Bacteriological profile and antimicrobial susceptibility patterns of blood culture isolates among bloodstream infection suspected patients attending in a referral hospital

Mahfuza Nasrin¹, Most. Fahmida Begum¹, Rezina Karim¹, Mohammad Shah Alam¹, Farha Rahman², Mohammad Moniruzzaman Bhuiyan³

¹Department of Microbiology, Uttara Adhunik Medical College, Dhaka, Bangladesh, ²Department of Microbiology, Bangladesh Medical College, Dhaka, Bangladesh, ³Department of Pediatrics, Sir Salimullah Medical College, Dhaka, Bangladesh.

Abstract
Bloodstream infections (BSIs) are the leading cause of morbidity and mortality in people of all ages. On time diagnosis and appropriate antimicrobial treatment is of utmost important to save the lives of affected ones. The present study is aimed to determine the bacterial profile and antimicrobial susceptibility patterns of blood culture isolates among BSI suspected patients attending in Uttara Adhunik Medical College and Hospital from 1st July to 31st December, 2020. This cross sectional study was conducted among 1,675 BSI suspected patients. About 10 ml of venous blood for adults and 2-3 ml for children was collected aseptically and transferred into an automated blood culture bottle. The BD BACTEC FX40 automated blood culture method was used to isolate bacterial pathogens and antimicrobial susceptibility test was performed by Kirby-Bauer disc diffusion method following CLSI guidelines. The rate of bacteriologically confirmed cases was 222/1675 (13%). Majority of BSI were caused by gram negative bacteria predominantly Salmonella Typhi (51%) followed by Salmonella paratyphi (15%), Escherichia coli (15%), Klebsiella spp. (7%), Staphylococcus aureus (6%), Pseudomonas aeruginosa (4%). Salmonella Typhi and Salmonella Paratyphi isolates showed 100% susceptibility to meropenem and ceftriaxone. A higher percentage of strains of Salmonella Typhi (80%,84%,85%) and Salmonella Paratyphi (93%,95%,95%) were found sensitive to amoxiclav, chloramphenicol and cotrimoxazole whereas only 4% isolates were sensitive to ciprofloxacin which is an alarming situation. None of the antibiotic was 100% effective against Escherichia coli, Klebsiella spp. and Pseudomonas aeruginosa. Amikacin and meropenem were found more effective against gram negative bacteria. In this study, none of the S. aureus strains showed resistance to cloxacillin, amoxiclav, gentamicin and doxycycline and sensitivity to azithromycin and clindamycin were found 40% and 50% respectively. Multidrug resistance among blood stream isolates are increasing in a threatening way which need to be addressed through effective surveillance and infection control strategies.

Introduction
The term bloodstream infection (BSI) generally refers to the growth of a microorganism from a blood culture obtained from a patient with clinical signs of infection and where contamination has been ruled out¹. It is characterized by the presence of viable bacterial or fungal microorganisms in the bloodstream that elicit inflammatory response and often accompanied by alteration of clinical, laboratory and hemodynamic parameters². Bacterial bloodstream infection (BSI) is a global concern. It may range from self-limiting infections to life-threatening sepsis that requires rapid and aggressive antimicrobial treatment³. It is often associated with increased length of hospital stay which ultimately leads to significant amount of healthcare related costs and a high rate of morbidity and mortality⁴.

Many bacteria such as Staphylococcus aureus, alphahemolytic Streptococci, Escherichia coli, Klebsiella pneumoniae, Pseudomonas aeruginosa, Salmonella Typhi and Acinetobacter species have been reported as a cause of bacteremia with variation in distribution from place to place.⁵ As an example, Salmonella enterica is a frequently isolated pathogen from blood samples in both African and Asian regions, however their serotypes differ substantially. Salmonella Paratyphi is the predominant organism in the Salmonella group in Africa
Salmonella Typhi is the most frequently isolated organism in Asia. Besides their isolation rate and their antibiotic susceptibility pattern varies substantially.

Depending on the age, the severity of infection, other risk factors and the mortality rate for BSI varies between 4.0 and 41.5%. Globally, bloodstream infection affects about 30 million people leading to 6 million deaths, with 3 million newborns and 1.2 million children suffering from sepsis annually. Changing patterns of epidemiology, lack of proper antimicrobial guidelines in the locality, the emergence of antimicrobial resistance and paucity of good diagnostic facilities are connected to the surge in BSI associated morbidity and mortality.

