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# INFLUENCE OF PLANT GROWTH REGULATORS ON VEGETATIVE GROWTH, SEX EXPRESSION AND YIELD OF SUMMER BOTTLE GOURD

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#### Abstract

Field experiments on bottle gourd variety 'BARI Lau -4' were conducted at the Plant Physiology field of Horticulture Research Center, Bangladesh Agricultural Research Institute, Gazipur during two consecutive summer seasons of 2015 and 2016 to investigate the effect of plant growth regulators on growth, sex expression, yield and yield components of the crop. The experiment consisted of different concentrations of Gibberellic acid (GA<sub>3</sub>), Naphthalene acetic acid (NAA), Maleic hydrazide (MH) and single concentration of cycocel (CCC) viz., GA<sub>3</sub> @ 10 ppm, GA<sub>3</sub> @ 30 ppm, NAA @100 ppm, NAA @ 150 MH @ 50 ppm, MH @ 150 ppm and CCC @ 500 ppm along with ppm, distilled water considered as control. All growth regulators were sprayed to the seedlings at two-leaf stage and 4 days after the first spray. Growth regulator treatments had significant effect on primary branches/plant, node number of 1<sup>st</sup> male and female flower appearance, number of days to 1<sup>st</sup> male and female flower appearance, number of male and female flowers, sex ratio (male:female flower) number of fruits/plant, individual fruit weight and fruit yield. Spraying of MH @ 150 ppm gave the highest primary branches/plant (17.0 and 18.0 in first and 2<sup>nd</sup> year, respectively) and induced maximum female flowers (37.3 and 40.0 in first and 2<sup>nd</sup> year, respectively) at lower nodes followed by CCC @ 500 ppm (36.3) in the first year and MH @ 50 ppm (40.0) in the 2<sup>nd</sup> year. Application of MH @ 150 ppm caused early appearance of female flowers on the nearest node (from bottom). Application of MH @ 150 ppm gave the lower number of male flowers (81.7 and 96.0 in first and 2<sup>nd</sup> year, respectively) and the highest number of female flowers/plant, thereby producing lower male:female sex ratio (2.2 and 2.5 in first and 2<sup>nd</sup> year, respectively) and the maximum number of fruits/plant (12.0 and 14.0 in first and 2<sup>nd</sup> year, respectively). The maximum fruit weight/plant was obtained from the application of MH @ 150 ppm (29.3 and 35.8 in first and 2<sup>nd</sup> year, respectively) followed by CCC @ 500 ppm (26.0 in the 1<sup>st</sup> year and 29.0 in 2<sup>nd</sup> year). The highest fruit yield per hectare was recorded significantly with the application of MH @ 150 ppm (97.6 t/ha and 89.6 t/ha in first and 2<sup>nd</sup> year, respectively) closely followed by CCC @ 500 ppm (88.5 t/ha in 2015) and GA<sub>3</sub> @ 30 ppm (75.3 t/ha in 2016), as compared to

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other treatments. The highest mean yield over the years was also recorded at MH @ 150 ppm (93.6 t/ha) followed by CCC @ 150 ppm (80.6 t/ha) and GA<sub>3</sub> 30 (74.1). Application of MH @ 150 ppm gave the maximum gross return and net return with the highest BCR of 5.24 followed by CCC 500 ppm (4.15) and GA<sub>3</sub> 30 ppm (3.86).

Keywords: Growth regulators, sex expression, fruit yield, summer bottle gourd.

### Introduction

Bottle gourd [*Lagenaria siceraria* (Molina) Standle] locally known as 'Lau' or 'Kadu', is one of the important and popular vegetables of the family *Cucurbitaceae*, extensively grown throughout Bangladesh. Fruits at tender stage are used as a cooked vegetable and for preparation of sweets (e.g. *kheer, petha, burfi and halwa*), pickles and rayta. Hard shells of mature fruits are used as water jugs, domestic utensils, floats for fishing nets and making musical instruments, etc. As a vegetable it is easily digestible. It has cooling effect and has diuretic and having cardio-tonic properties. Fruit pulp is used as an antidote against certain poisons and is good for controlling constipation, night blindness and cough. It is especially recommended in the diet of patients suffering from high blood pressure. The composition of immature fruits of bottle gourd per 100 g of fresh edible portion consists of water 93.9 g, energy 88 kJ (21 kcal), protein 0.5 g, fat 0.1 g, carbohydrate 5.2 g, fiber 0.6 g, Ca 44 mg, P 34 mg, Fe 2.4 mg,  $\beta$ -carotene 25 µg, thiamin 0.03 mg, niacin 1.2 mg, ascorbic acid 10 mg (Leghari *et al.*, 2014).

The growth and yield of cultivated crop plants is mainly influenced by genetic and crop management factors; first factor involves in various breeding techniques while second factor involves cultural operation, plant protection and other agronomic practices, both these factors have been fully exploited by researchers. In recent years, scientists have given attention to the idea of regulating plant growth as third most important factor in improving growth and yield. It helps in efficient utilization of metabolites in certain physiological processes going in plant system (Krisnamurthy, 2011).

