

Editorial:

Mobile Colistin Resistance (MCR) Gene, Antimicrobial Resistance: A Global Public Health Threat

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Colistin (Polymyxin E) is a novel antibiotic which gains importance in recent years though it was discovered a long time ago in 1947 from a bacteria *Paenibacillus polymyxa* var. *colistinus*.¹⁻⁴ Colistin possess high nephrotoxic and neurotoxic adverse effects; thereafter, it was abandoned in the 1970s and 80s as there was the development of less toxic highly effective newer antimicrobials.⁵⁻⁷ As the years passed, emergence and widespread distribution of multidrug resistance gram-negative bacteria such as carbapenem-resistant *Enterobacteriaceae* (CRE), carbapenem-resistant *Acinetobacter baumannii* (CRAB), multidrug resistance *Pseudomonas* species, extensively drug-resistant (XDR) *Pseudomonas aeruginosa*, and XDR *Acinetobacter baumannii* were posing a serious threat to human health and safety.⁸⁻⁹ This paved the way for a reconsideration of colistin as a valuable therapeutic alternative as a last resort used against infections caused by these potentially dangerous gram-negative pathogens.^{10, 11} "Colistin is an amphiphilic, complex, multi-component, pentacationic decapeptide antibiotic."¹² Colistin binds electrostatically to the outer membrane of gram-negative bacteria, increasing the permeability of membrane which leads to extravasation of cytoplasmic content, cell lysis and ensues death of

pathogens.^{13, 14} It is only effective against gram-negative microorganisms, thereafter, designated as a narrow spectrum bactericidal agent. Two forms of colistin are commercially available, colistin methanesulfonate sodium (CMS) used parenterally and colistin sulfate (CS) given oral and topically as powder for skin infections.^{1, 15} Nevertheless, alarming news is extensive use of colistin to treat the sturdy bacterial infections especially in ICU setting has led to development of gram-negative colistin-resistant microorganism; such as colistin-resistant *Enterobacteriaceae*, colistin-resistant *Acinetobacter baumannii* (CoRAB), *Pseudomonas* etc.¹⁶⁻¹⁹ Colistin resistance was thought due to chromosomal mutations caused by modifications of Lipopolysaccharide (LPS) at outer cell wall of gram-negative pathogens. Additionally, affinity of colistin towards LPS reduced through amendment of negative charge.^{14, 20} These chromosomal mutations among gram-negative organisms towards colistin only vertically transmissible.^{21, 22} So it was believed that colistin was one of the last drugs where resistance was not spread from cell to cell.^{21, 23, 24} A plasmid-borne transmissible mobile colistin-resistant gene (mcr-1) was identified among gram-negative pathogens *Escherichia coli* SHP45 in Chinese pork meat in 2015.²⁵ The colistin-

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resistant *E. coli* EC11 strain was identified from dairy farms by 2016 in same city in Shanghai, China.²⁶ Until now nine different mcr genes were identified.²⁷ In Belgium mcr-2 was first found in bovine and porcine *E. coli* and rarely found outside European union,²⁸ mcr-3,²⁹ mcr-7,³⁰ mcr-8³¹ in China, mcr-4 found in Italy, Spain and Belgium,³² mcr-5 in Germany,³³ mcr-6 (formally known as mcr-2.2) in UK.³⁴ mcr-9 was detected in multidrug-resistant *Salmonella enterica* serotype typhimurium identified isolated from a patient.^{27, 35} The presence mcr-1 gene has been evidenced throughout the planet.³⁶ Additionally, mcr-1 positive *E. coli* widespread presence in environment, food, and animals especially livestock and birds. Nevertheless, human carriers of mcr-1 so far been rare. It has been observed that less than 2% and ≤0.2% among *Enterobacteriaceae* and *E. coli* clinical specimens contain mcr-1 gene in China and Europe, respectively.^{25, 37-39} The low human carrier of mcr gene may be due to restricted use of colistin in hospitals than commercial animal and poultry industries.²¹ On the other hand, high levels of colistin are utilized in rooster and animal farms, especially to control colibacillosis originated diarrhea diseases,⁴⁰ that in-turn promotes a strong selective pressure for colistin resistance animals.²¹ So far, to date mcr-1 carrying isolates were recognized from 31 countries.³⁶ The nations with the highest numbers of mcr-1 positive tested specimens were in China, Vietnam, and Germany. *E. coli* is the main reservoir for mcr-1, but the other gram-negative pathogens were also documented as mcr-1 positive. Those pathogens were *Salmonella enterica*, *Klebsiella pneumoniae*, *Escherichia fergusonii*, *Kluyvera ascorbata*, *Citrobacter braakii*, *Cronobacter sakazakii*, and *Klebsiella aerogenes*.³⁶