Increasing antimicrobial resistance is a worldwide concern. It is a serious challenge for health care professionals in prescribing suitable antimicrobial therapy as many bacterial pathogens have developed resistance to most of the antibiotics. This is specially true in countries like Bangladesh due to lack of national guideline for antibiotic use, absence of good laboratory facilities to do antimicrobial drug susceptibility test and overuse or misuse of antibiotics without any rational prescription. In most of the cases, antimicrobial therapy is initiated empirically before the results of blood culture are available. Selection of right antibiotic for empirical therapy is of utmost importance. Continuous monitoring trends in the microbiology of BSI pathogens and their antibiotic susceptibility patterns are therefore important to guide empiric antibiotic treatment strategies and infection control programs.

The present study is aimed to determine the bacterial profile and antimicrobial susceptibility patterns among BSI suspected patients in a referral hospital to assist the health care professionals to choose an appropriate antimicrobial therapy for treatment of BSIs.

Materials and Methods
For this cross sectional study, a total of 1,675 samples from clinically suspected cases of bloodstream infection were collected at Uttara Adhunik Medical College and Hospital, Dhaka for a period of six months from 1st July 2020 to 31st December 2020. All the samples were collected from inpatient’s and outpatient’s department of our hospital during the study period and processed in Microbiology laboratory. About 10 ml of venous blood for adults and 2-3 ml for children was collected aseptically using 70% alcohol and 2% tincture iodine and transferred in to automated blood culture bottles. The BD BACTEC FX40 automated blood culture method was used. In case of a positive growth, the BD BACTEC FX40 automatically gives an alert. Blood culture bottles with no alert signal of bacterial growth after recommended days of incubation is considered culture negative. The positive bottles were sub cultured on MacConkey’s agar, blood agar and chocolate agar media. The chocolate agar plates were incubated inside a candle jar to provide 5-10% CO₂, whereas the other two agar plates (blood agar and MacConkey’s agar) were incubated aerobically for 18-24 h at 37°C.

Isolates were further processed according to standard operating procedure (SOP) of the laboratory for its complete identification. Pure cultures of bacterial isolates were subsequently subjected to species identification and confirmation. gram positive isolates were identified using catalase and coagulase tests. Isolates of members of Enterobacteriaceae family were identified biochemically by means of a series of tests: catalase, indole, citrate, urease, H₂S production and triple-sugar iron. Non lactose fermenting Gram negative bacteria were identified by indole, triple-sugar iron, urease, oxidase and catalase tests. Antimicrobial susceptibility tests were performed by using the Kirby-Bauer disc diffusion method and susceptibility patterns were determined following CLSI guidelines. Diameters of the zone of inhibition were measured to the nearest millimeter and categorized as sensitive, intermediate and resistant according to CLSI guidelines. Isolates were classified as either susceptible or resistant to an antibiotic and all the isolates with intermediate resistance were classified as resistant. Culture media and antibiotic discs used in the study were obtained from Oxoid Ltd., UK.

Quality control
Reference strains of Escherichia coli (ATCC 25922), Pseudomonas aeruginosa (ATCC 27853) and Staphylococcus aureus (ATCC 25923) were used as a control strains for identification and drug susceptibility testing. Quality control for media was done by randomly taking the prepared culture media and incubating overnight to see for any growth. In this study multi-drug resistance (MDR) was defined as simultaneous resistance to more than two antimicrobial agents. Isolates of Staphylococcus aureus were further tested for methicillin resistance according to the CLSI guidelines by using cefoxitin disc.
Results

Table-1: Distribution of bacterial growth among all participants (n=1,675)

<table>
<thead>
<tr>
<th>Bacterial growth</th>
<th>Number</th>
<th>Percentage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Growth</td>
<td>222</td>
<td>13</td>
</tr>
<tr>
<td>No growth</td>
<td>1453</td>
<td>87</td>
</tr>
<tr>
<td>Total</td>
<td>1675</td>
<td>100</td>
</tr>
</tbody>
</table>

During the study period, 1,675 blood cultures were analyzed, of which 222 (13%) bacteria were isolated. Of the total positive cases, 132 (59%) were males and 90 (41%) were females giving a male: female ratio of 1.5:1 (Figure -1).