Bangladesh Agricultural Research Institute (BARI) has developed five bottle gourd varieties for cultivation in both winter and summer seasons. The variety 'BARI Lau-4' is recommended for cultivation in both winter and summer season. In summer season, the yield of bottle gourd is low. Besides many other reasons for low yield of bottle gourd, there is one acute problem of fewer female flowers and high sex ratio (male:female) during summer. In cucurbitaceous plants, sex expression is determined by gene as well as the environment. Huyskens *et al.* (1992) reported that significantly more male flowers were produced during summer, under long days and high temperature, than in winter, under short days and low temperature condition. Plant growth regulators (PGRs) such as Gibberellic acid (GA<sub>3</sub>), Naphthalene acetic acid (NAA), Maleic hydrazide (MH), Cycocel (CCC), etc. are found beneficial for induction of female flowers and reduction of male flowers in summer bottle gourd and sex expression can also be

controlled by using different growth regulators (Kooner *et al.*, 2000; Rahman and Karim, 1997; Hidayatullah *et al.*, 2012; Gaurav *et al.*, 2008 and Kumar *et al.*, 2000). Very little information is available on the effect of growth regulators on sex expression and yield of bottle gourd in Bangladesh. Therefore, this investigation was carried out to assess the effect of PGRs (GA<sub>3</sub>, NAA, MH and CCC) on vegetative growth and to narrow the sex ratio by increasing female flowers side by side, decreasing male flowers per plant for yield improvement of bottle gourd in summer.

#### **Materials and Methods**

The experiment was conducted at the Plant Physiology field of HRC, BARI, Gazipur during two consecutive summer seasons of 2015 and 2016. The variety 'BARI Lau-4' was used in this experiment. The experiment consisted of eight treatments *viz.*,  $T_0 = \text{control}$  (distilled water),  $T_1 = \text{GA}_3 @ 10 \text{ ppm}$ ,  $T_2 = \text{GA}_3 @ 30 \text{ ppm}$ ,  $T_3 = \text{NAA} @ 100 \text{ ppm}$ ,  $T_4 = \text{NAA} @ 150 \text{ ppm}$ ,  $T_5 = \text{MH} @ 50 \text{ ppm}$ ,  $T_6 = \text{MH} @ 150 \text{ ppm}$  and  $T_7 = \text{CCC} @ 500 \text{ ppm}$ . The sources of GA<sub>3</sub> and CCC were Isha, Chemical Pvt. Ltd. (India), and those of MH and NAA were BDH Chemicals Ltd. (England) and SD Fine Chemicals Ltd. (Mumbai, India), respectively. All chemicals were Laboratory Grade.

250 mg of gibberellic acid was accurately weighed out using sensitive electronic balance and dissolved in a few ml of alcohol (95%). The solution thus prepared was transferred to 250 ml volumetric flask containing distilled water. The volume of the solution was made upto 250 ml to get the 1000 ppm stock solution. NAA stock solution (1000 ppm) was prepared by dissolving 500 mg NAA powder in 500 ml of distilled water. 500 mg NAA powder was accurately weighed out and dissolved by keeping it in a 500 ml volumetric flask and by adding 1 N NaOH solution drop by drop till the powder was completely dissolved. The volume was adjusted to 500 ml with distilled water in a volumetric flask. Thus the stock solution was prepared. 500 mg MH was accurately weighed out using sensitive electronic balance and dissolved in a few ml of 1 N NaOH. 100 ml of distilled water was added in a 500 ml volumetric flask. The solution was then adjusted to 500 ml by adding distilled water. This gives 1000 ppm stock solution. 2 ml (50% ai) of CCC was taken by 10 ml pipette ans dissolved in distilled water by addingshaking method. The volume was adjusted to 1000 ml with distilled water in a volumetric flask to obtain a stock solution of 1000 ppm. Finally, the required lower concentrations of GA3 (10 and 30 ppm), NAA (100 and 150 ppm), MH (50 and 150 ppm) and CCC (500 ppm) were prepared from their stock solutions by using the formula:  $V_1 \times S_1 = V_2 \times S_2$ ; where  $S_1$ : concentration of stock solution (1000 ppm) of the desired chemical (GA<sub>3</sub>, NAA, MH or CCC),  $V_1 =$ volume of stock solution of desired chemical (what we have to calculate),  $S_2$ : concentration of desired chemical and V<sub>2</sub>: amount of solution required for spray treatment-wise. Then calculated amount  $(V_1)$  of desired chemical was taken from stock solution and poured into a pot of known volume and then required amount

of water was added into this pot. Hand sprayer was used for spray of the chemicals.