It has been evidenced that the mcr-1 gene and other microbial resistance gene can overlap and exists in the same microbial class. The mcr-1 often presents with ESBLs (CTX-M, SHV, and TEM type) and AmpC cephalosporinase (CMY type), quinolones resistance genes (qnrS and aac (6')-Ib-cr).^{37, 38, 41, 42} Colistin sulfate to be the most commonly used antibiotic in the poultry industry in Bangladesh.⁴³ Colistin-resistant *E. coli* carrying mcr-1 has not been isolated from hospital settings, and this is the first report of the occurrence of colistin-resistant *E. coli* carrying resistant marker mcr-1 from the environment (city sludge specimens) of Bangladesh.⁴⁴ There are several

phenotypic and genotypic methods for the detection of mcr-1 producers.^{45, 46} Broth microdilution (BMD) is considered the reference test to see the susceptibility profile of polymyxin.¹ The other method to document the mcr-1 gene is Colistin MAC tet (CMT), Combined Disc test (CDT), Colistin MIC reduction (CMR), Modified Rapid Polymyxin NP test (RPNP) and alteration of Zeta potential tests.⁴⁶⁻⁴⁸ Polymerase Chain Reaction (PCR) and whole-genome Sequence (WGS) is considered the reference tests to identify the mcr gene from cultured bacteria as well as in clinical, fecal, environmental and food samples.^{47, 49} The advantage of PCR is to detect known mcr genes and the WGS can identify all known or unknown colistin resistance genes within 2 days.⁵⁰ Polymyxin / colistin is an essential therapeutic agent in medicine both for human and food-producing industries; thereafter, it is imperative need to arrest the speedy spread of mcr-1 concealing plasmid.^{1, 47, 51} Subsequently to halt such rapid spread of colistin gene utilization this last resort needs to be more rational and minimum in absolute clinical necessity. As irrational imprudent use of antibiotics acts as a promoter of resistance.^{52, 53} It is extremely significant to appraise the frequency of human carriers of *E. coli* mcr-1 +.³⁸ The spread of *E. coli* in the hospital environment by maintaining the proper sanitization protocol,⁵⁴ by screening the patients whether they work in food chain.⁵⁵ Antimicrobial resistance (AMR) is a global public health threat not only to human life but also for food industries to feed us. Consequently, it is essential to address it on multiple levels. Tourists, vacationers, explorers to those geographic parts with high levels of AMR should have vaccines up-to-date.⁵⁶ Resistance to colistin increases potentially problematic for clinicians to choose antimicrobials. As only a few antimicrobials are remaining for Colistin-, Carbapenem-Resistant *Klebsiella pneumonia* (C-C-RKp) infections.⁵⁷ The antimicrobial selection procedure should be on strict and appropriate scientific, rational, and prudent basis, such as hospital antibiogram of clinical specimen, the location of infection, pharmacokinetics/pharmacodynamics (PK/PD) effects and potential adverse drug reactions.⁵⁷⁻⁵⁹ Researchers suggest recently that the best available treatment options colistin-resistant infection is tigecycline, gentamicin, fosfomycin, and ceftazidime/avibactam.^{57, 60} As conclusive statement more research advocated using

the “one health” method to delay/stop antimicrobial resistance and safeguard human life.⁶¹ There is need to aware of the food industry operators along the entire food chain and to reduce colistin as growth promoters.^{55, 62} Wide-ranging research should be commenced immediately to discover more novel antimicrobial molecules to fight against the Colistin-Carbapenem resistant pathogens. National

and International medicine control agencies must develop stringent policies with regulatory policing activities to control misuse and irrational utilization antimicrobials; and need to develop regular educational intervention programs both for health professionals and consumers to promote prudent use of antibiotics.⁶³⁻⁶⁷

References:

1. Poirel L, Jayol A, Nordmann P. Polymyxins: Antibacterial Activity, Susceptibility Testing, and Resistance Mechanisms Encoded by Plasmids or Chromosomes. *Clin Microbiol Rev.* 2017;30(2):557–596. doi:10.1128/CMR.00064-16