Figure-1: Gender variation regarding growth positive cases (n=222)

![Gender variation](image)

Table-2: Distribution of growth positive cases by age group (n=222)

<table>
<thead>
<tr>
<th>Age</th>
<th>No. of growth positive cases (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Up to 18</td>
<td>62 (28%)</td>
</tr>
<tr>
<td>&gt; 18</td>
<td>160 (72%)</td>
</tr>
</tbody>
</table>

Among 222 blood culture positive cases, 62 patients were up to 18 years of age and 160 patients were above 18 years of age. The rate of isolation was noted higher among age group above 18 years of age when compared to the age group up to 18 years.

Table-3: Types of bacteria isolated in blood culture (n=222)

<table>
<thead>
<tr>
<th>Gram negative organisms</th>
<th>Types of bacteria isolated</th>
<th>No. of isolates (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Salmonella Typhi</td>
<td>114 (51%)</td>
</tr>
<tr>
<td></td>
<td>Salmonella Paratyphi</td>
<td>34 (15%)</td>
</tr>
<tr>
<td></td>
<td>Escherichia coli</td>
<td>34 (15%)</td>
</tr>
<tr>
<td></td>
<td>Klebsiella spp.</td>
<td>15 (7%)</td>
</tr>
<tr>
<td></td>
<td>Pseudomonas aeruginosa</td>
<td>9 (4%)</td>
</tr>
<tr>
<td></td>
<td>Acinetobacter spp.</td>
<td>2 (1%)</td>
</tr>
<tr>
<td>Gram positive organisms</td>
<td>Staphylococcus aureus</td>
<td>12 (6%)</td>
</tr>
<tr>
<td></td>
<td>Streptococcus pneumoniae</td>
<td>2 (1%)</td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>222</td>
</tr>
</tbody>
</table>

Table-3 shows the distribution of organisms found throughout this study period. Salmonella Typhi was the most frequently isolated blood borne bacterial pathogen in this study, accounting for 51% of the total isolates. Other frequently isolated pathogens were Salmonella Paratyphi (15%), Escherichia coli (15%), Klebsiella species (7%), Staphylococcus aureus (6%), Pseudomonas aeruginosa (4%), Acinetobacter species (1%) and Streptococcus pneumoniae (1%).

Table-4: Antibiotic susceptibility pattern of isolated Salmonella species (n=148)

<table>
<thead>
<tr>
<th>Organisms</th>
<th>AMC %</th>
<th>CIP %</th>
<th>CFM %</th>
<th>CRO %</th>
<th>C %</th>
<th>SXT %</th>
<th>AZM %</th>
<th>MEM %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Salmonella Typhi (n=114)</td>
<td>80</td>
<td>4</td>
<td>96</td>
<td>100</td>
<td>84</td>
<td>85</td>
<td>83</td>
<td>100</td>
</tr>
<tr>
<td>Salmonella Paratyphi (n=34)</td>
<td>93</td>
<td>4</td>
<td>95</td>
<td>100</td>
<td>95</td>
<td>95</td>
<td>68</td>
<td>100</td>
</tr>
</tbody>
</table>

AMC-Amoxiclav, CIP-Ciprofloxacin, CFM-Cefixime, CRO-Ceftriaxone, C-Chloramphenicol, SXT-Cotrimoxazole, AZM-Azithromycin, MEM-Meropenem

Salmonella Typhi and Salmonella Paratyphi isolates showed 100% susceptibility to meropenem and ceftriaxone. A higher percentage of strains of Salmonella Typhi (80%,...
84% and 85%) and Salmonella Paratyphi (93%, 95% and 95%) were found sensitive to amoxiclav, chloramphenicol and cotrimoxazole indicating that the resistance trend might have started reversing. On the other hand, 83% Salmonella Typhi and 68% Salmonella Paratyphi were susceptible to azithromycin. Only 4% isolates of Salmonella Typhi and Salmonella Paratyphi were sensitive to ciprofloxacin which is an alarming situation (Table-4).

