Original Article

Diagnosis of Bacterial Vaginosis by Acridine Orange Staining and its Comparison to Conventional Methods and Association of *Gardnerella vaginalis* with Bacterial Vaginosis.

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Abstract

The present study was undertaken to establish the efficacy of acridine orange staining for diagnosis of Bacterial Vaginosis (BV) and association of *Gardnerella Vaginalis* with BV. Two hundred sexually active females in the age group of 15-45 years, with vaginal discharge and itching, were selected for the study. A detailed history and a thorough clinical examination of all the cases were done. After making the clinical diagnosis of BV by Amsel’s criteria, diagnosis also carried out with Acridine orange staining, Gram stain, Nugent criteria and isolation of *Gard. vaginalis* from vaginal fluid. Out of 200 women, 48 (24%) cases were diagnosed as having bacterial vaginosis by applying Amsel’s clinical criteria. The rate of detection of bacterial vaginosis was 23% by Gram stain Nugent criteria and 24.5% by acridine orange staining. Out of the total 48 BV cases, the rate of detection of BV was 100% by Acridine orange staining and 93.87% by Gram-stain Nugent criteria. By Acridine orange staining *Lactobacillus* and pus cell were also detected which provided an important information about vaginal ecosystem. Acridine orange staining was the most sensitive (100%) method considering Amsel’s criteria as gold standard. The sensitivity of Gram stain Nugent criteria was 93.75% in this study. The specificity of Acridine orange staining was 99.55% and the specificity of Gram stain Nugent criteria was 99.10%. The positive predicative values of acridine orange staining and Gram staining Nugent criteria were 97.96% and 95.74% and the negative predictive values of these tests were 100% and 98.65% respectively. The results of Acridine orange staining test correlated well with that of Amsel’s clinical criteria among the study cases and healthy controls.

Key words: Bacterial Vaginosis, Sexually Transmitted Diseases, Amsel’s criteria, Acridine orange, Nugent criteria, *Gardnerella vaginalis*.

Introduction

Bacterial vaginosis (BV) is the most common cause of abnormal vaginal discharge and one of the most prevalent lower genital tract infection affecting almost one third (29%) of the women of reproductive age.¹ The BV is a clinical condition characterized by a thin, gray, homogenous, malodorous adherent vaginal discharge having pH > 4.5, release of a fishy odour on addition of 10% KOH to the vaginal fluid (whiff test), presence of clue cell and a few or no Lactobacilli.²

The normal flora of vagina, Lactobacillus, which under physiologic condition, produces an acidic milieu by transforming glycogen into lactic acid through hydrogen per oxide production, this lactic acid suppresses the growth of other organisms. Change in the normal vaginal flora causes change in pH which allows BV associated organisms like *Gardnerella vaginalis* and other anaerobes to overgrow and cause chronic infection and discharge.³ Symptom of BV depends on concentration of *Gardnerella vaginalis*.⁴ Almost
50% of BV patients remain asymptomatic. Although symptoms occur, the manifestation of BV is mild, so usually overlooked in developing countries like Bangladesh. Diagnosis of BV is important for its serious complications such as premature rupture of membrane, miscarriage, development of pelvic inflammatory diseases, increase risk of acquiring STD such as HIV and also increase genital tract HIV shedding. 
The predominant organism that causes BV are *Gardnerella vaginalis*, *Mycoplasma hominis* and *Ureaplasma urealiticum*. Other anaerobes such as *Prevotella*, *Mobiluncus*, Bacteroides and Peptostreptococcus have also been identified as flora associated with BV. Recently discovered strict anaerobe *Atopobium vaginae* is another organism that is strongly associated with BV, but its role in BV is still unknown. In women with BV, *Gardnerella vaginalis* is almost universally present (99%) and majority have *A. vaginae* (96%) infection. *Gardnerella vaginalis* has been reported to be present in small numbers (< 10⁴ cfu/ml) in asymptomatic women and its count of 2×10⁶ cfu/ml have always been associated with symptom of BV.

The optimal method of diagnosing BV remains controversial. There are two main categories of diagnostic tests for BV: clinical criteria and laboratory based testing. The most widely accepted clinical criteria are Amsel’s clinical criteria. This clinical criteria require 3 of the following 4 signs such as vaginal fluid pH >4.5, >20% epithelial cells are clue cell, specific characteristic vaginal discharge of BV and a positive whiff test. Evaluation of the clinical parameters are more subjective and vary with the skill of the clinician. 