2. Benedict RG, Langlykke AF. Antibiotic activity of *Bacillus polymyxa*. *J Bacteriol.* 1947; 54: 24.
3. Jeong H, Choi SK, Ryu CM, Park SH. Chronicle of a Soil Bacterium: *Paenibacillus polymyxa* E681 as a Tiny Guardian of Plant and Human Health. *Front Microbiol.* 2019; 10: 467. doi:10.3389/fmicb.2019.00467
4. Biswas S, Brunel JM, Dubus JC, Gaubert MR, Rolain JM. Colistin: an update on the antibiotic of the 21st century. *Expert Rev Anti Infect Ther.* 2012; 10 (12): 917-934. doi: 10.1586/eri.12.78.
5. Li J., Nation R.L., Turnidge J.D., Milne R.W., Coulthard K., Rayner C.R., Paterson D.L. Colistin: The re-emerging antibiotic for multidrug-resistant Gram-negative bacterial infections. *Lancet Infect. Dis.* 2006; 6 (9): 589–601. doi: 10.1016/S1473-3099(06)70580-1.
6. Zavascki A.P., Goldani L.Z., Li J., Nation R.L. Polymyxin B for the treatment of multidrug-resistant pathogens: A critical review. *J. Antimicrob. Chemother.* 2007; 60 (6): 1206–1215. doi: 10.1093/jac/dkm357.
7. Nation RL, Rigatto MHP, Falci DR, Zavascki AP. Polymyxin Acute Kidney Injury: Dosing and Other Strategies to Reduce Toxicity. *Antibiotics (Basel).* 2019; 8 (1): 24. doi:10.3390/antibiotics8010024
8. Eichenberger EM, Thaden JT. Epidemiology and Mechanisms of Resistance of Extensively Drug-Resistant Gram-Negative Bacteria. *Antibiotics (Basel).* 2019; 8 (2): 37. doi: 10.3390/antibiotics8020037
9. van Duin D, Paterson DL. Multidrug-Resistant Bacteria in the Community: Trends and Lessons Learned. *Infect Dis Clin North Am.* 2016; 30 (2): 377-390. doi: 10.1016/j.idc.2016.02.004
10. Lim LM, Ly N, Anderson D, Yang JC, Macander L, Jarkowski A 3rd, Forrest A, Bulitta JB, Tsuji BT. Resurgence of colistin: a review of resistance, toxicity, pharmacodynamics, and dosing. *Pharmacotherapy.* 2010; 30 (12): 1279-1291. doi:10.1592/phco.30.12.1279
11. Dhariwal AK, Tullu MS. Colistin: re-emergence of the ‘forgotten’ antimicrobial agent. *J Postgrad Med.* 2013; 59 (3): 208-215. doi: 10.4103/0022-3859.118040.