**Table-5: Antibiotic susceptibility pattern of isolated S. aureus (n=12)**

<table>
<thead>
<tr>
<th>Organisms</th>
<th>AMC %</th>
<th>DOX %</th>
<th>CIP %</th>
<th>SXT %</th>
<th>CN %</th>
<th>AZM %</th>
<th>CD %</th>
<th>CLOX %</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. aureus</td>
<td>100</td>
<td>100</td>
<td>70</td>
<td>70</td>
<td>100</td>
<td>40</td>
<td>50</td>
<td>100</td>
</tr>
</tbody>
</table>

AMC-Amoxiclav, CIP-Ciprofloxacin, DOX-Doxycycline, CD-Clindamycin, SXT-Cotrimoxazole, CN-Gentamycin, AZM-Azithromycin, CLOX-Cloxacillin

All the isolates of S. aureus were sensitive (100%) to amoxiclav, doxycycline, cloxacillin and gentamicin. S. aureus isolates were 70% responsive to ciprofloxacin and cotrimoxazole. On the other hand, sensitivity to azithromycin and clindamycin were found 40% and 50% respectively (Table-5).

**Table-6: Antibiotic susceptibility pattern of gram negative isolates**

<table>
<thead>
<tr>
<th>Organisms</th>
<th>AMC %</th>
<th>CIP %</th>
<th>CXM %</th>
<th>CFX %</th>
<th>CRO %</th>
<th>CAZ %</th>
<th>MEM %</th>
<th>AK %</th>
<th>TET %</th>
<th>ATM %</th>
<th>SXT %</th>
<th>CN %</th>
<th>TAZP %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Escherichia Coli (n=34)</td>
<td>25</td>
<td>36</td>
<td>30</td>
<td>30</td>
<td>30</td>
<td>95</td>
<td>88</td>
<td>82</td>
<td>29</td>
<td>47</td>
<td>76</td>
<td>85</td>
<td></td>
</tr>
<tr>
<td>Klebsiella spp. (n=15)</td>
<td>37</td>
<td>56</td>
<td>44</td>
<td>50</td>
<td>50</td>
<td>44</td>
<td>88</td>
<td>70</td>
<td>55</td>
<td>44</td>
<td>50</td>
<td>62</td>
<td>57</td>
</tr>
</tbody>
</table>

AMC-Amoxiclav, CIP-Ciprofloxacin, CXM-Cefuroxime, CFX-Cefixime, CRO-Ceftriaxone, CAZ-Ceftazidime, MEM-Meropenem, SXT-Cotrimoxazole, AK-Amikacin, TET-Tetracycline, ATM-Aztreonam, CN-Gentamycin, TZP-Tazobactam-piperacillin

Antibiotic susceptibility pattern of isolated gram negative bacteria is shown in Table-6. The observed sensitivity of E. coli was 95% to meropenem, 88% to amikacin, 85% to tazobactum-piperacillin and 76% to gentamicin. On the other hand, about 88%, 70% and 62% Klebsiella were sensitive to meropenem, amikacin and gentamycin respectively. Lower percentage of susceptibility were seen in case of E. coli (25%, 30% and 36%) and Klebsiella (37%, 50% and 56%) against amoxiclav, 3rd generation cephalosporins and ciprofloxacin respectively.

**Table-7: Antibiotic susceptibility pattern of Pseudomonas aeruginosa (n=9)**

<table>
<thead>
<tr>
<th>Organisms</th>
<th>AK %</th>
<th>ATM %</th>
<th>CIP %</th>
<th>LEV %</th>
<th>CAZ %</th>
<th>FEP %</th>
<th>CN %</th>
<th>TAZP %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pseudomonas Aeruginosa</td>
<td>75</td>
<td>62</td>
<td>62</td>
<td>62</td>
<td>75</td>
<td>62</td>
<td>95</td>
<td>75</td>
</tr>
</tbody>
</table>

CIP-Ciprofloxacin, CAZ-Ceftazidime, LEV-Levofloxacin, AK-Amikacin, ATM-Aztreonam, CN-Gentamycin, TZP-Tazobactam-piperacillin, FEP-Cefepime, MEM-Meropenem

Among the isolates of P. aeruginosa, 95% were found to be sensitive to tazobactum-piperacillin and on the contrary, 75% of the P. aeruginosa isolates were sensitive to meropenem, amikacin and cefepime (Table-7).