The experiment was laid out in a randomized complete block design (RCBD) with three replications. For raising seedlings sandy loam soil and well decomposed cowdung were thoroughly mixed in 1:1 ratio and then plastic pots were filled with this mixture. Seeds were placed in plastic pots on 01 March, 2015 and 11 February, 2016. Seeds were soaked in distilled water for 24 hours to facilitate germination. After final land preparation, pits were prepared by spade and size of each pit was 30 cm x 30 cm x 30 cm. One pit was made in the middle of each plot and one seedling was planted in that pit. Plot size was 2.0 m x 1.5 m and 2.0 m x 2.0 m in 2015 and 2016, respectively. Manure and fertilizers were applied as cow dung, N, P, K, S, B and Zn @ 10000, 80, 45, 88, 25, 1.8 and 4.5 kg/ha, respectively (Quamruzzaman et al., 2015). Half of cow dung; full doses of S, Zn and B, and P and K @ 30 kg/ha each were applied during final land preparation. The rest half of cow dung and P and K @ 15 kg/ha was applied as basal in pit. Rest of N and K was applied in 4 equal installments at 20 days interval starting from 20 days after transplanting. Seventeen day- old seedlings were transplanted on 19 March 2015 and 01 March 2016. First spraying of plant growth regulators (PGRs) were done at the 2 true leaf stage on March 23, 2015 and 03 March, 2016. Second spraying of PGRs was done after 4 days of first spray. Control plants were sprayed with distilled water. Trellises made of bamboos and iron nets were used for the support of each plant individually. Plants were allowed to be grown individually and not be allowed to intermingle with other plants grown beside. Weeding and irrigation were done as required. When 1<sup>st</sup> flower was seen, it was tagged and each male and female flower was tagged separately throughout the growing period, and the number of male and female flowers was counted.

Data were recorded on main vine length, number of primary branches/plant, number of leaves/plant, CCI (Chlorophyll Content Index), Fv/Fm (efficiency of photosystem II), days to first male and female flowers, node order of first and female flowers (from the bottom), number of male and female flowers/plant, number of fruits/plant, individual fruit weight, fruit length, fruit circumference and yield/plot. Then plot yield was converted to per hectare yield. Data on CCI (Chlorophyll Content Index) was taken by Chlorophyll Content Meter (Model: CCM-200, Opti-sciences, USA). The leaf discs were previously adapted to the dark for 20 minutes. The fluorescence data (Fv/Fm) were collected at 70 days after sowing within 10.00 am to 12.00 pm. In the first year (2015), the fruit harvest started from 21 May and ended in 18 June; while, in the 2<sup>nd</sup> year (2016), fruit harvest started from 09 May and ended in 29 June. Each fruit was harvested after twelve days of anthesis. Thus every fruit was harvested by sharp knife 12 days after anthesis. A total of 7 and 8 harvests were required in 2015 and 2016, respectively. Recorded data were statistically analyzed by MSTAT-C and mean separation was done by Tukey's W test at 5% level of probability.

## **Results and Discussion**

## Vegetative growth of bottle gourd plants

Plant growth regulators had significant effect on number of primary branches/plant in both the years. The number of leaves/plant was significantly influenced by growth regulators in 2015 but insignificant effect on length of main vine in both years (Table 1). The maximum number of primary branches/plant was recorded in the plants treated with MH @ 150 ppm (17.0 and 18.0 in 2015 and 2016, respectively; mean 17.5) closely followed by CCC @ 500 ppm (15.0 in both 2015 and 2016; mean 15.0) and MH @ 50 ppm (13.0 in 2015 and 15.0 in 2016, mean 14.00) and GA<sub>3</sub> @ 30 ppm (12.0 in 2015 and 16.0 in 2016, mean 14.0) and its minimum number from control in both years (6.0 in 2015 and 8.0 in 2016; mean 7.0). Ansari and Choudhury (2014) obtained significantly higher number of branches in bottle gourd by the application of GA<sub>3</sub> (50-100 ppm) and MH @ 50 ppm. Sadiq et al. (1990) reported to have the highest number of branches per plant in cucumber while spraying with CCC @ 500 ppm at 4 leaf stage. In the first year, spraying of MH @ 150 ppm gave the maximum number of leaves/plant (343.0) which was identical with MH @ 50 ppm (337.0) and CCC @ 500 ppm (341.0). Number of leaves/plant did not vary significantly with PGR treatments in the 2<sup>nd</sup> year, but numerically the PGR treated plants produced more leaves than control. The highest mean number of leaves/plant (325.0) was recorded at both MH @ 150 ppm and CCC @ 500 ppm. The control treatment gave the minimum number of leaves/plant in both the years.

Treatment		n vine le (m)	ength	Primary	y branches (no.)	s/plant	Leave	es/plant (	no.)
	2015	2016	Mean	2015	2016	Mean	2015	2016	Mean
$T_0$	12.2	12.0	12.1	6.0 e	8.0c	7.0	277 d	298	288.
$T_1$	12.2	12.1	12.1	11.0 cd	10.0c	10.5	304cd	310	307
$T_2$	12.1	13.2	12.6	12.0 bcd	16.0a	14.0	309 bcd	318	314
$T_3$	12.3	12.4	12.4	9.0 de	11.0bc	10.0	285d	300	294
$T_4$	11.9	13.2	12.6	9.0 de	10.0c	9.5	298 d	308	303
$T_5$	12.4	12.6	12.5	13.0 abc	15.0ab	14.0	337 abc	309	323
$T_6$	13.0	11.8	12.4	17.0 a	18.0a	17.5	343 a	307	325
$T_7$	13.00	11.8	12.4	15.0 ab	15.0ab	15.0	341 ab	309	325
CV(%)	3.94	10.37	-	10.97	10.73	-	3.62	5.31	-

 Table 1. Effect of plant growth regulators on vegetative growth of summer bottle gourd

Means showing dissimilar letters in a column are significantly different at 5% level by Tukey's W test.

