabolism dysregulation. Administration of blood transfusion in β thalassemia will improve the accumulation of iron substance , which is 0.34 mg/kg body weight /day on patients with the need for 600 Red Packed Cell / 4 weeks ¹. Iron plasma overload

1. Hadi Sarosa, Diponegoro University Postgraduate Program of Medical Sciences, Semarang, Indonesia & Departement of Physiology, Medical Faculty of Sultan Agung Islamic University Semarang, Indonesia
2. Udin Bahrudin, Department of Cardiology and Vascular Medicine, Diponegoro University Faculty of Medicine, Semarang, Indonesia
3. Ag Soemantri, Department of Pediatric, Diponegoro University Faculty of Medicine, Semarang, Indonesia
4. Siti Fatimah-Muis, Department of Nutrition, Diponegoro University Faculty of Medicine, Semarang, Indonesia
5. Nur Arfian, Departement of Anatomy, Faculty of Medicine, Public Health, and Nursing Universitas Gadjah Mada Yogyakarta, Indonesia
6. Ichiro Hisatome, Division of Regenerative Medicine, Tottori University Graduate School of Medical Science, Yonago, Japan.

Correspondence to: Hadi Sarosa, Diponegoro University Postgraduate Program of Medical Sciences, Semarang, Indonesia & Departement of Physiology, Medical Faculty of Sultan Agung Islamic University Semarang, Indonesia. Postal address Medical Faculty of UNIVERSITAS ISLAM SULTAN AGUNG ,Jalan Raya Kaligawe KM4, Semarang 50112, Central Java Indonesia. Email idahasoras@yahoo.com

cause the accumulation of iron in the internal organs. The accumulation of iron in cardiovascular muscle is cardiovascular complication and the main cause of death on repeated transfusion²

On the physiological state, transferrin receptor 1 (TfR1) plays an important role in the process of iron influx to the intra cell. Transferrin receptor 1 (TfR1) which located in the cardiac cell wall is important in the influx of iron to the myocardium, however during the presence of high intra cell iron level, TfR1 expression will be suppressed³⁻⁵. On the state of high iron plasma, several studies suggested the type L calcium channel (L-type calcium channels/LTCC)^{2,6-11} while others stated that type T calcium channel (*T-type calcium channels/TTCC*)^{12,13} are playing important roles in the influx of iron in the myocardium.

The accumulation of iron will cause cell damage due to the DNA oxidation. DNA oxidation triggers the apoptosis process and increase growth factor (TGF- β 1, PDGF), cytokine and extracellular matrices¹⁴. Cascade transforming growth factor- β 1 (TGF- β 1) plays a major role in inducing fibroblast into myofibroblast and increasing MMP2 and MMP 9 expressions which cause fibrosis^{14,15}. Cardiomyopathy and heart failure/ cardiac arrest due to the myocardium fibrosis are the leading cause of death for iron plasma overload^{2,9}.

Management of iron accumulation to this day is not yet satisfactory, one of them is iron chelation. Deferasirox is the latest development of iron chelation, it has the ability to bind plasma iron and penetrate cell membranes to bind intra cell iron¹⁶. Deferasirox is a tridentate molecule, forming 2 electrochemical bonds of Fe(2:1) ions which are quite stable, given orally once a day¹⁷. The use of iron chelating drugs to eliminate iron accumulation in the heart is still not yet satisfactory

In a state of high plasma iron, several studies have shown L-type calcium channels (*L-type calcium channels/LTCC*)¹¹ while others suggested that T-type calcium channel (*T-type calcium channels/TTCC*)¹³ plays an important role in the influx of iron into the myocardium. The type L calcium channel density is affected by estrogenic receptors, an increase in estrogen will reduce the number of type L calcium channels and reduce the entry of Ca²⁺ in the myocardium¹⁸. Azelnidipine is along acting third generation partition L type calcium channel, very soluble in fat¹⁹. Azelnidipine has the hydroxyl

radical scavenger effect to reduce ROS, antioxidant and antifibrotic effects²⁰.

This research is going to look at the effect of azelnidipine on TGF β , collagen expression and iron overload model heart fibrosis on male experimental animals. Iron overload model is by administration of intraperitoneal iron sucrose in Balb/c mice intermittently.