**Discussion**

Bloodstream infection (BSI) is a challenging problem and sometimes it may be life threatening; therefore, timely detection, identification and antimicrobial susceptibility testing of blood-borne pathogens are one of the most important functions of diagnostic microbiology laboratory. There exists a strong relationship between delay in effective initiation of therapy and in hospital mortality of septic shock. Each hour of delay in therapy initiation is associated with an average decrease in survival of 8%.

Knowledge of the hospital epidemiology and the antimicrobial susceptibility pattern of blood isolates help physicians to effectively manage blood stream infections. Bacteria isolation rate in this study was 13%, which is comparable with the result done in India (14%) and Germany (13%)11,12. It was however, lower than reports from Ethiopia (28%) and Gambia (34%)13,14. The reason may be most of the cases must have taken antibiotics before admission in our hospital and also self-medication is very common in Bangladesh as the antibiotics are commonly available over the counter. Such variation in blood culture positivity across the countries can be explained by various factors such as difference in blood culture system, volume or the number of blood culture samples, geographical location, nature of patient population, epidemiological difference of the etiological agents, and difference in infection control policies among nations.
In our study, more frequent bacteremia cases noted in case of males than females. This may be explained as men are involved in more physical activities for livelihood and less frequent hand hygiene practice which could potentially provide environment for large reservoirs of common pathogens responsible for causing blood stream infections.

In this study, S. Typhi (51%) was the predominant gram negative bacteria followed by S. Paratyphi (15%), E. coli, Klebsiella spp. and S. aureus. Several studies from Bangladesh have identified S. Typhi as a common cause of bloodstream infection in this region and reported Salmonella species to be responsible for almost half of the disease burden associated with BSI in Dhaka16. More or less similar observations have been seen in cases of bacteremia in different countries, though the proportion and prevalence of the bacterial agents varied8,15. As the only source of Salmonella infection is the infected human and fecal contamination of drinking water and food supplies, the highest percentage of Salmonella isolates in this study indicate the necessity of proper waste management and infection control practices.

Among the isolated Salmonella species Salmonella Typhi was more prominent than Salmonella Paratyphi. There is no such well-established reason behind the variation of Salmonella serovars; however, it can be assumed that the higher incidence of S. Typhi could be achieved via waterborne transmission as requires smaller inoculum than S. Paratyphi which requires larger inoculum via a foodborne transmission17.

In this study, we found all Salmonella Typhi and Salmonella Paratyphi isolates were susceptible to ceftriaxone and meropenem, but a very high level of reduced susceptibility exists against ciprofloxacin (4%). A high percentage of Salmonella Typhi (>80%) and Salmonella Paratyphi (>90%) isolates appeared to be sensitive to first line antityphoid agents like amoxiclav, chloramphenicol and cotrimoxazole, which is consistent with studies carried out in Nepal and Pakistan17,18. This might give us some hope that in future we can again start using these antimicrobials for treatment to Salmonella Typhi and Paratyphi. Our finding shows 83% Salmonella Typhi and 68% Salmonella Paratyphi were susceptible to azithromycin. A study from Nepal also reported a low rate of azithromycin resistance among tested antibiotics17. Several clinical studies have demonstrated good efficacy of azithromycin for the treatment of uncomplicated enteric fever in clinical and in vitro studies. On the other hand, a study from Netherland shows a high percentage of increased MICs for azithromycin19. In our study, we only used disc diffusion method for azithromycin; MIC was not done which is our limitation. From the susceptibility findings of Salmonella it is noticed that first line anti-typhoidal drugs were becoming more sensitive and sensitivity to ciprofloxacin is remarkably decreased. Ceftriaxone and meropenem use should be reserved for the treatment of MDR and XDR cases of enteric fever. It is necessary to understand the importance of proper reporting in context of microbiology, evaluation and monitoring of antimicrobial therapy18.

In this study all the isolates of S. aureus were found to be sensitive (100%) to cloxacillin, amoxiclav, doxycycline and gentamicin.

In the current study, most of the isolates of Escherichia coli, Klebsiella spp. and Pseudomonas aeruginosa showed extended drug resistance. None of the antibiotic was 100% effective against Escherichia coli, Klebsiella spp. and Pseudomonas aeruginosa.