 $T_0$  = Control (distilled water),  $T_1$  = GA<sub>3</sub> @10 ppm,  $T_2$  = GA<sub>3</sub> @ 30 ppm,  $T_3$  = NAA @ 100 ppm,  $T_4$  = NAA @ 150 ppm,  $T_5$  = MH @ 50 ppm,  $T_6$  = MH @ 150 ppm,  $T_7$  = CCC @ 500 ppm.

## Effect of PGRs on CCI and Fv/Fm values

In both the years all growth regulator treated plants produced identical chlorophyll content index (CCI) and Fv/Fm with control (Table 2). All the treatments gave Fv/Fm values close to 0.80 indicating photosystem II of the plants were not affected by the application of PGRs. The Fv/Fm values close to 0.800 revealed that all the plants under treatment were in healthier condition.

 Table 2. Effect of plant growth regulators on chlorophyll content and quantum yield (Fv/fm values) of summer bottle at 70 days after sowing

Treatment		CCI value			Fv/Fm value	
	2015	2016	Mean	2015	2016	Mean
$T_0$	20.2	21.3	20.7	0.80	0.81	0.81
$T_1$	20.3	21.5	20.9	0.80	0.81	0.81
$T_2$	21.4	22.3	21.9	0.80	0.82	0.81
$T_3$	20.5	20.2	20.4	0.81	0.81	0.81
$T_4$	20.6	21.2	20.9	0.81	0.81	0.81
$T_5$	20.7	20.1	20.4	0.80	0.80	0.80
$T_6$	20.7	20.8	20.8	0.80	0.82	0.80
$T_7$	20.4	21.3	20.9	0.80	0.80	0.80
CV (%)	5.20	4.88	5.13	3.08	4.60	3.52

 $T_0$  = Control (distilled water),  $T_1$  = GA<sub>3</sub> @ 10 ppm,  $T_2$  = GA<sub>3</sub> @ 30 ppm,  $T_3$  = NAA @ 100 ppm,  $T_4$  = NAA @ 150 ppm,  $T_5$  = MH @ 50 ppm,  $T_6$  = MH @ 150 ppm,  $T_7$  = CCC @ 500 ppm.

## Node number and number of days to first male and female flowers

Number of node and days required for the appearance of the first male flower were significantly lowered by all PGRs treatments except control (Table 3). The first male flower appeared at the lowest node (18.0 in 2015 and 16.3 in 2016; mean 17.15) on the plants treated with MH @ 150 ppm. The node numbers required for the first male flower were identical in NAA @ 100 ppm, NAA @ 150 ppm, MH @ 50 ppm, MH @ 150 ppm and CCC @ 500 ppm in the 1st year whereas, in 2nd year, node numbers were identical in all treatments except control. The plants treated with MH @ 150 ppm produced the first male flower in the least number of days in both years (59.0 in 2015, 63.0 in 2016 with a mean value 61.0. MH @ 150 ppm was statistically similar to GA<sub>3</sub> @ 30 ppm, and CCC @ 500 ppm with regard to number of days to first male flower appearance in 2<sup>nd</sup> year. All the PGR treatments significantly lowered the number of nodes and days required for the appearance of the first female flower than control in 2015 but, in 2016, application of MH both at 50 and 150 ppm significantly reduced node number at which female flower appeared and all the PGR treatments significantly lowered number of days required for the appearance of first female flower as compared to control (Table 3). The first

female flower appeared at the lowest node (21.0 in 2015 and 20.0 in 2016; mean 20.5) in the plants treated with MH @ 150 ppm, closely followed by CCC @ 500 ppm, MH @ 50 ppm and NAA @ 150 ppm during 2015, but in 2016, there was no significant differences among PGR treatments in respect of node number of female flower except control. Heslop-Harrison (1957) reported that the sexual differentiation is controlled by endogenous level of auxin content, which favours the formation of female flowers. Browning et al. (1992) explained that paclobutrazol induced a large, but transient increase in IAA (natural auxin) concentration within two days after treatment of the plants. These seem to be a reasonable explanation for the early appearance of female flowers at lower node. Similar findings have been reported with CCC by Sharma et al. (1988) and with MH by Singh and Choudhury (1988), Patel (1992) and Pandya and Dixit (1997) in bottle gourd. The plants treated with MH @ 150 ppm were able to produce the first female flower in least number of days in both the years (70.0 in 2015, 74.3 in 2016 and mean value 72.15. MH at 50 & 150 ppm and GA<sub>3</sub> @ 30 ppm gave identical number of days to 1<sup>st</sup> female flower in 2015 whereas, in 2016, MH @ 150 ppm was statistically similar to GA<sub>3</sub> @ 30 ppm with regard to number of days to first female flower appearance.