12. Wallace SJ, Li J, Nation RL, Prankerd RJ, Velkov T, Boyd BJ. Self-assembly behavior of colistin and its

- prodrug colistin methanesulfonate: implications for solution stability and solubilization. *J Phys Chem B.* 2010; 114 (14): 4836-4840. doi:10.1021/jp100458x
13. Mohamed YF, Abou-Shleib HM, Khalil AM, El-Guink NM, El-Nakeeb MA. Membrane permeabilization of colistin toward pan-drug resistant Gram-negative isolates. *Braz J Microbiol.* 2016; 47 (2): 381-388. doi: 10.1016/j.bjm.2016.01.007
 14. Trimble MJ, Mlynářík P, Kolář M, Hancock RE. Polymyxin: Alternative Mechanisms of Action and Resistance. *Cold Spring Harb Perspect Med.* 2016; 6 (10): a025288. doi: 10.1101/cshperspect.a025288
 15. Rhouma M, Beaudry F, Thériault W, Letellier A. Colistin in Pig Production: Chemistry, Mechanism of Antibacterial Action, Microbial Resistance Emergence, and One Health Perspectives. *Front Microbiol.* 2016; 7: 1789. doi:10.3389/fmicb.2016.01789
 16. Garcia Casallas JC, Robayo-Amortegui H, Corredor-Rozo Z, Carrasco-Márquez AM, Escobar-Perez J. Bacteremia by colistin-resistant *Acinetobacter baumannii* isolate: a case report. *J Med Case Rep.* 2019;13(1):141. doi:10.1186/s13256-019-2062-3
 17. Gelbíčová T, Baráková A, Florianová M, Karpíšková R. Detection of colistin-resistant *Acinetobacter baumannii* with the mcr-4 gene. *Klin Mikrobiol Infekc Lek.* 2019; 25 (1):4-6.
 18. Oikonomou O, Sarrou S, Papagiannitsis CC, Georgiadou S, Mantzarlis K, Zakynthinos E, Dalekos GN, Petinaki E. Rapid dissemination of colistin and carbapenem-resistant *Acinetobacter baumannii* in Central Greece: mechanisms of resistance, molecular identification, and epidemiological data. *BMC Infect Dis.* 2015; 15: 559. doi: 10.1186/s12879-015-1297-x.
 19. Agodi A, Voulgari E, Barchitta M, Quattrocchi A, Bellocchi P, Poulou A, Santangelo C, Castiglione G, Giunta L, Romeo MA, Vrioni G, Tsakris A. Spread of a carbapenem- and colistin-resistant *Acinetobacter baumannii* ST2 clonal strain causing outbreaks in two Sicilian hospitals. *J Hosp Infect.* 2014; 86 (4):260-266. doi: 10.1016/j.jhin.2014.02.001.
 20. Trebosc V, Gartenmann S, Tötzl M, Lucchini V, Schellhorn B, Pieren M, Locuro S, Gitzinger M, Tigges M, Bumann D, Kemmer C. Dissecting Colistin Resistance Mechanisms in Extensively Drug-Resistant *Acinetobacter baumannii* Clinical Isolates. *MBio.* 2019; 10 (4): e01083-19. doi:10.1128/mBio.01083-19
 21. Forde BM, Zowawi HM, Harris PNA, Roberts L, Ibrahim E, Shaikh N, Deshmukh A, Sid Ahmed MA, Al Maslamani M, Cottrell K, Tremblizzi E, Sundac L, Yu HH, Li J, Schembri MA, Whiley DM, Paterson DL, Beatson SA. Discovery of mcr-1-Mediated Colistin Resistance in a Highly Virulent *Escherichia coli* Lineage. *mSphere.* 2018; 3(5): e00486-18. doi:10.1128/mSphere.00486-18
 22. Imirzalioglu C, Falgenhauer L, Schmiedel J, Waezsada SE, Gwozdzinski K, Roschanski N, Roesler U, Kreienbrock L, Schiffmann AP, Irrgang A, Käsbohrer A, Bauerfeind R, Domann E, Chakraborty T. Evaluation of a Loop-Mediated Isothermal Amplification-Based Assay for the Rapid Detection of Plasmid-Encoded Colistin Resistance Gene mcr-1 in Enterobacteriaceae Isolates. *Antimicrob Agents Chemother.* 2017; 61 (4): e02326-16. doi:10.1128/AAC.02326-16
 23. Aruhomukama D, Sserwadda I, Mboowa G. Investigating colistin drug resistance: The role of high-throughput sequencing and bioinformatics. *F1000Res.* 2019; 8: 150. doi: 10.12688/f1000research.18081.2
 24. Lima T, Domingues S, Da Silva GJ. Plasmid-Mediated Colistin Resistance in *Salmonella enterica*: A Review. *Microorganisms.* 2019; 7(2): 55. doi:10.3390/microorganisms7020055
 25. Liu Y-Y, Wang Y, Walsh TR, Yi L-X, Zhang R, Spencer J, Doi Y, Tian G, Dong B, Huang X, Yu L-F, Gu D, Ren H, Chen X, Lv L, He D, Zhou H, Liang Z, Liu J-H, Shen J. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect Dis.* 2016; 16 (2): 161–168. doi:10.1016/S1473-3099(15)00424-7.