For laboratory testing Gram stain of vaginal smear was standardized by Nugent score. The result of Nugent score varies from person to person specially when different microscopes are being used. Culture of vaginal fluid is often used as a primary laboratory test in the past but this is found to be of little value in the diagnosis of BV as *Gardnerella vaginalis* is isolated in 83% to 94% of women with sign of BV but is also recovered in 36% to 55% of healthy women, as it is a normal flora of the human vagina. Oligonucleotide probe test for detection of *Gardnerella vaginalis*, detection of amine and fatty acid by electrophoresis and gas liquid chromatography, proline aminopeptidase and sialidase assay etc. are available. Among other laboratory methods, ELISA for detection of anti-hemolysin antibody of *Gardnerella vaginalis*, PCR for detection of BV associated bacteria, Affirm VP III microbial identification system, Quick Vue R advanced pH and amine test card for detection of BV are available in the advanced countries and these tests are restricted in the research laboratory.

The use of acridine orange stain for detection of clue cells is shown to be very much effective for diagnosis of BV. Acridine orange differentially stains the micro-organisms from cellular materials. Acridine orange staining technique has been recommended for the rapid identification of clue cells in vaginal smears. By acridine orange stain clue cell (orange staining bacteria on green epithelial cell) is clearly seen, number of Lactobacilli and pus cell can be observed. Other two common vaginal pathogens, Candida and Trichomonas can also be detected by this method.

Diagnosis of BV by Amsel’s clinical criteria and Gram stain Nugent criteria is not yet practiced in clinical laboratories in Bangladesh as it is time consuming and laborious. Diagnosis of BV is still based on only detection of clue cell by wet microscopy and Gram stain method.

Therefore the present study has been designed to establish an easy, simple, relatively inexpensive test for early diagnosis of bacterial vaginosis by acridine orange staining test which can help in proper diagnosis and treatment of BV and prevention of its complications.

**Methods**

This was a prospective study carried out in the department of Microbiology, Dhaka Medical College. Two hundred patients attending Gynae outpatient department of Dhaka Medical College and Maternal and Child Health Training Institute, Azimpur, Dhaka with history of abnormal characteristics vaginal discharge and itching suspected to have BV were included in the study.

Under all aseptic precaution, vaginal examination was done. At the time of speculum examination, the presence of vaginal discharge was noted, the vaginal pH was recorded and amine test was done.

(i) Physical examination of discharge:
Specific characteristic vaginal discharge of BV patient was noted which was thin, gray/white, homogenous, foul smelling (fishy odor) and adherent to the vaginal wall.

(ii) Measurement of vaginal fluid pH:

pH was measured by placing a pH paper (Merck, UK) by sponge holding forceps into the vagina during vaginal examination and colour change was observed. When the pH of vaginal fluid was > 4.5, the cases were suspected as BV cases.

(iii) Amine test (Whiff test):

One drop of 10% KOH was added with vaginal discharge and immediately sniffed for the fishy odor.

(iv) Wet Mount preparation:

One drop of normal saline was taken on a glass slide and vaginal discharge was added and covered with a cover slip. Then the slide was examined under light microscope.
BV was diagnosed by the presence of clue cell in association with at least 2 of the other 3 Amsel’s clinical criteria.

Diagnosis of bacterial vaginosis by Gram stain Nugent criteria:
Each Gram stained smear was evaluated for the following morphotypes under oil immersion (×1000 magnification) according to the Nugent criteria.

Acridine orange test:
The unfixed dried smear was stained with the Acridine orange staining and microscopic examination was done initially with 10 × objective to see the distribution of fluorescing material, and then with 40×objective to identify T. vaginalis, yeast cells, clue cell and pus cell.

Identification of different structures in acridine orange staining:
Normal epithelial cells were stained pale green colour with green coloured nucleus. Lactobacillus stained green colour. Gardnerella vaginalis stained yellowish orange. Leucocytes (pus cells)-yellow-green.

Trichomonas. vaginalis was identified by orange red staining with yellow-green nucleus. Yeast cells were stained–orange colour.

In bacterial vaginosis, the orange staining bacteria adhered to the green epithelial cells were clue cells. Inflammatory cell stained yellowish green with yellow to orange coloured nucleus.

Culture for isolation of Gardnerella vaginalis:
Human blood bilayer Tween 80 (HBT) agar media was inoculated directly from swab sample and was incubated in the candle extinction jar containing water-soaked cotton at 37°C for 48-72 hours for primary isolation of Gard. vaginalis. The plates were examined by oblique lighting after 24 hrs., 48 hrs. and 72 hrs.