#### Male and female flowers

Spraying of CCC @ 500 ppm gave the highest number of male flowers (143.3 in 2015) followed by control treatment (123.0 in 2015) but in 2016, this treatment produced lower male flower than control (Table 3). The lowest male flowers/plant (81.7 in 2015, 96.0 in 2016 and mean 88.8) was recorded at MH @ 150 ppm, which was identical with NAA @ 100 and MH @ 50 ppm in 2015; but in 2016, it was identical with NAA @ 100 and 150 @ ppm. In 2015, all treatments significantly produced the maximum female flowers/plant while in 2016, except NAA @ 150 ppm all other treatments significantly increased number of female flowers/plant as compared to control. In the first year, the maximum number of female flowers/plant was found in MH @ 150 ppm (37.3 in 2016, 40 in 2016; mean 38.7) closely followed by CCC @ 500 ppm (36.3 in 2015, 38.0 in 2016; mean 38.0) and GA<sub>3</sub> @ 30 ppm (34.0 in 2016).

The CCC has been reported to cause reduction in respiration rate and accumulation of photosynthates, an effect similar to low temperature and short days (Choudhury, 1996) which might promote female flower production in cucurbits. Desai *et al.* (2011) obtained male flowers ranging from 86.27 -151 (CCC @ 250 ppm); 111.43 by control treatment and female flowers with a range of 29.7 (control) to 51.1 (PBZ @ 25 ppm); 40.5 by CCC. The present experimental result corroborates the rfindings of Hidayatullah *et al.* (2012). Pandey and Singh (1976) reported that spraying of NAA @ 100 and 150 @ ppm and GA<sub>3</sub> @ 10 ppm increased the female flowers and decreased the male flowers per plant in sponge gourd compared to control. The present experimental results were also in agreement with the results of Pandey and Singh (1976) and Arora *et al.* (1982).

Table 3. Effect of plant grow	fect of pl:	ant growth	h regulat	ors on sex	expression	n of summ	vth regulators on sex expression of summer bottle gourd	burd					384
	Node at	Node at which the	e 1 <sup>st</sup> male	Numbe	Number of days to 1st male	) 1 <sup>st</sup> male	Node at v	Node at which the 1st female	st female	Number	Number of days to 1st female	1 <sup>st</sup> female	
Treatment	flo	flower appeared	red		flower		flo	flower appeared	ed		flower		
	2015	2016	Mean	2015	2016	Mean	2015	2016	Mean	2015	2016	Mean	
$\mathrm{T}_{0}$	25.3a	21.3a	23.3	67.0a	69.7a	68.3	28.3a	27.0a	27.7	78.3a	81.7a	80.0	
$\mathrm{T}_{\mathrm{l}}$	23.0b	19.3ab	21.2	62.3b	65.7bc	64.0	26.3b	24.3ab	25.3	75.7b	78.7b	77.2	
$\mathrm{T}_2$	22.3c	17.0bc	19.7	60.3cd	63.3cd	61.8	23.0de	21.0ab	22.0	71.00cd	74.7d	72.8	
$\mathrm{T}_3$	21.3cd	18.0bc	19.7	61.0bc	66.7b	63.8	25.3bc	23.0ab	23.2	76.0b	79.00b	77.5	
$\mathrm{T}_4$	21.0cd	18.0bc	19.5	61.3bc	67.3ab	64.3	24.3cd	23.0ab	23.7	76.3b	79.3b	77.8	
$T_5$	20.3d	17.0bc	18.7	60.3cd	65.7bc	63.0	22.3ef	20.3b	21.3	72.0cd	76.7c	74.3	
$T_6$	18.0e	16.3c	17.2	59.0e	63.0d	61.0	21.0f	20.0b	20.5	70.0d	74.3d	72.2	
$\mathrm{T}_7$	21.0cd	17.0bc	19.0	60.0cd	65.0bcd	62.5	22.0ef	21.3ab	21.7	72.7c	76.7c	74.7	
CV (%)	4.87	5.35	1	4.30	2.26	1	4.02	3.96		4.12	3.76		
Table 3. cont'd.	nt'd.												
E		M	Male flower (no.)	. (no.)		Female flower (no.)	ower (no.)		Sex ratio	Sex ratio (male : female)	imale)		
I reaument		2015	2016	16	Mean	2015	2016	Mean	2015		2016	Mean	
$\mathrm{T}_{0}$	1	123.0 b	131	131.3a	127.2	15.7 c	18.3d	17.0	7.9 a		2a	7.5	
$T_1$	1	103.7 c	115.	l15.3bc	109.5	24.0 b	27.3bc	25.7	4.3 bc		4.3b	4.3	
$\mathrm{T}_2$	1	103.2 c	118	118.3b	107.8	23.0 b	34.0ab	28.5	4.5 bc		3.5bc	4.0	
$T_3$	x	82.0 de	98.3de	3de	90.2	25.3 b	28.3bc	26.8	3.2 c		3.4bc	3.4	
$\mathrm{T}_4$		94.7 d	104	104.0e	99.3	23.3 b	25.0cd	24.2	4.1bc		4.2b	4.1	I
$T_5$		93.0 cde	107.3cd	3cd	100.2	24.3 b	29.0bc	26.7	$3.8 \ bc$		8b	3.8	10
$T_6$		81.7e	96.0e	0e	88.8	37.3 a	40.0a	38.7	2.2 c		2.5d	2.3	INI
$T_7$	1	143.3 a	120	120.3b	131.8	36.3 a	38.0a	37.2	$3.9 \ bc$		3.2cd	3.6	ΧUΖ
CV(%)		4.18	4.87	87	ı	5.74	8.26	I	9.14	11.	11.14	·	LLP
Means shov	ving dissin	milar letter	s in a col	umn are si	gnificantly	different at	Means showing dissimilar letters in a column are significantly different at 5% level by Tukey's W test.	by Tukey's	W test.				AIVI A
$T_0 =$ Control (distilled water),	ol (distille		$\Gamma_1=GA_3$	@ 10 ppm,	, $T_2 = GA_3$	@ 30 ppm,	, $T_3 = NAA$	@ 100 pp	m, $T_4 = N$	AA @ 150	ppm, T <sub>5</sub> :	$T_{1} = GA_{3} @ 10 ppm, \ T_{2} = GA_{3} @ 30 ppm, \ T_{3} = NAA @ 100 ppm, \ T_{4} = NAA @ 150 ppm, \ T_{5} = MH @ 50 ppm, \ T_{5} = MH \ T_{5} = MH$	4IN (
ppm, $T_6 = MH \otimes 150 ppm, T$	AH @ 15(		= CCC @	= CCC @ 500 ppm									zi u

