Original Article

Antibiotic Susceptibility Pattern of Bacteria Isolated from an Urban Referral Hospital in Dhaka City

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Abstract

The present study was to assess the pattern of antibiotic susceptibility of the isolated bacteria in an urban referral hospital in Dhaka City. A total of 393 bacterial strains were isolated from various specimens over a 10-months period. The majority of the organisms were Escherichia coli (33.33%) followed by Klebsiella species (27.48%), Staphylococcus aureus (17.05%), Acinetobacter species (8.14%), Pseudomonas species (7.12%), and others. The thirdgeneration Cephalosporins like Ceftriaxone, Ceftazidime and Cefotaxime were sensitive against 45-66% isolated Enterobacteriaceae. The first- and second-generation Cephalosporins were less effective. The sensitivity to Ciprofloxacin of various Enterobacteriaceae was only between 33-40% compared to 52.8-67.9% against Gentamicin. Majority of the Enterobacteriaceae were resistant to Ampicillin, whereas almost all of the Enterobacteriaceae (94-100%) were sensitive to Imipenem. About 97.0% Acinetobacter species were susceptible to Imipenem. Sensitivity of the organism (Acinetobacter) to third-generation Cephalosporins ranged between 50-56%, whereas 40.6% were found sensitive to Ciprofloxacin. The sensitivity to Chloramphenicol, Co-trimoxazole, Cephalexin and Ampicillin ranged between 9.3% to 34.3%. About 93.0% of Pseudomonas species were sensitive to Imipenem. The rate of susceptibility to Gentamicin and Netilmicin was higher than those of the Ciprofloxacin and Ceftriaxone (67.8% and 53.5% vs. 39.2%). About 70% of isolated S. aureus were resistant to Oxacillin but all were sensitive to Vancomycin. The result of this study would help the physicians to make a judicious choice of antibiotics for therapeutic purposes.

Key words: Antibiotic susceptibility, bacterial isolates, urban hospital, Enterobacteriaceae

Introduction

Infectious diseases still remain a major cause of morbidity and mortality in third-world countries including Bangladesh. Bacterial pathogens resistant to commonly

antimicrobials are now creating a challenge to the clinicians and researchers. The multi-drug-resistant organisms is a serious medical problem that has significantly affected the treatment of infectious diseases1,2 and has become a major clinical concern globally.3,4 Bacterial resistance pattern to antimicrobial agents can differ significantly from one country to another and within a country as evidenced by several recent surveillance studies,5-8

Introduction of newer antimicrobial agents is usually followed sooner or later by emergence of bacterial resistance Fax: 880-2-861 3004 to these drugs for many reasons.9 Development of multi-drug

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resistance in clinical isolates like *Salmonella typhi*, Pseudomonas species and Klebsiella species has been reported in Bangladesh. 10-12 The study on antibiotic susceptibility pattern is particularly important in developing countries that do not control antibiotic usage and maintain adequate epidemiological surveillance.

Therefore, the present study has been designed to find out the antibiotic resistance patterns of medically important bacteria in an urban hospital of Dhaka City. The outcome of this study might enable to determine the trend of drug resistance prevailing in Bangladesh.

Methods

This was a prospective study carried out in an urban hospital of Dhaka, Bangladesh. All the samples were collected from hospitalized and non-hospitalized patients of Bangladesh Institute of Research of Diabetic, Endocrine and Metabolic Disorders (BIRDEM) hospital, Dhaka. Samples were collected over a 10-months period during January to October, 2005 from both sexes and different age groups. A total of 393 clinical isolates were tested and the specimens included were Urine, Pus, Sputum, Vaginal swab, Throat swab and Conjunctival swab.