E. coli isolates showed <30% sensitivity to 2nd and 3rd generation of cephalosporins. Resistance to fluoroquinolones remains high. The fact that cephalosporins are one of the most commonly used antibiotics for inpatient’s as well as for outpatient’s could be the reason for such low degree of sensitivity. Hence, rationalization of treatment strategies is very much warranted considering the local trends of BSIs12. The observed sensitivity of E. coli was 95% to carbapenems, 85% to tazobactum-piperacillin and 88% to amikacin. Similar findings have been observed across Saudi Arabia and China19,20. Klebsiella isolates showed sensitivity rate of 88% and 70% to meropenem and amikacin. For the Pseudomonas aeruginosa isolates tazobactum-piperacillin (95%), amikacin (75%) and meropenem (75%) were found more active. However other studies from Ethiopia and Nepal showed all the isolates of Gram negative bacteria were susceptible to meropenem21,22. Our study shows that multidrug resistant gram negative pathogens are on rise in our hospital.

We acknowledge several limitations of our study, like this was single center study with shorter duration and small number of isolates which may not reflect the true status of the antimicrobial pattern of wider community or even other hospitals.
Conclusion
Bacterial bloodstream infections are of great concern globally. The knowledge of bacteriological profile and antimicrobial sensitivity patterns of hospital is crucial for effective management of blood stream infection. Results of this study will help in providing useful guidelines for choosing an effective antibiotic in our hospital. Robust infection control practices and antibiotic stewardship programs may help to reduce incidence of blood stream infections as well as prevent the emergence of resistance. Furthermore, research should also focus on diagnostic stewardship, to establish newer, rapid, automated bacterial identification method and sensitivity analysis that will not only help early initiation of appropriate antimicrobial therapy for better patient outcome but will also defer bacterial resistance.

References
infection (BSI) is a global concern. It may range from often accompanied by alteration of clinical, laboratory the growth of a microorganism from a blood culture The term bloodstream infection (BSI) generally refers to requires rapid and aggressive antimicrobial treatment 3. It organism in the frequently isolated pathogen from blood samples in both health care professionals to choose an appropriate antimi- pathogens and their antibiotic susceptibility patterns are antibiotic for empiric therapy is of utmost importance. alcohol and 2% tincture iodine and transferred in to is the most frequently isolated organism Reference strains of Quality control measured to the nearest millimeter and categorized as subsequently subjected to species identification and strains for identification and drug susceptibility testing. Among 222 blood culture positive cases, 62 patients were seen for any growth. In this study multi-drug resistance species (n=222) analyzed, of which 222 (13%) bacteria were isolated. Of this study, accounting for 51% of the total isolates. Other Salmonella Typhi and Paratyphi isolates were susceptible to azithromycin. A Our finding shows 83% Salmonella Typhi and Paratyphi. TABLE-5: Antibiotic susceptibility pattern of isolated bacteria is shown in Table-6. The observed sensitivity of isolates were 70% responsive to ciprofloxacin such as difference in blood culture system, volume 2.  Viscoli C. Bloodstream infections: The peak of the therapy for better patient outcome but will also defer infection control practices. 2.  Viscoli C. Bloodstream infections: The peak of the hospital mortality of septic across the countries can be explained by various factors involved in more physical activities for livelihood and less prevalence of the bacterial agents varied 8,15. As the only CARPATELLI C, ABBETTO B. Acinetobacter E. coli, for outpatient’s could be the reason for such low degree of for the treatment of MDR and XDR should be reserved for the treatment of MDR and XDR 4.  Fichera F, Incianni F, Spadaro R. Three-year antimicrobial resistance trends in bloodstream infections at a large teaching hospital in China: a 20-year surveillance study (1998-2017). Antimicrobial resistance & infection control. 2019 Dec;8(1):1-8. 5.  Birru M, Woldemariam M, Manilal A, Aklilu A, Tsalla T, Mitiku A, et al. Bacterial profile, antimicrobial susceptibility patterns, and associated factors among bloodstream infection suspected patients attending Arba Minch General Hospital, Ethiopia. Scientific reports. 2021 Aug 5;11(1):1-3. 6.  Simkhada P, Raj SKC, Lamichhane S, Subedi S, Shrestha UT. Bacteriological profile and antibiotic susceptibility pattern of blood culture isolates from patients visiting tertiary care hospital in Kathmandu, Nepal. Global journal of medical research. 2016;16(1):1-8.