Then semi quantitative estimation of Gardnerella vaginalis in culture was done. The number of colonies were counted and expressed as 1+ to 4+ according to the colonies found in different streaking zone.

Growth was quantitated as follows:
1+, < 10 colonies in the first streaking zone;
2+, > 10 colonies in the first zone and , < 10 colonies in the 2nd zone;
3+, > 10 colonies in the 2nd zone and < 10 colonies in the 3rd zone;
4+, > 10 colonies in the 3rd zone.

Identification of Gardnerella vaginalis:
i. Colonial morphology: Colonies on HBT agar were identified as small white colonies with β-hemolysis after 48 or 72 hours of incubation.

   ii. Clear β-hemolysis with diffuse edges on HBT media, but no hemolysis on sheep blood agar.
   iii. The typical cell morphology in a Gram stained smear from a colony were pleomorphic, Gram-variable or Gram-negative cococcabilli or small rods.
   iv. Catalase test negative.
   v. Oxidase test negative.

Results
All BV cases were diagnosed using the presence of clue cells’ and two or more of the other three Amsel’s criteria.

Of the 200 patients, 48 (24%) were positive for BV by Amsel’s clinical criteria, 46 (23%) patients were positive for BV by Gram stain Nugent criteria 49(24.5%) were positive by acridine orange staining and 51 (25.5%) were culture positive for G. vaginalis (Table 1).

Out of 49 acridine orange staining positive patients, 48 were positive by Amsel’s criteria and one was negative. All the patients who were positive by Amsel’s clinical criteria were positive by acridine orange staining. Out of 46 cases detected by Gram stain Nugent criteria, one was negative by Amsel’s clinical criteria. Amsel’s criteria detected additional 3 cases of BV who were negative by Gram stain Nugent criteria, Of the 51 women who were positive for Gard. vaginalis by culture, 5 were negative by Amsel’s criteria.

Table 1: Comparison of Amsel’s criteria with Gram stain Nugent criteria, acridine orange staining and culture of vaginal fluid for Gard. vaginalis for diagnosis of BV.

<table>
<thead>
<tr>
<th>BV diagnosis by Amsel’s criteria</th>
<th>Gram stain</th>
<th>Acridine orange Staining</th>
<th>Culture for G. vaginalis</th>
</tr>
</thead>
<tbody>
<tr>
<td>BV cases</td>
<td>Gram stain</td>
<td>Acridine orange Staining</td>
<td>Culture for G. vaginalis</td>
</tr>
<tr>
<td>(n=48)</td>
<td>Positive</td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>non BV</td>
<td>45 (93.75)</td>
<td>3 (6.25)</td>
<td>48(100.00)</td>
</tr>
<tr>
<td></td>
<td>46(95.83)</td>
<td>2 (4.17)</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>200</td>
<td>154</td>
<td>49</td>
</tr>
</tbody>
</table>

Notes: ** highly significant
Among the BV cases diagnosed by different methods, 46 (95.83%) of the 48 cases were diagnosed by Amsel’s criteria, 43 (93.48%) of the 46 cases by Gram stain Nugent criteria and 47 (95.92%) of the 49 cases by Acridine orange staining were positive for G. vaginalis in culture (Table-2).

Table 2: Isolation of G. vaginalis among BV cases diagnosed by different methods.

<table>
<thead>
<tr>
<th>Culture</th>
<th>BV positive cases</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Amsel’s criteria</td>
</tr>
<tr>
<td></td>
<td>(n=48)</td>
</tr>
<tr>
<td>G. vaginalis</td>
<td>Gram stain Nugent criteria</td>
</tr>
<tr>
<td></td>
<td>(n=46)</td>
</tr>
<tr>
<td></td>
<td>Acridine orange stain</td>
</tr>
<tr>
<td></td>
<td>(n=49)</td>
</tr>
<tr>
<td>Positive</td>
<td>46 (95.83)</td>
</tr>
<tr>
<td>Negative</td>
<td>2 (4.17)</td>
</tr>
</tbody>
</table>

Figures in parentheses represent percentages.

In wet film examination, 51 (25.5%) were positive for clue cell, 16 (8%) were positive for Trichomonas vaginalis and 62 (31%) were positive for Candida. In 11 cases, both clue cell and Trichomonas vaginalis were found. By Gram staining method, 49 (24.5%) were positive for clue cell, 9 (4.5%) were positive for Trichomonas vaginalis and 48 (24%) were positive for Candida. By Acridine orange staining 49 (24.5%) were positive for clue cell, 22 (11%) were positive for Trichomonas vaginalis and 69 (34.50%) were positive for Candida (Table-3).