All samples were routinely cultured on MacConkey and Blood agar plates. In addition to these plates, Chocolate agar media were used for culturing Pus, Vaginal swab and Conjunctival swab specimens. After overnight incubation, plates were checked for the presence of suspected pathogens. All the suspected colonies were identified by colony characteristics, motility, Gram staining results, and biochemical reactions.¹³

Antimicrobial susceptibility test of the isolated organisms was done by disk diffusion method using the Kirby-Bauer technique¹⁴ and as per recommendations of the National Committee for Clinical Laboratory Standards.⁶ Antimicrobial agents used for determining antibiogram of isolated organisms were Penicillin (10 units), Ampicillin (10 μg), Cefuroxime (30 μg), Ceftazidime (30 μg), Cefotaxime (30 μg), Co-trimoxazole (25 μg), Imipenem (10 μg), Gentamicin (10 μg), Tetracycline (30 μg), Ciprofloxacin (5 μg),

Vancomycin (30 μ g), Chloramphenicol (30 μ g), Netilmicin (30 μ g), Oxacillin (1 μ g), Erythromycin (15 μ g), Rifampicin (5 μ g), Fusidic acid (10 μ g), Cephalexin (30 μ g) and Ceftriaxone (30 μ g). All disks were obtained from Oxoid Ltd., Basinstoke, Hampsire, UK.

In order to monitor the quality of the test results, on each day of testing the reference ATCC strains *Escherichia coli* ATCC 25922 and *Staphylococcus aureus* ATCC 29213 were included. The zone of inhibition was compared with recommended standard values.¹⁵

Results

A total of 393 strains of bacteria were isolated from various specimens. The specimens were Urine (189, 48.09%), Sputum (88, 22.39%), Pus (71, 18.07%), Throat swab (20, 5.09%) and others (25, 6.36%), including Vaginal and Conjunctival swabs.

Out of 393 organisms isolated, majority were *Escherichia coli* (131, 33.33%) followed by Klebsiella species (108, 27.48%). Other isolates included *Staphylococcus aureus* (67, 17.05%), Acinetobacter species (32, 8.14%), Pseudomonas species (28, 7.12%) and Providencia species (11, 2.80%). (Table I)

Table I: Pattern of organisms isolated from various samples

| Serial | Name of organisms | Number | Percentage |
|--------|-----------------------|--------|------------|
| 1. | Escherichia coli | 131 | 33.33 |
| 2. | Klebsiella species | 108 | 27.48 |
| 3. | Staphylococcus aureus | 67 | 17.05 |
| 4. | Acinetobacter species | 32 | 8.14 |
| 5. | Pseudomonas species | 28 | 7.12 |
| 6. | Providencia species | 11 | 2.80 |
| 7. | Proteus species | 09 | 2.29 |
| 8. | Enterobacter species | 01 | 0.25 |
| 9. | Others* | 06 | 1.53 |
| | Total | 393 | 100 |

^{*}Others indicate S. saprophyticus (n=3), S. epidermidis (n=2) and betahemolytic streptococci (n=1)

Among the total bacterial isolates, 260 (66.6%) were members of the Enterobacteriaceae family including *E. coli*, Klebsiella species, Providencia species, Proteus species, and Enterobacter species. Considering susceptibility pattern with different antimicrobials, it is found that almost all of the Enterobacteriaceae were sensitive to Imipenem (94-100%). The third-generation Cephalosporins like Ceftriaxone, Ceftazidime and Cefotaxime were sensitive against 45-66% isolated Enterobacteriaceae. The sensitivity to Ciprofloxacin of various Enterobacteriaceae was only between 33-40%. Sensitivity to Gentamicin ranged between 52.8-67.9%. (Table II)

Table II: Sensitivity of the isolated Enterobacteriaceae (n=260)