 26. Bai F, Li X, Niu B, Zhang Z, Malakar PK, Liu H, Pan Y, Zhao Y. A mcr-1-Carrying Conjugative IncX4 Plasmid in Colistin-Resistant *Escherichia coli* ST278 Strain Isolated from Dairy Cow Feces in Shanghai, China. *Front Microbiol.* 2018; 9: 2833. doi:10.3389/fmicb.2018.02833
 27. Carroll LM, Gaballa A, Guldmann C, Sullivan G, Henderson LO, Wiedmann M. Identification of Novel Mobilized Colistin Resistance Gene mcr-9 in a Multidrug-Resistant, Colistin-Susceptible *Salmonella enterica* Serotype Typhimurium Isolate. *MBio.* 2019; 10 (3). pii: e00853-19. doi: 10.1128/mBio.00853-19.
 28. Xavier BB, Lammens C, Ruhal R, Kumar-Singh S, Butaye P, Goossens H, Malhotra-Kumar S. Identification of a novel plasmid mediated colistin-resistance gene, mcr-2, in *Escherichia coli*, Belgium, June 2016. *Euro Surveill.* 2016; 21 (27): pii=30280. doi: 10.2807/1560-7917.ES.2016.21.27.30280.
 29. Yin W, Li H, Shen Y, Liu Z, Wang S, Shen Z, Zhang R, Walsh TR, Shen J, Wang Y. Novel plasmid-mediated colistin resistance gene mcr-3 in *Escherichia coli*. *MBio.* 2017; 8(3). pii: e00543-17. doi: 10.1128/mBio.00543-17.
 30. Yang YQ, Li YX, Lei CW, Zhang AY, Wang HN. Novel plasmid-mediated colistin resistance gene mcr-7.1 in *Klebsiella pneumoniae*. *J Antimicrob Chemother.* 2018;

- 73 (7) :1791-1795. doi: 10.1093/jac/dky111.J
31. Wang X, Wang Y, Zhou Y, Li J, Yin W, Wang S, Zhang S, Shen J, Shen Z, Wang Y. Emergence of a novel mobile colistin resistance gene, mcr-8, in NDM-producing *Klebsiella pneumoniae*. *Emerg Microbes Infect.* 2018; 7(1): 122. doi: 10.1038/s41426-018-0124-z.
 32. Carattoli A, Villa L, Feudi C, Curcio L, Orsini S, Luppi A, Pezzotti G, Magistrali CF. Novel plasmid-mediated colistin resistance mcr-4 gene in *Salmonella* and *Escherichia coli*, Italy 2013, Spain and Belgium, 2015 to 2016. *Euro Surveill.* 2017; 22 (31). pii: 30589. doi: 10.2807/1560-7917.ES.2017.22.31.30589.