Materials and Methods

Animal subjects

Experiments were conducted after approval by the ethical Committee of the Faculty of Medicine, Diponegoro University, no 16/EC/H/FK-RSDK/V/2017. Male Balb/c mice (n = 25) age 8 weeks old, with 30-40 gr body weight were obtained from the Experimental Animal Care Unit (UPHP) LPPT of Gadjah Mada University. Mice were caged maintained by the Department of Biology, Faculty of Medicine Unissula, with a light-dark cycle of 12:12 hour, humidity : 50-60 %, temp : 24° -26°. Research subjects were randomly selected into 5 groups. Group 1 (NaCl+S) were given 0,3 cc Na Cl 0.9% intra peritoneal and drug solvent (Aquabidest, CMC dan Nipagin) 0.5 cc by turns orally every day, group 2 (Fe+S) were given 1,5 mg (0.3 cc) Fe+ sucrose (Venofer®) and drug solvent 0.5 cc orally, group 3 (Fe+Dfx) were given 1,5 mg (0.3 cc) Fe+ sucrose (Venofer®) and deferasirox 20 mg/kg body weight/day orally, group 4 (Fe+Azl) were given 1,5 mg (0.3 cc) Fe+ sucrose (Venofer®) and azelnidipine 14 mg/day orally and group 5 (Fe+Dfx-Azl) were given 1,5 mg (0.3 cc) Fe+ sucrose (Venofer®) and mixture of deferasirox 20 mg/kg body weight/day and azelnidipine 14 mg/day orally, Fe-sucrose was diluted with NaCl 0.9 %, intraperitoneal injection and was done intermittent day. Mice got standard feeding and free access to water *ad libitum*. Mice were terminated at day 70 after operation

Heart harvesting

Before termination, the mice were anesthetized with pentobarbital (60 mg/kg ip), then the abdomen and thorax were opened. Heart were harvested and half ventricle was in RNA later ® for extraction RNA and other half was fixated in PFA 4% in PBS for 24 hour and paraffin was used embedded tissue process.

Histological analysis

Paraffin section with 4 mm thickness was analyzed. Paraffin section were deparaffinized and stained with Sirius Red to quantify the fibrosis interstitial fraction

area. Quantification of fibrosis area was done using Image J software, with 10 fields in each sample and with 400 x magnification.

Reverse Transcriptase PCR

RNA was extracted from heart tissue using FavorPrep™Tri-RNA Reagent (Favorgen, FATRR 001, Biotech Corp). cDNA was synthesized using ReverTra-Ace (TOYOBO Co, Ltd, TRT-101x10). Reverse transcriptase PCR (RT-PCR) was done for examining the expression of following gen : collagen 1 (forward, 5'-ATGCCGCGACCTCAAGATG-3'; reverse, 5'-GAGGCACAGACGGCTGAGTA-3'), TGF β1 (forward, 5'-TTCCGCTGCTACTGCAAGTCA-3'; reverse 5'-GGGTAGCGATCGAGTGTCCA-3'), Caspase 3 (forward 5'YCYGACTGGAAAGCCGAAACTC-3'; reverse, 5' TCCCACTGTCTGTCTCAATGCCAC-3'), GAPDH (forward, 5'-TTGCTGTTGAAGTCGCAGGAG-3'; reverse, 5'-TGTGTCCGTCGTGGATCTGA-3') was used reference.

Statistical Analysis

Data were presented as mean ± SD collagen and TGF β level and fraction area fibrosis were analysed with ANOVA test and continued with Post Hoc test

Ethical clearance: This research study was approved by ethics committee of Medical Faculty of Sultan Agung Islamic University Semarang, Indonesia.