| Organisms | Number (%) sensitive to- | | | | | | | | | | | | | |
|------------------------------|--------------------------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--|
| | Amp | Cepl | CXM | CAZ | CRO | CTX | Tet | Gen | Cip | Chl | Cot | Imp | Net | |
| E. coli (n=131) | 10 | 28 | 49 | 74 | 76 | 80 | 43 | 89 | 53 | 83 | 31 | 129 | 78 | |
| | (7.6) | (21.3) | (37.4) | (56.4) | (58.0) | (61.1) | (32.8) | (67.9) | (40.4) | (63.3) | (23.6) | (98.4) | (59.5) | |
| Klebsiella spp (n=108) | 6 | 19 | 38 | 60 | 56 | 62 | 44 | 57 | 39 | 56 | 33 | 102 | 67 | |
| | (5.5) | (17.5) | (35.1) | (55.5) | (51.8) | (57.4) | (40.7) | (52.8) | (36.1) | (51.8) | (30.5) | (94.4) | (62.0) | |
| Providencia | 01 | 02 | 05 | 06 | 06 | 05 | 06 | 07 | 04 | 05 | 04 | 11 | 08 | |
| spp (n=11) | (9.0) | (18.1) | (45.4) | (54.5) | (54.5) | (45.4) | (54.5) | (63.6) | (36.3) | (45.4) | (36.3) | (100) | (72.7) | |
| Proteus | 01 | 02 | 04 | 05 | 05 | 06 | 04 | 06 | 03 | 04 | 02 | 09 | 05 | |
| spp (n=9) | (11.1) | (22.2) | (44.4) | (55.5) | (55.5) | (66,6) | (44.4) | (66.6) | (33.3) | (44.4) | (22.2) | (100) | (55.5) | |
| Enterobactes | 00 | 00 | 00 | 00 | 00 | 00 | 00 | 00 | 00 | 00 | 00 | 01 | 00 | |
| spp (n=1) | | | | | | | | | | | | (100) | | |

Amp=Ampicillin, Cepl=Cephalexin, CFM=Cefixime, CAZ=Ceftazidime, CRO=Ceftriaxone, CTX=Cefotaxime, Tet=Tetracycline, Gen=Gentamycin, Cip=Ciprofloxacin, Chl=Chloramphenicol, Cot=Co-trimoxazole, Imp=Imipenem, Net=Netilmycin

Almost all of Acinetobacter species (31, 96.8%) were susceptible to Imipenem, while 23 (71.8%) and 21 (65.6%) of the isolates were sensitive to Tetracycline and Gentamicin respectively. The sensitivity to third-generation Cephalosporins ranged between 50-56%, whereas 40.6% were found sensitive to Ciprofloxacin. The sensitivity of the organism to Chloramphenicol, Co-trimoxazole, Cephalexin and Ampicillin were 34.3%, 31.2%, 15.6% and 9.3% respectively.

Out of 28 isolates of Pseudomonas species, 26 (92.8%) were sensitive to Imipenem. Only 01 (3.5%) isolate was susceptible to Ampicillin, while 12 (42.8%) were sensitive to Carbenicillin and Cefotaxime. The sensitivity of the organism to Ciprofloxacin and Ceftriaxone was 39.2% each, while the rate of susceptibility to Gentamicin and Netilmicin was higher than the formers (67.8% and 53.5%). Susceptibility to Co-trimoxazole, Chloramphenicol and Tetracycline were between 17.8%-32.1% respectively. (Table III)

Table III: Antibiogram of isolated Acinetobacter species and Pseudomonas species by disk diffusion method

| Organisms | | Number (%) sensitive to- | | | | | | | | | | | | |
|----------------|-------|--------------------------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--|
| | Amp | Cepl | CXM | CAZ | CRO | CTX | Tet | Gen | Cip | Chl | Cot | Imp | Net | |
| Acinetobacte | 03 | 05 | 15 | | 18 | 16 | 23 | 21 | 13 | 11 | | 31 | 18 | |
| (n=32) | (9.3) | (15.6) | (46.8) | (53.1) | (56.2) | (50.0) | (71.8) | (65.6) | (40.6) | (34.3) | (31.2) | (96.8) | (56.2) | |
| Pseudomonas | 01 | i e | 127 | 13 | 11 | 12 | 09 | 19 | 11 | 08 | 05 | 26 | 15 | |
| spp (n=28)* | (3.5) | * | (10) | (46.4) | (39.2) | (42.8) | (32.1) | (67.8) | (39.2) | (28.5) | (17.8) | (92.8) | (53.5) | |

^{*} Carbenicillin was tested in addition and 12 (42.8%) Pseudomonads were sensitive