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E		Fruit	Fruits/plant (no.)		Fruits/plant (no.) Fruit length (cm) Fruit	Fruit length (cm)		Fruit	Fruit circumference (cm)	e (cm)
I reatment		2015	2016	Mean	2015	2016	Mean	2015	2016	Mean
$\mathrm{T}_{0}$	7	4.7 c	6.7f	5.7	46.7	47.3	47.0	37.9	37.6	37.8
$T_1$	•	7.7 b	9.7cde	8.7	47.5	47.7	47.6	38.3	39.9	39.1
$\mathrm{T}_2$		8.7 b	12.0ab	10.4	49.6	49.5	49.5	39.8	40.2	40.0
$T_3$	9	6.7 bc	7.7ef	7.2	48.9	49.2	49.1	39.1	39.9	39.5
$\mathrm{T}_4$		7.0 b	9.0de	8.0	47.7	47.7	47.7	41.3	40.2	40.8
$T_5$		8.0 b	10.7bcd	9.3	46.8	47.9	47.4	38.1	40.3	39.2
${ m T_6}$	1	12.0 a	14.0a	13.0	47.2	48.1	47.7	38.7	39.6	39.2
$\mathrm{T}_7$	1	11.3 a	11.7bc	11.5	47.2	48.2	47.7	35.8	38.6	27.2
CV (%)		9.76	7.87		5.61	6.56		10.59	8.13	
										Mean yield
1	:			ţ					,	Mean yield
Treat-	Individ	Individual fruit we	weight (kg)	Fru	Fruit weight/plant (kg)	t (kg)		Fruit yield (t/ha)	ha)	increase over
ment										control (%)
	2015	2016	Mean	2015	2016	Mean	2015	2016	Mean	
$\mathrm{T}_{\mathrm{0}}$	2.4	2.5	2.5	11.5e	16.1e	13.8	38.5 e			ı
$\mathbf{T}_{\mathbf{I}}$	2.4	2.5	2.4	18.3cd	20.8d	19.6	61.1 cd			30.2
$\mathrm{T}_2$	2.7	2.6	2.6	21.8bc	30.1b	26.0	72.8 bc			46.7
$T_3$	2.4	2.6	2.5	16.2de	20.1d	18.2	53.5 de			24.1
$\mathrm{T}_4$	2.8	2.6	2.7	18.6cd	22.8cd	20.7	62.0 cd			33.7
$T_5$	2.7	2.6	2.6	21.1c	26.5bc	23.8	70.4 c		68.3	42.3
$\mathrm{T}_{6}$	2.6	2.7	2.6	29.3a	35.8a	32.6	97.6 a			54.2
$\mathbf{T}_7$	2.5	2.6	2.6	26.6ab	29.1b	27.8	88.5 ab			51.1
CV (%) 6.	6.18	10.34	I	8.33	6.13		8.27	6.10	8.59	·
eans sho	Means showing dissimilar let	milar letters	s in a column	are signific:	ters in a column are significantly different at 5% level by Tukey's W test	at 5% level b	y Tukey's V	W test.		
= Cont	$T_0 =$ Control (distilled water)	d water), T	$1 = GA_3 @ 10$	0 ppm, $T_2 =$	GA <sub>3</sub> @ 30 pp	$m, T_3 = NAA$	A @ 100 ppr	m, $T_4 = NAA$	@ 150 ppm, 7	), $T_1 = GA_3 @ 10 \text{ ppm}, T_2 = GA_3 @ 30 \text{ ppm}, T_3 = NAA @ 100 \text{ ppm}, T_4 = NAA @ 150 \text{ ppm}, T_5 = MH @ 50 \text{ ppm}, T_5 = MH $
$m, T_6 =$	ppm, $T_6 = MH \otimes 150 ppm, T$	0 ppm, $T_7 =$	$\gamma = \text{CCC} @ 500 \text{ ppm}$	) ppm.						