Results

On group Fe+S the highest average of TGF β 1 and collagen I expression. The lowest average of TGF β expression was on group Fe+Dfx and collagen expression the lowest expression was on group Fe+Azl

Table1 : TGF β1expression on various treatment groups on Balb-c mice

Mice	NaCl+S	Fe+S	Fe+Dfx	Fe+Azl	Fe+Dfx-Azl
1	1.56	1.52	1.30	1.37	1.36
2	1.37	1.46	1.43	1.43	1.40
3	1.39	1.45	1.26	1.24	1.43
4	1.50	1.86	1.25	1.48	1.58
5	1.52	1.75	1.36	1.45	1.67
Mean	1.47±0.08	1.61±1.86	1.32±0.06	1.40±0.1	1.49±0.13

Table 2 : Collagen I expression on various

treatment groups on Balb-c mice

Mice	NaCl+S	Fe+S	Fe+Dfx	Fe+Azl	Fe+Dfx-Azl
1	1.20	1.32	1.13	1.25	1.22
2	1.21	1.24	1.12	1.11	1.19
3	1.16	1.21	1.07	0.96	1.16
4	1.23	1.56	1.04	1.16	1.35
5	1.30	1.50	1.17	1.20	1.39
Mean	1.22±0.05	1.37±0.16	1.11±0.05	1.14±0.11	1.26±0.10

The administration of Fe indicated that there was increasing expression of TGF β 1and collagen I expression which will cause damage on heart muscle and cause fibrosis. The results of ANOVA test, there is significant difference on TGF β 1expression (p < 0.05) and collagen I expression (p < 0.01). The result of Post Hoc test between 2 treatment groups indicated that there were no difference in TGF β expression between groups NaCl+S with Fe+Dfx (P>0.05), Fe+Dzl (P>0.05). There are no significant in collagen expression between groups NaCl+SwithFe+Dfx (P > 0.05) ,Fe+Dzl (P>0.05).

Discussion

Myocardium is composed from several types of cells, cardiomyocyte, cardiofibroblast and endothelial cell. Cardiofibroblast generate cytokines, and growth factors and have the function to preserve the structural and functional integrities of the extracellular matrices. The extracellular matrix has a very important role as the mechanical and chemical properties between cardiomyocytes, cardiofibroblast cells and blood vessels ¹⁴. The myocardial extracellular matrix is woven between cells arranged by proteins, fibrous hidden in materials similar to gels composed of complex carbohydrates. A dilute gel is called interstitial fluid, which is the space between blood vessels and tissue cells which are part of nutrients, residual results, and materials that dissolve in the air ²¹. The extracellular matrix composes three components of macromolecules namely glycosaminoglycan, fibrous protein collagen and glycoprotein ²².Inflammation²³, oxidative stress²⁴ and hypoosmolar hyperhydration²⁵are contributors to cardiomyocyte impairment. Damage to the myocardium increases several growth factors (TGF β1, PDGF), cytokines (TNF α, IL 1β, IL6), extracellular matrix and causes cardiofibroblast differentiation to myocardiofibrobast. Myocardiofibroblasts have contraction ability, can move and produce more extracellular matrix proteins ¹⁴. Cardiofibroblast hyperactivity and myocardiofibroblasts increase the production of collagen I, II fibilar proteins and

extracellular matrix deposits¹⁵. Cardiofibroblast accumulation and accumulation of collagen fibrillary proteins that are maternity, supported by tissue and myocardial function^{14,26,27}.

This study indicated that that intraperitoneal administration of Fe 1,5 mg intraperitoneal by intermittent days can increase the TGF β 1, Collagen I expression and myocardium fibrosis.