 33. Borowiak M, Fischer J, Hammerl JA, Hendriksen RS, Szabo I, Malorny B. Identification of a novel transposon-associated phosphoethanolamine transferase gene, mcr-5, conferring colistin resistance in d-tartrate fermenting *Salmonella enterica* subsp. *enterica* serovar Paratyphi B. *J Antimicrob Chemother.* 2017; 72 (12): 3317-3324. doi: 10.1093/jac/dkx327.
 34. AbuOun M, Stubberfield EJ, Duggett NA, Kirchner M, Dormer L, Nunez-Garcia J, Randall LP, Lemma F, Crook DW, Teale C, Smith RP, Anjum MF. mcr-1 and mcr-2 variant genes identified in *Moraxella* species isolated from pigs in Great Britain from 2014 to 2015. *J Antimicrob Chemother.* 2018; 73 (10): 2904. doi: 10.1093/jac/dky272.
 35. Carroll LM, Wiedmann M, den Bakker H, Siler J, Warchocki S, Kent D, Lyalina S, Davis M, Sischo W, Besser T, Warnick LD, Pereira RV. 2017. Whole-genome sequencing of drug-resistant *Salmonella enterica* isolates from dairy cattle and humans in New York, and Washington States reveals source and geographic associations. *Appl Environ Microbiol.* 2017; 83 (12). pii: e00140-17. doi: 10.1128/AEM.00140-17.
 36. Wang R, van Dorp L, Shaw LP, Bradley P, Wang Q, Wang X, Jin L, Zhang Q, Liu Y, Rieux A, Dorai-Schneiders T, Weinert LA, Iqbal Z, Didelot X, Wang H, Balloux F. The global distribution and spread of the mobilized colistin resistance gene mcr-1. *Nat Commun.* 2018; 9 (1): 1179. doi:10.1038/s41467-018-03205-z
 37. Hasman H, Hammerum AM, Hansen F, Hendriksen RS, Olesen B, Agersø Y, Zankari E, Leekitcharoenphon P, Stegger M, Kaas RS, Cavaco LM, Hansen DS, Aarestrup FM, Skov RL. Detection of mcr-1 encoding plasmid-mediated colistin-resistant *Escherichia coli* isolates from human bloodstream infection and imported chicken meat, Denmark 2015. *Euro Surveill.* 2015;20(49). doi: 10.2807/1560-7917.ES.2015.20.49.30085.
 38. Skov RL, Monnet DL. Plasmid-mediated colistin resistance (mcr-1 gene): three months later, the story unfolds. *Euro Surveill.* 2016; 21 (9): 30155. doi: 10.2807/1560-7917.ES.2016.21.9.30155.
 39. Grami R, Mansour W, Mehri W, Bouallègue O, Boujaâfar N, Madec JY, Haenni M. Impact of food animal trade on the spread of mcr-1-mediated colistin resistance, Tunisia, July 2015. *Euro Surveill.* 2016; 21 (8): 30144. doi: 10.2807/1560-7917.ES.2016.21.8.30144.
 40. Kempf I, Fleury MA, Drider D, Bruneau M, Sanders P, Chauvin C, Madec JY, Jouy E. What do we know about resistance to colistin in Enterobacteriaceae in avian and pig production in Europe? *Int J Antimicrob Agents.* 2013; 42 (5): 379-383. doi: 10.1016/j.ijantimicag.2013.06.012.
 41. Fernandes MR, Moura Q, Sartori L, Silva KC, Cunha MP, Esposito F, Lopes R, Otutumi LK, Gonçalves DD, Dropa M, Matté MH, Monte DF, Landgraf M, Francisco GR, Bueno MF, de Oliveira Garcia D, Knöbl T, Moreno AM, Lincopan N. Silent dissemination of colistin-resistant *Escherichia coli* in South America could contribute to the global spread of the mcr-1 gene. *Euro Surveill.* 2016; 21 (17). doi: 10.2807/1560-7917.ES.2016.21.17.30214.
 42. Rapoport M, Faccone D, Pasteran F, Ceriana P, Albornoz E, Petroni A; MCR Group, Corso A. mcr-1-mediated colistin resistance in human infections caused by *Escherichia coli*: First description in Latin America. *Antimicrob Agents Chemother.* 2016; 60 (7):4412-4413. doi: 10.1128/AAC.00573-16.