INFLUENCE OF PLANT GROWTH REGULATORS ON VEGETATIVE

## Sex ratio of male and female flower

All the treatments resulted in narrowing down of male-female sex ratio over control in both years (Table 3). The lowest sex ratio (male:female) was recorded at MH @150 ppm in both the years (2.2 in 2015 and 2.5 in 2016; mean 2.30). In 2015, all the treatments except control produced identical values of sex ratio, but in 2016, MH @ 150 and CCC @ 500 were identical with regard to sex ratio. Again, in 2016, there was no significant difference among GA<sub>3</sub> @10 (T<sub>1</sub>), GA<sub>3</sub> @ 30 (T<sub>2</sub>), NAA @100 (T<sub>3</sub>), NAA @ 150 (T<sub>4</sub>) and MH @ 150 (T<sub>5</sub>) ppm in respect of sex ratio. Although CCC @ 500 ppm produced the maximum male flowers compared to control in 2015, it gave lower sex ratio (3.9) than control (7.9) because of producing higher female flowers.

The marked suppression of male flowers and induction of female flowers by all concentrations of GA<sub>3</sub>, NAA, MH and CCC @ 500 ppm was noticed. Induction of female flowers by all concentrations of PGRs was the evidence of lowered sex ratio. The sex ratio recorded in this investigation was comparable to the results reported by Hidayatullah *et al.* (2012) with GA<sub>3</sub>, Sharma *et al.* (1988) as well as Pandya and Dixit (1997) with CCC and Ingle *et al.* (2000) as well as Arora *et al.* (1982) with MH in bottle gourd.

# Yield attributes and yield of bottle gourd

Growth regulator treatments had significant effect on yield components and yield of bottle gourd except fruit length, fruit circumference and individual fruit weigh in both the years (Table 4). The maximum number of fruits/plant was found in MH @ 150 ppm in both the years (12.0 in 2015 and 14.0 in 2016; mean 13.0). In the first year, MH @ 150 ppm gave identical fruit number with CCC @ 500 ppm (11.3), but in the 2<sup>nd</sup> year, number of fruits produced by the plants treated with MH 150 ppm was statistically similar to those of  $GA_3 @ 30 ppm (12.0)$ . The lowest number of fruits was obtained from control in both the years (4.7 in 2015 and 6.7 in 2016; mean 5.7). The maximum fruit weight/plant was recorded at MH @ 150 ppm in both the years (29.3 kg and 35.8 kg in 2015 and 2016, respectively). In 2015, MH @ 150 ppm was identical with CCC @ 500 ppm in producing fruit weight/plant. In the  $2^{nd}$  year, the second highest fruit weight/plant was obtained from the application of GA<sub>3</sub> @ 30 ppm (30.1 kg) which was statistically similar to CCC @ 500 ppm (29.1 kg). The mean values over two years indicated the highest fruit weight/plant (32.6 kg) was recorded from MH @ 150 ppm followed by CCC @ 500 ppm (27.8 kg) and GA<sub>3</sub> @ 30 ppm (26.0 kg). The lowest fruit weight/plant was found from the control (11.5 kg in 2015 and 16.1 kg in 2016; mean 13.8 kg) in both the years.

Fruit yield per hectare followed the same trend of fruit weight/plant. MH 150 ppm gave the highest fruit yield (97.6 t/ha in 2016, 89.6 t/ha in 2016 and mean 93.6 t/ha). In 2015, CCC @ 500 produced the identical yield (88.5 t/ha) of MH @ 150 ppm. In 2016, application of GA<sub>3</sub> @ 30 ppm gave the  $2^{nd}$  highest yield

(74.1 t/ha) which was statistically similar to CCC @ 500 ppm (72.7 t/ha). It was observed from the mean values that the maximum fruit yield (93.6 t/ha) was recorded from MH @ 150 ppm followed by CCC @ 500 ppm (80.6 t/ha) and GA<sub>3</sub> @ 30 ppm (74.1 t/ha). The control treatment gave the lowest yield (38.5 t/ha and 40.4 t/ha in 2015 and 2016, respectively; mean 39.4) (Table 4). MH (50-150 ppm) produced yields in the range of 70.4 -97.6 t/ha in 2015 and 66.2-89.6 t/ha in 2016. Higher concentration of MH (150 ppm) produced higher yield than MH @ 50 ppm in both the years. On the other hand, GA<sub>3</sub> (10-30 ppm) gave the yields ranged from 61.1 -72.8 t/ha in 15 and 52.0-75.3 t/ha in 2016 and GA<sub>3</sub> @ 30 ppm produced the highest yield in both the years. NAA (100-150 ppm) produced fruit yield ranged from 53.5-62.0 t/ha in the first year and from 50.4-56.9 t/ha in the 2<sup>nd</sup> year and NAA @ 150 ppm gave the highest fruit yield in both the years. In mean values over two years, plants of bottle gourd treated with 150 pm MH, 500 ppm CCC and 30 ppm GA<sub>3</sub> gave 54.16%, 51.06% and 46.74 % higher yield, respectively than control. An increase in yield could be attributed to earliness and increased number of female flowers as well as narrowed male:female sex ratio. Arora et al. (1982) reported that MH @ 50 ppm was most effective in producing the maximum fruit weight/plant and ultimately the yield. Foliar spray of MH (50-150 ppm) increased the yield in most of the cucurbits (Sonkar, 2003; Jatoi et al., 2010). Plants sprayed with MH @ 50 ppm at 2-leaf stage produced the best yield (Baruah and Das, 1997). Saimbhi and Thakur (2006) reported that single spray of CCC @ 500 ppm increased number of fruits per plant and yield in squash melon (Citrullus vulgaris). Sadiq et al. (1990) obtained maximum number of fruits/plant and yield/vine from application of CCC 5@ 00 in cucumber. Hidayatullah et al. (2012) obtained maximum fruit weight/plant and yield in bottle gourd from the spraying of plants with GA<sub>3</sub> 30 ppm. Desai et al. (2011) obtained the maximum yield of bottle gourd from spraying with paclobutrazol @ 25 ppm followed by CCC @ 200 ppm, CCC @ 500 ppm and MH @ 200 ppm.