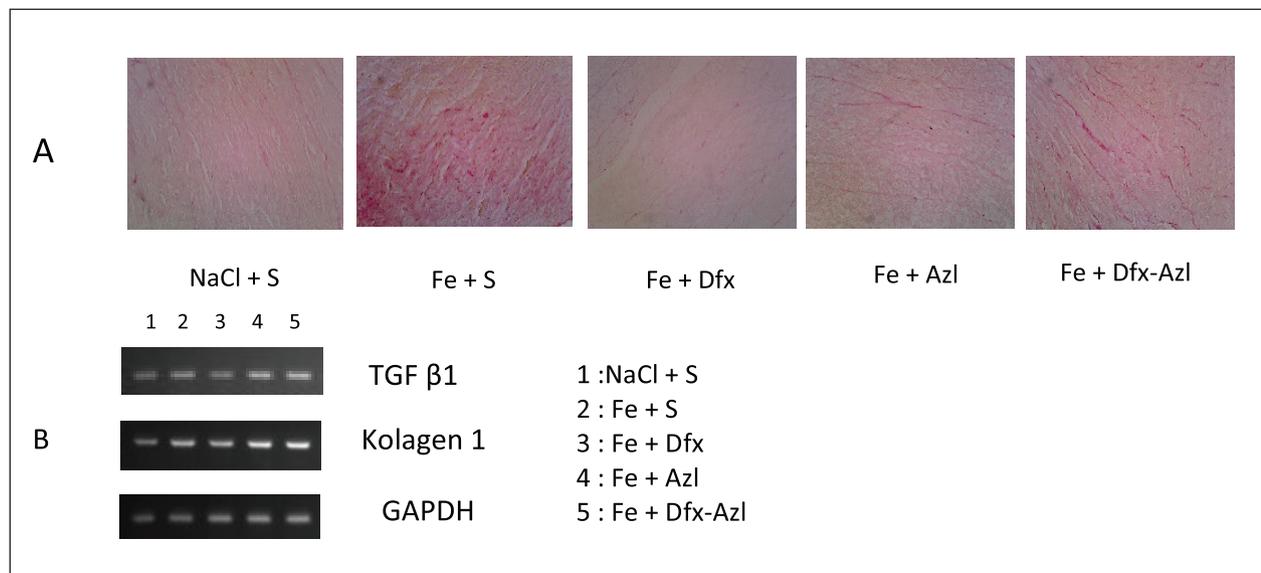


Figure 1 : (A)Histochemistry examination with Sirius Red (SR) staining indicated that group Fe+S with the most fibro- sis area fractions, no significant difference of area fractions (%) NaCl + S with Fe + Azl

(B) reverse transcriptase PCR (RT-PCR) TGF β 1, Collagen I and GAPDH, and densitometry analysis with Image J software indicated that there is no difference ($P>0.05$) Group Fe+Azl vs NaCl+S, Fe+Azl vs Fe+Dfxdan Fe+Azl vs Fe+Dfx-Azl. There is significant difference ($P<0.05$) Group Fe+Azl Vs Fe+S.

Myocardial iron deposits result in oxidative stress due to an increase in the hydroxyl radical prooxide. Increased hydroxyl radicals increase peroxidase and damage to lipids, proteins and DNA²⁸. DNA damage is measured by expression of growth factor (TGF- β 1). Cascade transforming growth factor- β 1 (TGF- β 1) induces fibroblasts into myofibroblasts and increases the expression of collagen fibrillar proteins, which results in fibrosis^{14,15}.

L- type calcium channels (LTCC) are mostly in the myocardium, play an important role during calcium entry as triggers of heart muscle contraction, regulation of the duration of action potential and regulation of gene expression²⁹. The relationship between chronic heart failure and serum calcium³⁰. In the state of iron overload, LTCC plays a role in the influx of iron into the heart muscle^{2,6-11}. The lowest of collagen expression average were in the Fe-Azl group, Azelnidipine is type L Ca Channel

blocker which is an antihypertensive drug with better effect of anti-inflammatory and anti-oxidant than amlodipine, furthermore, it has the antifibrotic effect by inhibiting TGF- β 1 in the liver²⁰, and inhibits apoptosis by reducing cytochrome C levels in HL-1 cardiomyocytes³¹

Azelnidipine is an anti-hypertension that can prevent heart damage. Azelnidipine has better anti inflammatory and antioxidant effects than amlodipine³²antifibrotic effects by inhibiting TGF- β 1 in the liver²⁰, and inhibiting apoptosis by reducing cytochrome C levels in HL-1 cardiomyocytes³¹. Cardiofibroblast hyperactivity and myocardial fibroblasts increase the production of collagen I, II fibrillar proteins and extracellular matrix deposits¹⁵. Fibrillar collagen type I and II are the most common components of the myocardial extracellular matrices¹⁴. TGF β 1 pathway is an important pathway for collagen production³³and where fibrosis takes place¹⁵.

L-type calcium channels (LTCC) play a role in the entry of Fe²⁺^{2,6-11}.This study shows that administration of azelnidipine has a protective effect on the occurrence of myocardial fibrosis, lowering TGF β 1 and collagen I expression . L Type Calciumchannel (LTCC) plays a role in the entry of iron into the cell.