 43. Islam KBMS, Shiraj-Um-Mahmuda S, Hazzaz-Bin-Kabir M. Antibiotic usage patterns in selected broiler farms of Bangladesh and their Public Health Implications. *J Public Health Dev Ctries.* 2016; 2(3): 276-284.
 44. Islam A, Rahman Z, Monira S, Rahman MA, Camilli A, George CM, Ahmed N, Alam M. Colistin resistant *Escherichia coli* carrying mcr-1 in urban sludge samples: Dhaka, Bangladesh. *Gut Pathog.* 2017; 9: 77. doi:10.1186/s13099-017-0227-4
 45. Bardet L, Rolain JM. Development of New Tools to Detect Colistin-Resistance among Enterobacteriaceae Strains. *Can J Infect Dis Med Microbiol.* 2018; 2018: 3095249. doi:10.1155/2018/3095249
 46. Esposito F, Fernandes MR, Lopes R, Muñoz M, Sabino CP, Cunha MP, Silva KC, Cayô R, Martins WMBS, Moreno AM, Knöbl T, Gales AC, Lincopan N. Detection of Colistin-Resistant MCR-1-Positive *Escherichia coli* by Use of Assays Based on Inhibition by EDTA and Zeta Potential. *J Clin Microbiol.* 2017;55(12):3454–3465. doi:10.1128/JCM.00835-17
 47. Dalmolin TV, de Lima-Morales D, Barth AL. Plasmid-mediated Colistin Resistance: What Do We Know? *J Infectiology.* 2018; 1(2): 16-22.
 48. Coppi M, Cannatelli A, Antonelli A, Baccani I, Di Pilato V, Sennati S, Giani T, Rossolini GM. A simple phenotypic method for screening of MCR-1-mediated colistin resistance. *Clin Microbiol Infect.* 2018; 24 (2): 201.e1-201.e3. doi: 10.1016/j.cmi.2017.08.011.

49. Osei Sekyere J. Mcr colistin resistance gene: a systematic review of current diagnostics and detection methods. *Microbiologyopen*. 2019; 8 (4): e00682. doi:10.1002/mbo3.682
50. Nijhuis RH, Veldman KT, Schelfaut J, Van Essen-Zandbergen A2, Wessels E, Claas EC, Gooskens J3. Detection of the plasmid-mediated colistin-resistance gene mcr-1 in clinical isolates and stool specimens obtained from hospitalized patients using a newly developed real-time PCR assay. *J Antimicrob Chemother*. 2016; 71 (8): 2344-6. doi: 10.1093/jac/dkw192.
51. Gao R, Hu Y, Li Z, Sun J, Wang Q, Lin J, Ye H, Liu F, Srinivas S, Li D, Zhu B, Liu YH, Tian GB, Feng Y. Dissemination and Mechanism for the MCR-1 Colistin Resistance. *PLoS Pathog*. 2016;12 (11): e1005957. doi: 10.1371/journal.ppat.1005957
52. Xu F, Zeng X, Hinenoya A, Lin J. MCR-1 Confers Cross-Resistance to Bacitracin, a Widely Used In-Feed Antibiotic. *mSphere*. 2018; 3(5). pii: e00411-18. doi: 10.1128/mSphere.00411-18.
53. Manyi-Loh C, Mamphweli S, Meyer E, Okoh A. Antibiotic Use in Agriculture and Its Consequential Resistance in Environmental Sources: Potential Public Health Implications. *Molecules*. 2018; 23 (4). pii: E795. doi: 10.3390/molecules23040795.
54. Haque M, Sartelli M, McKimm J, Abu Bakar M. Healthcare-associated infections - an overview. *Infect Drug Resist*. 2018; 11: 2321–2333. doi:10.2147>IDR.S177247
55. Barlaam A, Parisi A, Spinelli E, Caruso M, Taranto PD, Normanno G. Global Emergence of Colistin-Resistant *Escherichia coli* in Food Chains and Associated Food Safety Implications: A Review. *J Food Prot*. 2019; 82 (8):1440-1448. doi: 10.4315/0362-028X.JFP-19-116.