#### **Partial Economic Analysis**

The present study (Table 5) revealed that the maximum gross return (Tk. 11,70,000.00) was found from the plants sprayed with MH @ 150 ppm followed by CCC @ 500 ppm (Tk. 10,07,380.00) and GA<sub>3</sub> @ 30 ppm (Tk. 9,25,750.00) and the minimum gross return was recorded from control (Tk. 4,93,000.00). Net return showed marked difference among the treatments and followed the same trend of gross return. Net return was the highest (Tk. 9,82,600.00) in MH @ 150 ppm followed by CCC @ 500 ppm (Tk. 8,15,580.00) and GA<sub>3</sub> @ 30 ppm (Tk. 7,35,200.00) while the lowest (Tk. 3,06,200.00) in control. The maximum benefit cost ratio (BCR) was obtained from MH @150 ppm (5.24) followed by CCC @ 500 ppm (4.25) and GA<sub>3</sub> @ 30 ppm (3.86); while the minimum from control (1.64). The cost and return analysis revealed that spraying of MH @ 150 ppm was superior to CCC @ 500 and GA<sub>3</sub> @ 30 ppm in terms of net income and BCR.

Treatment	Mean fruit yield (t/ha)	Gross return ('000 Tk./ha)	Cost of treatment ('000 Tk./ha)	Cost of cultivation ('000 Tk./ha)	Net return ('000 Tk./ha)	Benefit- cost ratio (BCR)
$T_0$	39.44	493.00	0.00	186.80	306.20	1.64
$T_1$	56.54	706.75	1.25	188.05	518.70	2.76
$T_2$	74.06	925.75	3.75	190.55	735.20	3.86
$T_3$	51.98	649.75	0.55	187.35	462.40	2.47
$T_4$	59.44	743.00	0.83	187.63	555.38	2.96
$T_5$	68.31	853.87	0.20	187.00	666.87	3.57
$T_6$	93.60	1170.00	0.60	187.40	982.60	5.24
<b>T</b> <sub>7</sub>	80.59	1007.38	5.00	191.80	815.58	4.25

 Table 5. Partial cost benefit analysis of summer bottle gourd production by using plant growth regulators

 $T_0$  = Control (distilled water),  $T_1$  = GA<sub>3</sub> @ 10 ppm,  $T_2$  = GA<sub>3</sub> @ 30 ppm,  $T_3$  = NAA @ 100 ppm,  $T_4$  = NAA @ 150 ppm,  $T_5$  = MH @ 50 ppm,  $T_6$  = MH @ 150 ppm,  $T_7$  = CCC @ 500 ppm.

Basic cost of cultivation: 186800 Tk./ha.

Cost of PGRs:

- 1. Gibberellic acid (GA<sub>3</sub>): Tk. 500.00/g
- 2. Naphthalelene Acetic Acid (NAA): Tk. 2200.00/100 g
- 3. Maleic hydrazide : Tk. 1500.00/100g
- 4. Cycocel (CCC) : Tk. 2000.00/100ml (50% ai)

Treatment	Dose/ha
GA <sub>3</sub> @ 10 ppm (T <sub>1</sub> )	2.5 g
GA <sub>3</sub> @ 30 ppm (T <sub>2</sub> )	7.5 g
NAA @ 100 ppm (T <sub>3</sub> )	25.0 g
NAA @ 150 ppm (T <sub>4</sub> )	37.5 g
MH @ 50 ppm (T <sub>5</sub> )	13.33 g
MH @ 150 ppm (T <sub>6</sub> )	40.0 g
CCC @ 500 ppm (T <sub>7</sub> )	500 ml

Market selling price of bottle gourd fruits: Tk. 12.50/kg (Tk 12500.00/ton)

# Conclusion

The experimental result revealed that PGR played a significant role in regulating number of branches/plant, male and female flowering, narrowing sex ratio and ultimately increasing number of fruits and yield. Treatments of MH @ 150 ppm, CCC @ 500 ppm and  $GA_3$  @ 30 ppm were superior to the rest of the treatments for the earliness of female flower appearance, narrowing down sex-ratio and

finally increasing yield. These three treatments also registered higher gross return, net return and BCR. Therefore, it can be concluded that spraying MH @150 ppm or CCC @ 500 ppm or  $GA_3$  @ 30 ppm to the seedlings of bottle gourd at two-leaf stage and  $2^{nd}$  spraying after four days of  $1^{st}$  spray should be done in summer to get higher yield with higher return.

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