Table 3: Detection of clue cell, Trichomonas vaginalis and Candida by wet film examination, Gram staining & acridine orange staining of vaginal fluid among the study cases.

<table>
<thead>
<tr>
<th>Vaginal infection</th>
<th>Wet film examination</th>
<th>Gram staining</th>
<th>Acridine orange staining</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clue cell</td>
<td>40+11=51(25.50)</td>
<td>49(24.50)</td>
<td>49(24.50)</td>
</tr>
<tr>
<td>Trichomoniasis</td>
<td>5+11=16 (8.00)</td>
<td>9(4.50)</td>
<td>22(11.00)</td>
</tr>
<tr>
<td>Candidiasis</td>
<td>62(31.00)</td>
<td>48(24.00)</td>
<td>69(34.50)</td>
</tr>
</tbody>
</table>

Figures in parentheses represent percentages.

*11 patients had both trichomoniasis and bacterial vaginosis.

In order to determine the pathogenic role of G. vaginalis in BV, quantitative estimation of G. vaginalis in culture has been recommended. In this study, most of the BV cases yielded 3+ to 4+ growth of G. vaginalis in culture but non BV cases and healthy controls yielded 1+ or 2+ growth.

In this study, wet film microscopy detected 51 (25.5%) clue cell, 16(8%) Trichomonas vaginalis and 62 (31%) Candida respectively. Gram stain method detected 49(24.50%) clue cell, 9(4.5%) Trichomonas vaginalis and 48(24%) Candida. Acridine orange stain detected 49 clue cells, 22 Trichomonas vaginalis and 69 Candida. Among 51 clue cell positive cases (by wet film), 48 were positive for BV by Amsel’s criteria. Of the 49 clue cell positive cases by Gram stain method, 45 women were categorized into BV group according to Gram stain Nugent criteria. By Acridine orange staining 49 BV...
cases were diagnosed by observing clue cell, condition of Lactobacillus and pus cell. Out of them 48 were also BV positive by Amsel’s clinical criteria. Wet film examination detected additional 3 clue cell positive cases who were BV negative by Amsel’s criteria. Gram stain method failed to detect 3 BV cases who were clue cell positive but negative by Nugent criteria. Amsel’s criteria also missed one case who was BV positive by acridine orange staining. So further study is required with molecular method to confirm whether the case was false positive or not. Moreover, for diagnosis of BV cases by Amsel’s criteria, only wet film microscopy is not sufficient. Presence of clue cell among the patients with BV ranges from 85 to 97 percent compared with 5 to 14 percent women without BV. For diagnosis of BV by clue cell in wet microscopy requires at least 20 fields examination and also additional 2 criteria which is time consuming and more subjective. On the other hand, BV can be diagnosed directly by examining the Acridine orange stained vaginal fluid smear. Moreover, wet film microscopy may over diagnose the BV cases. But by acridine orange staining method BV cases can be diagnosed accurately because by this technique clue cell, Lactobacillus and pus cell are better visualized. So Acridine orange staining is better than any other method.

The Acridine orange test detected highest number (49) of BV cases. In this study, acridine orange staining, Gram stain Nugent criteria and Amsel’s criteria detected 49(100%), 46(93.87%) and 48 (94.11%) BV cases respectively. In contrast to the present study, Nunns et al. (1996) detected 63 (77.81%), 40 (49.4%) and 48(59.3%) BV cases by Acridine orange staining, Gram stain Nugent criteria and Amsel’s criteria respectively.

The Gram stain Nugent criteria based on bacterial morphotype count is also laborious. The reliability of diagnosing BV by Gram stain method has been improved by a scoring system (0-10) of interpretation by Nugent. The ‘intermediate’ score group of Gram stain Nugent criteria has a higher (32%) chance of transition to BV. Women of this group are more susceptible to acquisition of HIV and other STDs as they do not receive any treatment and thereby increased risk of complications of BV.

The results of Gram stain varies from person to person specially when different microscopes are used as the actual field diameter is not mentioned in the system. Moreover, sometimes Gram stain cannot differentiate very short form of Lactobacillus from Gardnerella vaginalis Clue cell can also be found in intermediate score group. The women of this group may develop BV which can be missed if only Gram stain Nugent score is done for diagnosis of BV.

The acridine orange test yielded excellent sensitivity and specificity with respect to Amsel’s clinical criteria, Gram-stain Nugent criteria and culture of G. vaginalis. The sensitivity was very high (100%) within the range of acceptable values as a reliable diagnostic test, the specificity was also high (99.55%) and acceptable.

References