56. Frost I, Van Boeckel TP, Pires J, Craig J, Laxminarayan R. Global Geographic Trends in Antimicrobial Resistance: The Role of International Travel. *J Travel Med*. 2019 May 22. pii: taz036. doi: 10.1093/jtm/taz036.
57. Petrosillo N, Taglietti F, Granata G. Treatment Options for Colistin Resistant *Klebsiella pneumoniae*: Present and Future. *J Clin Med*. 2019;8(7):934. doi:10.3390/jcm8070934
58. Joshi S. Hospital antibiogram: a necessity. *Indian J Med Microbiol*. 2010; 28 (4): 277-80. doi: 10.4103/0255-0857.71802.
59. de With K, Allerberger F, Amann S, Apfaltrer P, Brodt HR, Eckmanns T, Fellhauer M7, Geiss HK, Janata O, Krause R, Lemmen S, Meyer E, Mittermayer H, Porsche U, Presterl E, Reuter S, Sinha B, Strauß R, Wechsler Fördös A, Wenisch C, Kern WV. Strategies to enhance rational use of antibiotics in hospital: a guideline by the German Society for Infectious Diseases. *Infection*. 2016;44(3):395–439. doi:10.1007/s15010-016-0885-z
60. Hawkey PM, Warren RE, Livermore DM, McNulty CAM, Enoch DA, Otter JA, Wilson APR. Treatment of infections caused by multidrug-resistant Gram-negative bacteria: report of the British Society for Antimicrobial Chemotherapy/Healthcare Infection Society/British Infection Association Joint Working Party. *J Antimicrob Chemother*. 2018; 73 (suppl_3): iii2-iii78. doi: 10.1093/jac/dky027.
61. Kniel KE, Kumar D, Thakur S. Understanding the Complexities of Food Safety Using a “One Health” Approach. *Microbiol Spectr*. 2018; 6(1). doi: 10.1128/microbiolspec.PFS-0021-2017.
62. Walia K, Sharma M, Vijay S, Shome BR. Understanding policy dilemmas around antibiotic use in food animals & offering potential solutions. *Indian J Med Res*. 2019; 149 (2): 107–118. doi: 10.4103/ijmr.IJMR_2_18
63. Haque M. Antibiotic Use, Antibiotic Resistance, and Antibiotic Stewardship –A Global Public Consequences. *Bang J Med Sci*. 2019; 18 (02): 169-170. doi: <https://doi.org/10.3329/bjms.v18i2.40680>
64. Carlet J, Jarlier V, Harbarth S, et al. Ready for a world without antibiotics? The Pensières Antibiotic Resistance Call to Action. *Antimicrob Resist Infect Control*. 2012;1(1):11. doi:10.1186/2047-2994-1-11
65. Saliba-Gustafsson EA, Borg MA, Rosales-Klitz S, Nyberg A, StålsbyLundborg C. Maltese Antibiotic Stewardship Programme in the Community (MASPIC): protocol of a prospective quasiexperimental social marketing intervention. *BMJ Open*. 2017; 7 (9): e017992. doi:10.1136/bmjopen-2017-017992
66. Che Roos NA, Bakar MA, Haque M. Knowledge, attitude and practice among Malaysian medical students, doctors, other health professionals, and common people regarding antibiotic use, prescribing and resistance: A systematic review. *Adv Hum Biol* 2019; 9 (3): 179-183. doi: 10.4103/AIHB.AIHB_42_19
67. Rahman SZ, Mishra A. Discovery of “Aligarh Super Bug” and Widespread variants of NDM1: A concern in antimicrobial resistance. *Bang J Med Sci*. 2017; 16 (03): 335-336. doi: org/10.3329/bjms. v16i3.32842