Considering sensitivity of *Staphylococcus aureus*, it was found that only 05 (7.4%) and 08 (11.9%) of the isolates were sensitive to Penicillin and Ampicillin respectively. Only 20 (29.8%) isolates were sensitive to 1 µg Oxacillin showing a calculated 70.2% of the isolates being Methicillin resistant or MRSA. Only 23 (34.3%) of the isolates were sensitive to Ciprofloxacin. However, all of them were sensitive to Vancomycin and the sensitivity to Rifampicin and Fusidic acid ranged between 85-88%. (Table IV)

Table IV: Antibiogram of isolated S. aureus by disk diffusion method

| Organisms | Number (%) sensitive to- | | | | | | | | | | | | | |
|-----------|--------------------------|--------|-------|-----------|--------|--------|--------|--------|--------|--------|--------|-------|--------|--------|
| | Pen | Anp | Ox | Cepi | Cip | Ery | Tet | Gen | Chl | Cot | Net | Van | Rif | Fus |
| S. aureus | 05 | 08 | 20 | 25 | 23 | 15 | 24 | 39 | 44 | 18 | 53 | 67 | 57 | 59 |
| (n=67) | (7.4) | (11.9) | (29.5 | 5)(37.31) | (34,3) | (22.3) | (35.8) | (58.2) | (65.6) | (26.8) | (79.1) | (100) | (85.0) | (88.0) |

Amp=Ampicillin, Cepl=Cephalexin, CXM=Cefuroxime, CAZ=Ceftazidime, CRO=Ceftriaxone, CTX=Cefotaxime, Tet=Tetracycline, Gen=Gentamicin, Cip=Ciprofloxacillin, Chl=Chloramphenicol, Cot=Co-trimoxazole, Imp=imipenem, Net=Netilmicin, Pen=Penicillin, Ox=Oxacillin, Ery=Erythromycin, Rif= Rifampicin, Van=Vancomycin and Fus = Fusidic acid

Discussion

Antibiotic resistance is a common phenomenon in developing countries where drugs are available freely without prescription. Now-a-days, antibiotics have been used extensively and newer antibiotics are continuously being added for the treatment of various infections. Proper use of antibiotics is very important in reducing unnecessary expenses, development of resistance to useful and life-saving antibiotics as well as to minimize many side effects. The resistance pattern varies from one country to another.

In the present study, most of the Enterobacteriaceae were sensitive to Imipenem ranging between 94-100%. This high level of sensitivity to Imipenem could be due to its restricted and limited use in the clinical practice. The drug has only recently been introduced in Bangladesh and is very expensive which has further restricted its widespread use. Similar effectiveness of Imipenem has also been reported from other countries, 5, 7, 16

The third-generation Cephalosporins like Ceftriaxone, Ceftazidime and Cefotaxime were sensitive against 45-66% isolated Enterobacteriaceae. The first- and second-generation Cephalosporins were less effective. In the United States, the frequency of resistance to Ceftazidime has increased from 1.5% to 3.6% from 1991 to 1997 as reported by the National Nosocomial Infections Surveillance system. A surveillance trial involving 102 medical centers in the United States detected 10.3% and 23.8% Ceftazidime-resistant *E. coli* and *K. pneumoniae* respectively.¹⁶

However, the sensitivity of various Enterobacteriaceae to Ciprofloxacin in the present study was only between 33-40%. This low-level of sensitivity to Quinolones and Cephalosporins was the result of very extensive use of these antibiotics in clinical practice. A large majority of patients were found prescribed by these drugs on their first contact with physicians.¹⁰

All the *Staphylococcus aureus* isolated in this study were sensitive to Vancomycin whereas 70% were Methicillin resistant (MRSA). The prevalence of MRSA differs strongly between countries.¹⁷ In the present study, very high isolation rate of MRSA was found as detected by 1 μg Oxacillin disk. Another study with wound specimens from diabetic patients in Bangladesh in 1994, reported an isolation rate of 37.2% and 21.6% MRSA amongst the hospitalized and non-hospitalized

diabetics respectively.¹⁸ The present findings show that the prevalence of MRSA in hospitalized patients has increased significantly over time.

This study would help the physicians to make judicious choice of antibiotics and would be helpful for formulation of an antibiotic policy.

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[Conflict of interest: None declared]