Acknowledgement

Source of Funding

This study was supported by Doctoral Dissertation grants from Ministry of Research Technology and Higher Education of Republic Indonesia

We would like to appreciate to those who are participating in this study. This work was performed in partial fulfillment of the requirements for Doctoral of Faculty of Medicine Diponegoro University

Conflict of Interest:

The authors declared that they have no conflict of interest.

Contribution of Authors:

Data gathering and idea owner of this study: HadiSarosa

Study design: HadiSarosa

Data gathering: HadiSarosa, Udin Bahrudin, Nur Arfian

Writing and submission of manuscript: Hadi Sarosa, Ag Soemantri, Siti Fatimah-Muis, Udin Bahrudin, Nur Arfian, Putri R Ayuningtyas

Editing and approval of final draft: HadiSarosa, Ag Soemantri, Siti Fatimah Muis, Udin Bahrudin, Ichiro Hisatome

References:

1. Marsella, M. & Borgna-Pignatti, C. Transfusional iron overload and iron chelation therapy in thalassemia major and sickle cell disease. *Hematol. Oncol. Clin. North Am.* **28**, 703–27, vi (2014).
2. Murphy, C. J. & Oudit, G. Y. Iron-overload cardiomyopathy: pathophysiology, diagnosis, and treatment. *J. Card. Fail.* **16**, 888–900 (2010).
3. Ke, Y. *et al.* Post-transcriptional expression of DMT1 in the heart of rat. *J. Cell. Physiol.* **196**, 124–130 (2003).
4. Parkes, J. G., Liu, Y., Sirna, J. B. & Templeton, D. M. Changes in gene expression with iron loading and chelation in cardiac myocytes and non-myocytic fibroblasts. *J. Mol. Cell. Cardiol.* **32**, 233–246 (2000).
5. Srail, S. K. Protein of Iron Homeostasis. in *Iron Physiology and Pathophysiology in Humans* (eds. Anderson, G. J. & McLaren, G. D.) 3–25 (Humana Press, 2012). doi:10.1007/978-1-60327-485-2
6. Parkes, J. G., Olivieri, N. F. & Templeton, D. M. Characterization of Fe²⁺ and Fe³⁺ transport by iron-loaded cardiac myocytes. *Toxicology* **117**, 141–151 (1997).
7. Oudit, G. Y. *et al.* L-type Ca²⁺ channels provide a major pathway for iron entry into cardiomyocytes in iron-overload cardiomyopathy. *Nat. Med.* **9**, 1187–1194 (2003).
8. Oudit, G. Y., Trivieri, M. G., Khaper, N., Liu, P. P. & Backx, P. H. Role of L-type Ca²⁺ channels in iron transport and iron-overload cardiomyopathy. *J. Mol. Med. (Berl)* **84**, 349–64 (2006).

9. Gujja, P., Rosing, D. R., Tripodi, D. J. & Shizukuda, Y. Iron overload cardiomyopathy: better understanding of an increasing disorder. *J. Am. Coll. Cardiol.***56**, 1001–12 (2010).
10. Chen, M., Cabantchik, Z. I., Chan, S., Chan, G. C. & Cheung, Y. Iron Overload and Apoptosis of HL-1 Cardiomyocytes : Effects of Calcium Channel Blockade. *PLoS One***9**, 1–9 (2014).
11. Abd Allah, E. S. H., Ahmed, M. a. & Abdel Mola, A. F. Comparative study of the effect of verapamil and vitamin D on iron overload-induced oxidative stress and cardiac structural changes in adult male rats. *Pathophysiology***21**, 293–300 (2014).
12. Kumfu, S. *et al.* T-type calcium channel as a portal of iron uptake into cardiomyocytes of beta-thalassemic mice. *Eur. J. Haematol.***86**, 156–166 (2011).
13. Kumfu, S., Chattipakorn, S., Chinda, K., Fucharoen, S. & Chattipakorn, N. T-type calcium channel blockade improves survival and cardiovascular function in thalassemic mice. *Eur. J. Haematol.***88**, 535–48 (2012).
14. Fan, D., Takawale, A., Lee, J. & Kassiri, Z. Cardiac fibroblasts, fibrosis and extracellular matrix remodeling in heart disease. *Fibrogenesis Tissue Repair***5**, 15 (2012).
15. Longo, D. L., Rockey, D. C., Bell, P. D. & Hill, J. a. Fibrosis — A Common Pathway to Organ Injury and Failure. *N. Engl. J. Med.***372**, 1138–1149 (2015).
16. Wang, Y. *et al.* Iron-induced cardiac damage: role of apoptosis and deferasirox intervention. *J. Pharmacol. Exp. Ther.***336**, 56–63 (2011).
17. Gammella, E., Recalcati, S., Rybinska, I., Buratti, P. & Cairo, G. Iron-Induced Damage in Cardiomyopathy : Oxidative-Dependent and Independent Mechanisms. *Oxid. Med. Cell. Longev.***2015**, (2015).
18. Philp, K. L. *et al.* Greater antiarrhythmic activity of acute 17beta-estradiol in female than male anaesthetized rats: correlation with Ca²⁺ channel blockade. *Br. J. Pharmacol.***149**, 233–242 (2006).
19. Inomata, J. I. *et al.* Differential effects of azelnidipine and amlodipine on sympathetic nerve activity in patients with primary hypertension. *J. Hypertens.***32**, 1898–1904 (2014).
20. Ohyama, T. *et al.* Azelnidipine is a calcium blocker that attenuates liver fibrosis and may increase antioxidant defence. *Br. J. Pharmacol.***165**, 1173–1187 (2012).
21. Sherwood, L. Sistem Reproduksi. in *Fisiologi Manusia Dari Sel ke Sistem (Introduction to Human Physiology)* 781–839 (EGC, 2013).
22. Alberts, B. *et al.* Cell Junctions and The Extracellular Matrix. in *Molecular Biology of The Cell* 1035–1090 (Garland Science, 2015).
23. Kawai, C. From Myocarditis to Cardiomyopathy : Mechanisms of Host Factors That Influence Susceptibility to. *Heart***99**, 1091–1100 (1999).
24. Chen, A. F. *et al.* Free radical biology of the cardiovascular system. *Clin. Sci. (Lond)*.**123**, 73–91 (2012).
25. Olga, Y. *et al.* Age-dependent cardioprotective action of meldonium on heart remodeling under the experimental hyposmolar hyperhydration. *Bangladesh J. Med. Sci.***18**, 390–6 (2019).
26. Krenning, G., Zeisberg, E. M. & Kalluri, R. The origin of fibroblast and mechanism of cardiac fibrosis. *J. cell Physiol.***225**, 631–637 (2010).
27. Van Linthout, S., Miteva, K. & Tschöpe, C. Crosstalk between fibroblasts and inflammatory cells. *Cardiovasc. Res.***102**, 258–269 (2014).
28. Kaur, K., Sharma, A. K., Dhingra, S. & Singal, P. K. Interplay of TNF-alpha and IL-10 in regulating oxidative stress in isolated adult cardiac myocytes. *J. Mol. Cell. Cardiol.***41**, 1023–1030 (2006).
29. Shaw, R. M. & Colecraft, H. M. L-type calcium channel targeting and local signalling in cardiac myocytes. *Cardiovasc. Res.***98**, 177–186 (2013).
30. Khrystyma, P. *et al.* Investigation of calcium metabolism in patients with coronary heart disease complicated by chronic heart failure, stage II-A. *Bangladesh J. Med. Sci.***17**, 395–401 (2018).
31. Bahrudin, U. *et al.* Simultaneous treatment with azelnidipine and olmesartan inhibits apoptosis of hl-1 cardiac myocytes expressing e334k CMYBPC. *Drug Res. (Stuttg)*.**63**, 515–520 (2013).
32. Komoda, H., Inoue, T. & Node, K. Anti-Inflammatory Properties of Azelnidipine, a Dihydropyridine-Based Calcium Channel Blocker. *Clin. Exp. Hypertens.***32**, 121–128 (2010).
33. Porter, K. E. & Turner, N. a. Cardiac fibroblasts: At the heart of myocardial remodeling. *Pharmacol. Ther.***123**, 255–278 (2009).